

Effects of cola on the rats' urine proteome

Abstract

The nature of biomarker is change. Contrast to the blood which is under homeostatic controls, urine reflects changes in the body earlier and more sensitive therefore is a better biomarker source. However, the urinary proteome is affected by many factors such as diet, medication, daily activities. We studied the effect of cola intake on urinary proteome, using animal models to minimize related factors. Urine samples before and after cola treatment were collected and analyzed by SDS-PAGE and LC-MS/MS. Spectral counts was used to perform semi-quantitative analysis. In total, 371 proteins were identified with FDR<1% and at least 2 peptides. To be conservative, all of the differential proteins met the following criteria: 1); T-test values P<0.05, 2) the variation trend in all three animals was consistent, and the fold change was at least 1.5 in each rat. Twelve proteins significantly changed after cola intake, including 4 increased and 8 decreased. Six significantly changed proteins were previously annotated as urinary candidate biomarkers. Cola is a common beverage around the world; thus, this differential protein should be considered for future diseases biomarker studies.

Keywords: cola, biomarker, urinary proteome

Introduction

Biomarkers are the measurable changes associated with physiological or pathophysiological process, and the most fundamental property of biomarkers is change.¹ Blood is under the control of homeostasis to remain stable and balanced for protecting organs from disturbing factors, so it tends to remove changes quickly. Urine, as a filtrate of blood, without homeostatic controls, can accumulate all kinds of changes so it is a better source for biomarker discovery than blood.^{1,2} So far, a wide range of candidate urinary biomarkers have been reported.³

However, urinary biomarkers research can be challenging for the fact that changes in urine are very complicated. There are various elements affecting urinary proteome, diet, medication, gender, age, daily rhythms, exercise, and hormone status.⁴ It is difficult to determine whether potential biomarkers are truly related to diseases. We tried to use rats to study their effects one factor at a time. The results can help to annotate the potential disease biomarkers that are affected by these confounding factors.

Cola is a very common beverage in our daily life. The purpose of our experiment is to find out whether drinking cola will cause changes on urinary proteome and determine which proteins are affected.

Materials and methods

Ethics statement and cola treatment

Four adult male Wistar rats (210-220g) were purchased from Charles River, China. All animals were kept with standard laboratory diet under controlled indoor temperature ($22\pm1^{\circ}\text{C}$) and humidity (65%-70%). The experiment was approved by Institutional Animal Care Use & Welfare Committee of Institute of Basic Medical Sciences, Peking Union Medical College (Animal Welfare Assurance Number: ACUC-A02-2013-015). For these 4 rats, all drinking water was replaced with cola (commercially available sucrose-sweetened carbonated drink, Coca-ColaTM, China). Before the cola treatment and at the fourth day after cola treatment, all rats were individually

Volume 4 Issue 3 - 2016

Kunyu Tang, Weiwei Qin, Ting Wang, Youhe Gao

College of Life Science, Beijing Normal University, China
*both authors have equal contribution

Correspondence: Youhe Gao, Department of Biochemistry and Molecular Biology, Beijing Normal University, Gene Engineering and Biotechnology Beijing Key Laboratory, Beijing, 100875, P.R. of China, Tel +86 10 58804382, Email gaoyouhe@bnu.edu.cn

Received: October 31, 2016 | **Published:** November 07, 2016

placed in the metabolic cages for 12hours. Then the twelve-hour urine samples were used for proteomics analysis.

Urinary proteins extraction and tryptic digestion

Urine was centrifuged at 3500g for 30min immediately after collection. Three volumes of ethanol were added after removing the pellets, and the samples were precipitated at 4°C. Then, 8M urea, 2M thiourea, 25mm dithiothreitol and 50mm Tris were used to re-dissolve the pellets. The protein concentration of each sample was measured by the Bradford protein assay. A filter-aided sample preparation method⁵ was used to digest the proteins in the three samples (each contained 200μg of urinary protein). Briefly, protein solution was reduced with 4.5mm DTT for 1h at 37°C, and then alkylated with 10mm IAA for 30min at room temperature in darkness. Finally, proteins were digested with trypsin (1μg/50μg protein) overnight at 37°C. The resulting peptides were desalting and then lyophilized for LC-MS/MS analysis.

LC-MS/MS analysis

The digested peptides were dissolved in 0.1% formic acid and loaded on a trap column (75μm×2cm, 3μm, C18, 100Å). The eluent was transferred to a reversed-phase analytical column (50μm×150mm, 2μm, C18, 100Å) by Thermo EASY-nLC 1200 HPLC system. Peptides were analyzed by Fusion Lumos mass spectrometer (Thermo Fisher Scientific). The Fusion Lumos was operated on data-dependent acquisition mode. Survey MS scans were acquired in the Orbitrap using a 350-1550m/z range with the resolution set to 120,000. The most intense ions per survey scan (top speed mode) were selected for collision-induced dissociation fragmentation, and the resulting fragments were analyzed in the Orbitrap. Dynamic exclusion was employed with a 30s window. Three technical replicate analyses were performed for each sample.

