

Assessment of acetylcholinesterase inhibition by *Bacopa monnieri* and acephate in hippocampus of chick brain for impediment of Alzheimer's disease

Abstract

Introduction: In Alzheimer's disease (AD), a depletion of the neurotransmitter, acetylcholine (ACh) occurs due to the destruction of cholinergic neurons. By inhibiting Acetylcholinesterase (AChE), the enzyme which catalyses break down of ACh, levels of this neurotransmitter can be elevated and function may be improved.

Objective: This study sought to find out whether the Acetylcholinesterase (AChE) activity, enzyme kinetics and histology in the hippocampus of Chick brain were affected in a selective manner by a herbal extract, and organophosphate Pesticide and combination of both:

- Ethanol extract of *Bacopa monnieri*
- Acephate
- Pretreatment of ethanol extract of *Bacopa* followed by acephate.

Method: AChE activity was determined by Ellman's method and AChE kinetics by LB plots.

Result: AChE activities due to *Bacopa monnieri*, acephate and combination of both in hippocampus of treated chicks were significantly inhibited to 72%, 70.9% and 94% respectively. Our kinetic study reveals that acephate is competitive while *Bacopa monnieri* was competitive-noncompetitive inhibitor of AChE. Slight histological variation was observed as the concentration of cytoplasm of CA1 and CA3 pyramidal cell layers in the Hippocampus of chick treated with acephate and ethanolic extract of *Bacopa monnieri*.

Conclusion: Common herb *Bacopa monnieri* may help to improve ACh level, but scientific evidence to prove that it can treat Alzheimer's disease, is still to confirm.

Keywords: hippocampus, chick brain, acephate, inhibition, kinetics, K_m , V_{max} , *Bacopa Monnieri*, acephate

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Abbreviations: IAEC, institutional animal ethics committee; AD, Alzheimer's disease; AChE, acetylcholinesterase; DTNB, dithionitrobenzoic acid; OD, optical density

Introduction

World Alzheimer Report of 2015¹ has estimated that 46.8million people are currently living with dementia worldwide which will be further increased to 74.7million by 2030 and 131.5million by 2050. Alzheimer's disease (AD) is a neuro-degenerative disorder and common form of dementia, which progress gradually with the age. The common symptom of AD is memory impairment and cognitive shortfall due to low levels of acetylcholine in the brain of AD patients. According to the cholinergic theory, the inhibition of acetylcholinesterase (AChE), an enzyme that catalyzes acetylcholine hydrolysis, increases the levels of acetylcholine in the brain, thus improving cholinergic functions in AD patients.² At present tacrine, donepezil, rivastigmine and galanthamine are few approved drugs for the symptomatic treatment of Alzheimer's disease. The clinical response of these drugs has limited effectiveness and has some kind of side effect in AD patients. These side effects include Dizziness

and syncope, Bradycardia, atrial arrhythmias, myocardial infarction, angina, seizures Sino-atrial and atrioventricular block.³ Galanthamine is a natural alkaloid first obtained from *Galanthus* spp. was approved by US-FDA in 2001. Recently Huperzine A, an alkaloid derived from *Huperzia* spp., has been recommended as commercialized AChEi as a dietary supplement to treat AD symptoms in China, which gives promising results in memory improvement.⁴ Screening of 23 pure Amaryllidaceae alkaloids and 26 extracts from different species of the genus *Narcissus* have shown positive results for their Acetylcholinesterase inhibitory activity using galanthamine as a reference.⁵ Herbal remedies for Alzheimer's disease have become more and more popular in the recent years and not without a reason that there is a possibility to slow down the brain's degeneration caused by Alzheimer's with natural treatments and it has drawn the attention of the scientific community.⁶ Thus, there is need to develop targeted effective therapeutics for the treatment of Alzheimer's disease with least or no side effects. OP pesticides act by their ability to inhibit AChE in vertebrates and invertebrates.⁷⁻⁹ Kinetic analysis of different species: mammalian (monkey, rat, and guinea pig), avian (chicken), piscine (catfish), and amphibian (frog) in brain Acetylcholinesterase have shown differential sensitivity to organophosphate insecticides.¹⁰⁻¹²

Bacopa monnieri is a perennial, creeping herb and commonly used nervine tonic and memory enhance herb. It has been used in the Ayurvedic system of medicine for centuries. Acephate is widely used organophosphate pesticide. Acephate kills insects by disrupting their nervous function. This paper reviews the AChE inhibitory effects of a commonly used medicinal herb *Bacopa monnieri* in hippocampus of brain of chick for the treatment of Alzheimer's disease and dementia and it was compared with acephate, which is potent AChE OP inhibitor. Animal studies have suggested that early stress is associated with alterations in the hippocampus, a brain area that plays a crucial role in learning and memory. Therefore, purpose of this study was to measure both AChE inhibitions, AChE inhibitory kinetics and alterations in hippocampus structure in chick brain due to herb *Bacopa monnieri* and acephate.

Materials and methods

Preparation of plant extract

Fresh *Bacopa monnieri* were procured from Jawaharlal Nehru Agricultural University, Jabalpur (M.P.) India. Plant material was validated by botanist. Leaves and twigs were cut and carefully washed in water; dried in shade. Dry plant material was ground to powder. Ethanol extract of *Bacopa monnieri* was prepared in Soxhlet apparatus using 90% alcohol at 75-80°C. Filtrate was dried to semisolid mass, freeze-dried and re-constituted in de-ionized water prior to assay.

Animals and treatment

20 chicks of 100-125g were divided into 4 groups and each group consisted of a minimum of five animals. Separate animals were used for each experiment. They were housed separately in polypropylene cages, maintained in laboratory condition at 30°C with 12/12hours light and dark cycle for 10days. They were allowed free access to commercial feed and water *ad libitum*. Ethical clearance for the use of animals was obtained from the Institutional Animal Ethics Committee (IAEC). The animals were orally dosed via feeding needle as: Group-I: Ground nut oil 1ml/kg b.w. for 4days, served as control (vehicle); Group-II: 37.8mg/kg b.w. acephate (1/25th of LD50) in ground nut oil daily for 4days; Group-III : Ethanol extract of *Bacopa monnieri* 100mg/kg in deionized water for 7days; Group-IV: Pretreatment of ethanol extract of *Bacopa monnieri* (100mg/1kg b.w.) for 7days daily followed by 37.8mg/kg b.w. acephate for 4days.

Tissue preparation

At the end of experiment, animals were euthanized and brains were dissected out according to Zeman and Innes.¹³ Hippocampus from each chick was quickly isolated, weighed and thoroughly washed with isotonic saline. Tissues were homogenized in buffer (7.4 p^H) followed by centrifugation at 4°C and supernatant was used as source of enzyme.

Estimation of AChE activity

The hippocampus AChE activity was measured using the Ellman's method.¹⁴ The end point was the formation of the yellow color because of the reaction of thiocholine with di-thio-bis-nitrobenzoate ions. The rate of formation of thiocholine from acetylcholine iodide in the presence of tissue cholinesterase was measured using a SL164 UV-VIS spectrophotometer. The sample was first treated with 5,50-dithionitrobenzoic acid (DTNB), and the optical density (OD) of the yellow color compound formed during the reaction at 412nm every minute for a period of 2min was measured. All measurements were done in duplicate. Specific activity was expressed in $\mu\text{mol}/\text{min}/\text{mg}$ protein.

Protein determination

Protein was determined by the Lowry method¹⁵ using BSA as the standard. Aliquots of homogenate were diluted with reagents then 0.5ml Folin's reagent was added and kept for incubation for 20min then the colour was read at 620nm against a reagent blank. Measurements were done in duplicate.

AChE kinetics

Kinetic parameters like K_m and V_{max} were calculated by Line weaver-Burk plots, which were drawn from assays using acephate (37.8mg/kg) and *Bacopa monnieri* extract (100mg/kg) at various substrate concentrations (0.66mM, 0.44mM, 0.33mM, 0.26mM ATCI). From these Kinetic parameters (K_m & V_{max}) were determined for each assay by plotting the reciprocals of velocity and substrate concentrations.

Histological examination

Tissues from each group of chicks were fixed in aqueous Bouin's fluid, dehydrated and embedded in paraffin wax. Serial sections were cut at 7 micron and stained with Haematoxylin and Eosin. The microphotographs of serial sections were taken at 400X with computer-aided microscope (Leica).