Protein identification and statistical analysis

The MS/MS data were processed using Mascot software (version 2.4.1, Matrix Science, London, UK) and searched against the Swiss-

Prot rat database (05/03/2013; containing 9,354 sequences). Search parameters were set as follows: 10 ppm precursor mass tolerance, 0.02 Da fragment mass tolerance, two missed cleavage sites allowed in the trypsin digestion, cysteine carbamidomethylation as fixed modification, oxidation (M) as variable modifications. The Mascot results were filtered and validated by Scaffold (version 4.0.1, Proteome Software Inc., Portland, OR, USA). Peptide identifications were accepted if they were detected with 90.0% probability and an FDR less than 0.1% by the Scaffold local FDR algorithm. Protein identifications were accepted if they were detected with FDR less than 1% and contained at least 2 identified peptides. Spectral counting was used to compare protein abundance between different experimental groups. To be conservative, all of the differential proteins met the following criteria: 1) T-test values $P < 0.05$, 2) the variation trend in all three animals was consistent, and the fold change was at least 1.5 in each rat.

Results and discussion

SDS-PAGE analysis of urinary proteins before and after cola treatment

25 μ g urinary proteins were separated using 12% SDS-PAGE gel. There were consistent changes before and after cola treatment. As is shown in Figure 1, during 55-100kDa and 130-180kDa, the bands are lighter after drinking cola, while about 15kDa, the bands are darker, indicating that drinking cola will have an obvious impact on the rat's urinary proteome (Figure 1).

Urine proteome changes identified by LC-MS/MS

To investigate the effects of cola intake on the urinary proteome, the urinary proteomes from three rats before and after cola intake were analyzed by LC-MS/MS. Six samples three rats (three samples from cola treatment before, and three after treatment) were analyzed by UPLC coupled with Fusion Lumos. In total, 371 proteins were

identified with FDR<1% and at least 2 peptides (Supporting Information Tables S1). For the total difference in spectra after the cola treatment, spectral counting was used to perform semi-quantitative analysis. The details of criteria are described in Materials and methods.

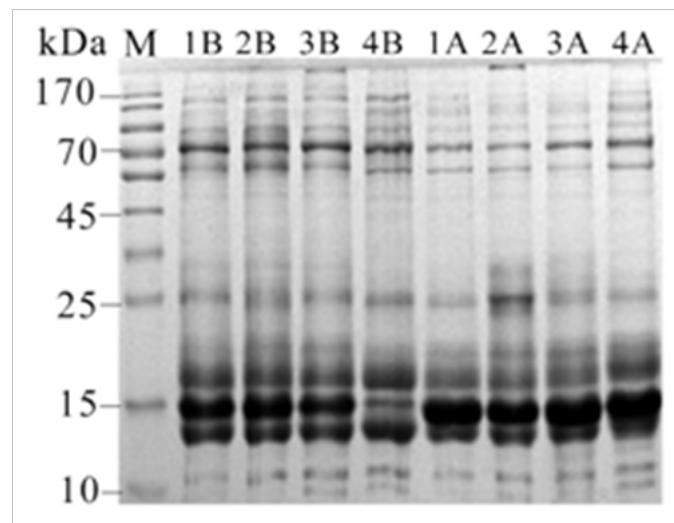


Figure 1 SDS-PAGE analysis of the urinary protein samples before and after cola treatment. (M, protein marker; B, before cola treatment; A, after cola treatment).

Twelve proteins were significantly altered after cola intake (Table 1). Four proteins increased and 8 decreased, 9 proteins have human orthologs. All significant changed proteins were searched against the Urinary Protein Biomarker Database.³ Six significantly changed proteins were previously annotated as urinary candidate biomarkers (Table 1). Cola is a common beverage around the world; thus, these protein changes should be considered when researching diseases biomarkers.

Table 1 Details of changed urinary proteins before and after drinking cola

ID	Protein name	MW (kDa)	PSM (Before)			PSM (After)			Fold change			Change tendency	Human ortholog	Diseases
			1	2	3	1	2	3	Rat1	Rat2	Rat3			
P20759	Ig gamma-1 chain C region 36	36	0	2	3	5	6	7	∞	3	2.3	UP	NO	None
P02625	Parvalbumin alpha	12	0	1	0	2	2	3	∞	2	∞	UP	P20472	Compound-induced skeletal muscle toxicity ⁶
P42854	Regenerating islet-derived protein 3-gamma	19	13	15	16	49	87	53	3.8	5.8	3.3	UP	NO	Membranous nephropathy ⁷
P27590	Uromodulin	71	77	74	63	149	140	164	1.9	1.9	2.6	UP	P07911	Sepsis-induced acute renal failure ⁸ Focal segmental glomerulosclerosis and mesangioproliferative glomerulonephritis ⁹ Preeclampsia ¹⁰
P07897	Aggrecan core protein	221	3	6	4	2	1	1	0.7	0.2	0.3	DOWN	NO	None
O35112	CD166 antigen	65	3	4	3	2	1	2	0.7	0.3	0.7	DOWN	Q13740	Type I diabetes ¹¹
P42123	L-lactate dehydrogenase B chain	37	5	4	6	3	1	2	0.6	0.3	0.3	DOWN	P07195	None
P01026	Complement C3	186	14	10	17	7	3	8	0.5	0.3	0.5	DOWN	P01024	None
P07150	Annexin A1	39	4	7	5	2	1	1	0.5	0.1	0.2	DOWN	P04083	Glomerular injury ¹² Interstitial fibrosis ¹³ HER2 breast cancer ¹⁴
P50399	Rab GDP dissociation inhibitor beta	51	4	4	3	2	2	2	0.5	0.5	0.7	DOWN	P50395	None