Statistical analysis

For the data of statistical comparison between different treatments and control groups, data were analyzed by Student's t-test to determine the effect of the treatment. The level for the accepted statistical significance was $P > 0.05$.

Results and discussion

A few studies have been undertaken to evaluate AChE inhibitory potential herbs and pesticides in different discrete regions of Brain of chick.¹⁶⁻¹⁹ Present investigation is aimed at the assessment of in vivo AChE inhibition, its kinetics and histological alterations in hippocampus of brain of chick subjected to intoxication with Acephate, treatment with *Bacopa monnieri* singly and in combination with acephate. Our result showed that AChE specific activity in hippocampus of control chick was $10.58 \pm 8.4 \times 10^{-6}$ M. However, daily oral dose of acephate (34mg/kg b. w.) for fourdays to chick produced 70.9% significant AChE inhibition (Table 1). Organophosphorus insecticides Chlorpyrifos and methyl parathion were potent inhibitor of Acetylcholinesterase in hippocampus.²⁰ Hippocampus has been reported to sensitive to AChE inhibition by multiple exposures to 2-organophosphate pesticides i.e. Tri-Ortho-Toly phosphate and Chlorpyrifos²¹ and Chlorpyrifos and methyl parathion.²⁰

Oral dose of 100mg/kg b. w ethanol extract of *Bacopa monnieri* yields 72 % AChE inhibition in hippocampus (Table 1). However, AChE inhibition increased to many fold i.e. 94% after 7days pretreatment of of *Bocopa monnieri* ethanolic extracts (100mg/kg b. w.) followed by acephate (34mg/kg b.w.) daily for 4days. The hydroalcohol extract from *Centella asiatica*, *Nardostachys jatamansi*, *Myristica fragrans*, *Evalvulus alsinoides* inhibited 50% of AChE activity.²² The Chinese herb *Huperzia serrata* has been shown to possess antioxidant and neuroprotective properties suggesting thereby its potential in the treatment of Alzheimer's disease.⁴ Huperzine A is a medicinally active plant derivative found in *Huperzia serrata*. This drug has been shown to inhibit acetylcholinesterase (such as tacrine and donepezil) and to improve memory and mental functioning in patients with Alzheimer's and other severe conditions.²³ Three herbal plants, *Melissa officinalis*, *Salvia officinalis* & *Rosmarinus officinalis*

have proved their efficacy in the management of patients with AD as they are active not only in inhibition of AChE or β -amyloid deposits inhibition *in-vitro*.^{24,25} Current treatments option available for dementia are very limited (cholinesterase inhibitors, NMDA receptor antagonist)²⁶ they only provide short term symptomatic relief and do not prevent disease progression. Also, over the last decade a large number of drug candidates have failed in clinical trials.²⁷

The primary mechanism of action of organophosphate pesticides and anticholinesterase herbs includes the inhibition of AChE kinetics.^{10,28,29} In the present study the inhibitory Kinetics of AChE was determined by applying Lineweaver-Burk plot. The kinetic parameters Michaelis Menten Constant (Km) and maximum velocity (V_{max}) were calculated in control and treated groups to compare inhibitory kinetic in hippocampus of chick. The km was determined by slops for the control and acephate inhibited AChE, intersecting separately on the abscissa represent the Km value and the slops intersecting on the ordinate indicate V_{max} values. Data of our kinetic study also substantiate AChE inhibition in hippocampus of chick. The km value in hippocampus of control group was 0.25 ± 0.15 mM, which significantly increased with treatment of acephate (0.83 ± 0.35 mM) and *Bacopa monneiri* (0.37 ± 0.25 mM). The Km also elevated to 0.44 ± 0.26 in the mixed treatment group IV (Figure 1) (Table 1). It was interesting to note that acephate and *Bacopa monneiri* display different nature of AChE inhibition in hippocampus. This was apparent as the V_{max} values were decreased while Km increased in chicks after treatment with *Bacopa monneiri* singly and in combination with acephate as compared to untreated groups of chick. Thus, *Bacopa monneiri* yields mixed AChE inhibition i.e. competitive-non competitive (Figure 1) (Table

1). Such type of mixed inhibition kinetics is typical of some medicinal plants; due to the great variety of compounds they contain all acting in different ways.³⁰ Contrary to this, acephate produced competitive AChE inhibition indicating increasing Km in treated chicks and constant V_{max} in both control and acephate treated chicks (Figure 1) (Table 1). Abdulaziz Al-Jafari^{31,32} reported Cyclophosphamide reversibly inhibits AChE activity of Chicken brain.

In our study a few histological variations was observed in the Hippocampus showing concentration of cytoplasm of CA1 and CA3 layers in chick treated with acephate and combined treatment group. No significant changes were observed in hippocampus structure in chick treated with *Bacopa monneiri*. (Figure 2). It has been reported that *Bacopa monneiri* in rat hippocampus significantly increased the density of cholinergic neurons in hippocampus.³² Sarin and soman affected adversely pyramidal cells of C_{A1} and C_{A3} in the hippocampus.³³ Similar study on the toxicity of alpha-cypermethrin at the dose of 250mg/kg b. w. for four weeks showed focal concentration in C_{A3} layer of hippocampus.³⁴ Other investigation on Chlorpyrifos and Cypermethrin intoxication resulted in increased density of the cytoplasm in neurocytes of hippocampus.³⁵ Shih et al.³⁶ opined that critical histological examination of brain sections is essential in determining the nature and extent of such changes to understand relationship of the severity of cellular injury and degree and duration of toxicant-induced convulsions. Our findings clearly show that *Bacopa monneiri* is potent inhibitor AChE as OP pesticide acephate does. Kinetic study confirms its mixed nature of inhibition. It does not produce any adverse effect on histology of hippocampus.

Table 1 Acetylcholinesterase activity (Activity/mg protein/min), % inhibition, Km $\times 10^{-3}$ and Vmax of AChE of Hippocampus of Chick brain treated with sub lethal dose of Acephate (34mg/kg b. w.), *Bacopa monneiri* (100mg/kg b. w.) and pretreatment of *Bacopa monneiri* (100mg/kg b. w.) Followed by Acephate (34mg/kg b.w.). The AChE specific activity is expressed in μ moles of ATCl hydrolyzed/mg protein/min

Parameters	Control	Acephate	<i>Bacopa Monneiri</i>	<i>Bacopa monneiri</i> and Acephate
AChE specific activity	10.58 \pm 8.2	3.49 \pm 2.1***	2.93 \pm 0.07	0.619 \pm 0.203*
AChE inhibition%	-	70.9	72	94
Km $\times 10^{-3}$ M	0.25 \pm 0.15	0.83 \pm 0.35*+232%	0.37 \pm 0.25***+48%	0.44 \pm 0.26**+76%
Vmax (A/min/mg protein)	0.12	0.12	0.11	0.17

Values are expressed as mean \pm S.D of 5 individual animals in each group analyzed by student's t-test P<0.01*; P<0.02**; P<0.05***, compared with control vs. treated