Table Continued....

ID	Protein name	MW (kDa)	PSM (Before)			PSM (After)			Fold change	Change tendency	Human ortholog	Diseases		
			1	2	3	1	2	3						
B5DFC9	Nidogen-2	153	3	4	4	1	1	2	0.3	0.3	0.5	DOWN	Q14112	None
Q9JJ40	Na(+)/H(+) exchange regulatory cofactor NHE-RF3	57	13	12	15	4	7	8	0.3	0.6	0.7	DOWN	Q5T2W1	Protein excretion with Sodium loading ¹⁵ Aldosteronism ¹⁶

Acknowledgements

Thanks to the Australian Proteome Analysis Facility (APAF) for funding this research.

Conflict of interest

The author declares no conflict of interest.

References

- Gao Y. Urine—an untapped goldmine for biomarker discovery? *Sci China Life Sci.* 2013;56(12):1145–1146.
- Li M, Zhao M, Gao Y. Changes of proteins induced by anticoagulants can be more sensitively detected in urine than in plasma. *Sci China Life Sci.* 2014;57(7):649–656.
- Shao C, Li M, Li X. A Tool for Biomarker Discovery in the Urinary Proteome: A Manually Curated Human and Animal Urine Protein Biomarker Database. *Mol Cell Proteomics.* 2011;10(11):M111.010975.
- Wu J, Gao Y. Physiological conditions can be reflected in human urine proteome and metabolome. *Expert Rev Proteomics.* 2016;12(6):623–636.
- Wisniewski JR, Zougman A, Nagaraj N, et al. Universal sample preparation method for proteome analysis. *Nat Methods.* 2009;6(5):359–362.
- Dare TO, Davies HA, Turton JA, et al. Application of surface-enhanced laser desorption ionization technology to the detection and identification of urinary arvalbumin- α : A biomarker of compound-induced skeletal muscle toxicity in the rat. *Electrophoresis.* 2002;23(18):3241–3251.
- Ngai HH, Sit WH, Jiang PP, et al. Serial Changes in urinary proteome profile of membranous nephropathy: implications for pathophysiology and biomarker Discovery. *J Proteome Res.* 2006;5(11):3038–3047.
- Holly MK, Dear JW, Hu XZ, et al. Biomarker and Drug Target Discovery Using Proteomics in a New Rat Model of Sepsis-Induced Acute Renal Failure. *Kidney Int.* 2006;70(3):496–506.
- Wang Y, Chen Y, Zhang Y, et al. Differential ConA-enriched urinary proteome in rat experimental glomerular diseases. *Biochem Biophys Res Commun.* 2008;371(3):385–390.
- Kononikhin AS, Starodubtseva NL, Bugrova AE, et al. An untargeted approach for the analysis of the urine peptidome of women with preeclampsia. *J Proteomics.* 2016;149:38–43.
- Suh MJ, Tovchigrechko A, Thovarai V, et al. Quantitative Differences in the Urinary Proteome of Siblings Discordant for Type 1 Diabetes Include Lysosomal Enzymes. *J Proteome Res.* 2015;14(8):3123–3135.
- Ka SM, Tsai PY, Chao TK, et al. Urine annexin A1 as an index for glomerular injury in patients. *Dis Markers.* 2014;2014:854163.
- Yuan Y, Zhang F, Wu J, et al. Urinary candidate biomarker discovery in a rat unilateral ureteral obstruction model. *Sci Rep.* 2015;5:9314.
- Gajbhiye A, Dabhi R, Taunk K, et al. Urinary proteome alterations in HER2 enriched breast cancer revealed by multipronged quantitative proteomics. *Proteomics.* 2016;16(17):2403–2418.
- Thongboonkerd V, Klein JB, Pierce WM, et al. Sodium loading changes urinary protein excretion: a proteomic analysis. *Am J Physiol Renal Physiol.* 2003;284(6):F1155–F1163.
- van der Lubbe N, Jansen PM, Salih M, et al. The phosphorylated sodium chloride cotransporter in urinary exosomes is superior to prostasin as a marker for aldosteronism. *Hypertension.* 2012;60(3):741–748.