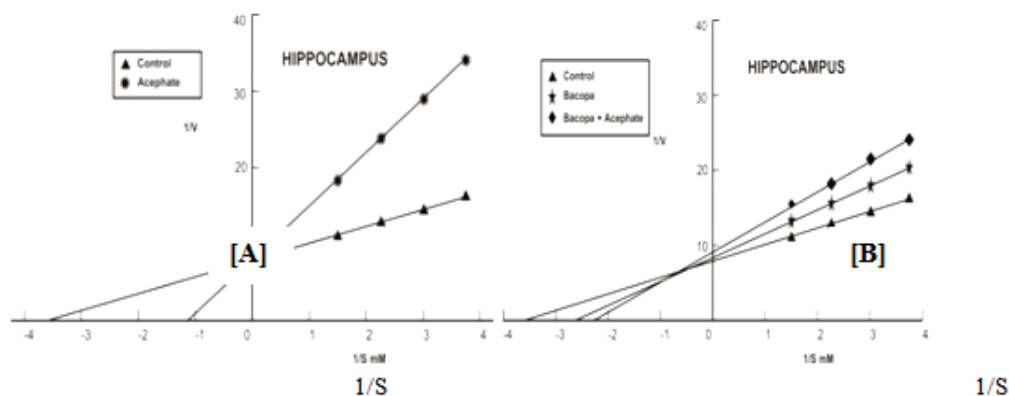


Figure 1 Lineweaver–Burk plot of in vivo inhibition of AChE by [A]- Acephate (34mg/kg b. w.) and [B]- *Bacopa monneiri* (100mg/kg b. w.) and pretreatment of *Bacopa monneiri* (100mg/kg b. w. for 7days) followed by treatment of Acephate (34mg/kg b. w. for 4days) in hippocampus of chick brain. S is the concentration of ATCl. Each point is mean of five assays.

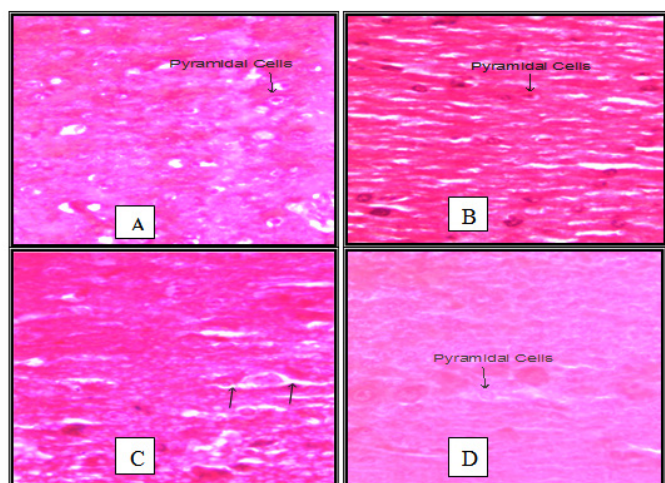


Figure 2 Photomicrograph of T.S. of hippocampus of Chick brain. H.E.X400: [A], Group I (Control) showing the normal structure. [B], Group II (Exposed to acephate 34mg/kg b.w.) showing concentration of cytoplasm of CAI layer. [C], Group III (Exposed to *Bacopa monnieri* 100mg/kg b.w.) showing no significant changes. [D], Group IV (Exposed to pretreatment of *Bacopa monnieri* 100mg/kg b.w. followed by acephate 34mg/kg b.w.) showing concentration of cytoplasm of CAI layer.

Conclusion

Since, AChE inhibition plays a critical role in slowing progression of AD; it seems that *Bacopa monnieri* could act as anti-Alzheimer with at least three mechanisms: AChE inhibition, antioxidant and anti-inflammatory activity. It is a potential cognitive enhancer and neuroprotectant against Alzheimer's disease.^{32,37} This approach has a particular success as these cholinergic neurons are found mainly in regions associated with learning and memory - spreading from the basal forebrain³⁸ and hippocampus³⁹ up to the cerebral cortex.⁴⁰ In order to find the mechanisms more investigations are necessary. Further works on *Bacopa monnieri* could lead to isolate bioactive compounds helpful in AD. The paucity of treatment is due to the multi-factorial nature of AD associated cognitive dysfunction. There is a need to develop combination therapies, however adverse effects associated with modern synthetic drugs could prove to be prohibitive when trying to develop them as combination treatments.

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None.

Conflict of interest

Author declares that there is no conflict of interest.

References

- Gemma CA, Maëllenn G, Yu-Tzu Wu, et al. The global prevalence of dementia. In *Alzheimer's disease International: World Alzheimer Report*. London: Published by Alzheimer's disease International (ADI); 2015:10–29.
- Murray AP, Faraoni MB, Castro MJ, et al. Natural AChE Inhibitors from Plants and their Contribution to Alzheimer's Disease Therapy. *Curr Neuroparmacology*. 2013;11(4):388–413.
- Rowland JP, Rigby J, Harper AC, et al. Cardiovascular monitoring with acetylcholinesterase inhibitors: a clinical protocol. *Advances in Psychiatric Treatment*. 2007;13(3):178–184.

- Kozikowski, Alan P, Tueckmantel, et al. Chemistry, Pharmacology, and Clinical Efficacy of the Chinese Nootropic Agent Huperzine A. *Acc Chem Res*. 1999;32(8):641–650.
- López S, Bastida J, Viladomat F, et al. Acetylcholinesterase inhibitory activity of some Amaryllidaceae alkaloids and Narcissus extracts. *Life Sci*. 2002;71(21):2521–2529.
- Hajimehdipoo H, Tehranifar T, Shafaroodi H. Acetylcholinesterase Inhibitory Effect of Some Medicinal Herbs Used in Iranian Traditional Medicine for Memory Improvement. *Global Journal of Botanical Science*. 2013;1(1):18–21.
- Moser VC, McDaniel KL, Phillips PM, et al. Time-Course, Dose-Response, and Age Comparative Sensitivity of N-Methyl Carbamates in Rats. *Toxicol Sci*. 2009;114(1):113–123.
- Satyadevan S, Kumar S, Tembhre M. Acetylcholinesterase activity and enzyme kinetics in brain of common carp, *Cyprinus carpio* subjected to sub lethal exposure to dimethoate. *Biosc Biotech Biochem*. 1993;57(9):1566–1567.
- Tembhre M, Ahirwar S, Gour S, et al. Inhibitory Potential of Acephate and Ethanol Extract of *Bacopa Monnieri* on AChE in Rat Cortex and Hippocampus. *IJBBB*. 2015;5(1):45–53.
- Wang C, Murphy SD. Kinetic analysis of species difference in acetylcholinesterase sensitivity to organophosphate insecticides. *Toxicol Appl Pharmacol*. 1982;66(3):409–419.
- Tembhre M, Ahirwar S, Gour S. Chlorpyrifos induced inhibition of AChE in *Cyprinus carpio* and recovery during drug leaching. *J of Cell and Tissue Res*. 2006;6(2):0793–0028.
- Ahirwar S, Tembhre M, Gour S, et al. Anticholinesterase Efficacy of *Bacopa monnieri* against the Brain Regions of Rat - A novel approach to therapy for Alzheimer's disease. *Asian J Exp Sci*. 2012;26(1):65–70.
- Zeman W, Innes JRM, Horne EC. *Craigie's Neuroanatomy of the Rat*. New York: Academic Press; 1963:21–25.
- Ellman G, Courtney D, Andres V. A new and rapid colorimetric determination of acetylcholinesterase activity. *Biochem Pharmacol*. 1961;7:88–95.
- Lowry OH, Rosebrough NJ, Farr AL, et al. Protein measurement with the Folin phenol reagent. *J Biol Chem*. 1951;193(1):265–275.
- Tembhre M, Gour M, Ahirwar S, et al. Diacetylmorphine reactivation of Acetylcholinesterase Butyrylcholinesterase Inhibited by Dichlorvos in Central and Peripheral Nervous System of Rat. *IPCBBE*. 2012;48(24):126–133.
- Suganthi N, Pandian SK, Devi KP. Cholinesterase inhibitory effects of *Rhizophora lamarckii*, *Avicennia officinalis*, *Sesuvium portulacastrum* and *Suaeda monica*; Mangroves inhabiting an Indian coastal area (Vellar Estuary). *J Enzyme Inhib Med Chem*. 2009;24(3):702–707.
- Shekarchi M, Hajimehdipoor H, Naghibi F, et al. Investigating acetylcholinesterase inhibitory effects of some *Ferula* species. *J Med Plants*. 2013;12(46):107–113.
- Gearhart JM, Jepson GW, Clewell HJ, et al. Physiologically based pharmacokinetic model for the inhibition of acetylcholinesterase by organophosphate esters. *Environ Health Perspect*. 1994;102(11):51–60.
- Betancourt AM, Filipo NM, Carr RL. Alteration of Neurotrophins in the Hippocampus and Cerebral Cortex of Young Rats Exposed to Chlorpyrifos and Methyl Parathion. *Toxicol Sci*. 2007;100(2):445–455.
- Jortner BS, Hancock SK, Hinkley JF, et al. Neuropathological Studies of Rats Following Multiple Exposure to Tri-Ortho-Tolyl Phosphate, Chlorpyrifos and Stress. *Toxicol Pathol*. 2005;33(3):378–385.

23. Mukherjee PK, Kumar V, Houghton PJ. Screening of Indian medicinal plants for acetylcholinesterase inhibitory activity. *Phytother Res*. 2007;21(12):1142–1145.
24. Ashani Y, Peggins JO, Doctor BP. Mechanism of inhibition of cholinesterases by huperzine A. *Biochem Biophys Res Commun*. 1992;184(2):719–726.
25. Moss M, Cook J, Wesnes K, et al. Aromas of rosemary and lavender essential oils differentially affect cognition and mood in healthy adults. *Int J Neurosci*. 2003;113(1):15–38.
26. Ożarowski M, Mikołajczak PL, Kozłowska BT, et al. Neuroactive compounds from medicinal plants of the Lamiaceae family showing potentially beneficial activity in treatment of Alzheimer's disease. *Herba Polonica*. 2009;55(4):148–163.
27. Ghezzi L, Scarpini E, Galimberti D. Disease-modifying drugs in Alzheimer's disease. *Drug Des Devel Ther*. 2013;7:1471–1478.
28. Cummings JL, Morstorf T, Zhong K. Alzheimer's disease drug-development pipeline: Few candidates, frequent failures. *Alzheimers Res Ther*. 2014;6(4):37.
29. Kousba AA, Sultatos LG, Poet TS, et al. Comparison of Chlorpyrifos-Oxon and Paraoxon Acetylcholinesterase Inhibition Dynamics; Potential Role of a Peripheral Binding Site. *Toxicology Sci*. 2004;80(2):239–248.
30. Kousba AA, Poet TS, Timchalk C. Age-Related Brain Cholinesterase Inhibition Kinetics following In Vitro Incubation with Chlorpyrifos-Oxon and Diazinon-Oxon. *Toxicol Sci*. 2007;95(1):147–155.
31. Mills S, Bone K. *Principles and Practice of Phytotherapy: Modern herbal medicine*. Edinburgh: Churchill Livingstone; 2000.
32. Al-Jafari AA. The toxicological effect of cyclophosphamide on acetylcholinesterase activity. *Toxicol Lett*. 1993;66(2):125–131.
33. Uabundit N, Wattanathorn J, Mucimapura S, et al. Cognitive enhancement and neuroprotective effects of *Bacopa monnieri* in Alzheimer's disease model. *J Ethnopharmacol*. 2010;127(1):26–31.
34. Le Mercier G, Carpentier P, Sentenac Roumanou H, et al. Histological and histochemical changes in the central nervous system of the rat poisoned by an irreversible anticholinesterase compound. *Acta Neuropathol*. 1983;61(2):123–129.
35. Luty S, Latuszyńska J, Halliop J, et al. Toxicity of dermally absorbed alpha-cypermethrin in rat. *Ann Agric Environ Med*. 1998;5(2):109–116.
36. Latuszyńska J, Lut, S, Raszewski G, et al. Neurotoxic effect of dermally applied chlorpyrifos and cypermethrin Reversibility of changes. *Ann Agric Environ Med*. 2003;10(2):197–201.
37. Shih T, Duniho SM, McDonough JH. Control of nerve agent-induced seizures is critical for neuroprotection and survival. *Toxicol Appl Pharmacol*. 2003;188(2):69–80.
38. Aparna V, Mallya SV, Srikanth P, et al. Comparative pharmacognosy of two medhya dravyas, Brahmi (*Bacopa monnieri* Linn.) and Mandukaparni (*Centella asiatica* Linn.). *The Journal of Phyto pharmacology*. 2015;4(1):1–5.
39. Wu CK, Thal L, Pizzo D, et al. Apoptotic signals within the basal forebrain cholinergic neurons in Alzheimer's disease. *Exp Neurol*. 2005;195:484–496.
40. Gron G, Brandenburg I, Wunderlich AP, et al. Inhibition of hippocampal function in mild cognitive impairment: targeting the cholinergic hypothesis. *Neurobiol Aging*. 2006;27:78–87.
41. Descarries L, Aznavour N, Hamel E. The acetylcholine innervation of cerebral cortex: new data on its normal development and its fate in the hAPP (SW, IND) mouse model of Alzheimer's disease. *J Neural Transm*. 2005;112(1):149–162.