

A computational comparative study of α -glucosidase enzyme divergence

Abstract

α -glucosidases (α -Gls) catalyze the last step of carbohydrate digestion in mammals and release glucose in bloodstream, which results in the increased blood glucose level. The evolution and classification of this enzyme has been a matter of debate. In the present study the amino acid sequences of α -Gls from 12 species were aligned and the Phylogenetic trees were constructed. The data indicated that only the mammalian enzyme contained the glucoamylase region. Five amino acid regions were found to be the conservative blocks of mammalian α -Gls. These blocks were not fully conserved in plants, fungi and bacteria. The results were in favor of Chiba's classification despite the Ile \rightarrow Thr substitution in *Aspergillus Niger* and Ala \rightarrow Pro substitution in chimpanzee and human α -Gls. The chimpanzee α -Gls showed the most similarity to human enzyme. Plants, fungi, bacteria, and the mammalian α -Gls seemed to separately create a specific class. Together, the data suggest that the eukaryotic enzyme had been diverged significantly from the prokaryotic α -Gls since the separation from the common ancestor.

Keywords: α -glucosidase, phylogenetic trees, evolution, multiple alignments

Volume 2 Issue 4 - 2015

Mohammad Hossein Mehraban,^{1,2} Younes Ghasemi,² Sadeq Vallian¹

¹Department of Biology, University of Isfahan, R Iran

²Department of Pharmaceutical Biotechnology, Shiraz University of Medical Sciences, IR Iran

Correspondence: Sadeq Vallian Professor of Human Molecular Genetics Division of genetics, Department of Biology, Faculty of Science, University of Isfahan, Isfahan, Iran, Tel +983117932456, Email svallian@sci.ui.ac.ir

Received: March 07, 2015 | **Published:** October 19, 2015

Introduction

α -Glucosidase (α -Gls) enzymes have a crucial role in digestion of carbohydrates and biosynthesis of glycoprotein.^{1,2} These enzymes hydrolyze terminal glycoside bonds and release α -glucose from the substrate chain. They are found in prokaryotic cells, plants and mammalian tissues such as liver, blood and especially intestine.¹ The last step of carbohydrate digestion is catalyzed by α -glucosidases (namely maltase-glucoamylase or MGAM in mammals) which results in liberation of glucose. In *Escherichia coli* and viruses such as human immunodeficiency virus (HIV) they are necessary for cell wall and envelope construction.² In plants, the glucose produced by the activity of these enzymes is used as an energy source for cellular growth and development. These enzymes are distributed in plant tissues like seeds, fruits, leaves and roots. In the absence of α -amylase it is suggested that α -Gls can initiate the degradation of natural starch granules in barely seeds and pea chloroplasts.³⁻⁵ Their important role in treating degenerative diseases such as diabetes mellitus type-II and HIV has been proved in many studies.⁶⁻¹¹ Although, the enzymes exist in monomeric form in prokaryotes, plants and primary eukaryotes, hetero and homodimeric structures were found in advanced Eukaryotes.^{1,12} Therefore, the classification of α -Gls enzyme is of great importance for enzymologists and evolutionists. According to Chiba's classification, *Escherichia coli* and other bacterial species constitute family I of α -Gls.¹³ On the other hand, α -Gls from mammalian tissues, plants, *Candida tsukubaensis*, and *Mucor javanicus* constitute the family II. The basis of this classification is the amino acid conservative blocks existed in these enzymes. The amino acid block of DLVINH, EVAH and YIENHD seems to be present in class I. However the conservative blocks of class II are GIWADMNEV and GADICGF.¹ In the present study the aim was to assess the similarities between the two classes using evolutionarily relationship and Phylogenetic data. Currently it is not known how α -Gls had been evolved and how many species have the enzyme, and what the amino acid sequence of the ancestor was. Answer to these questions will facilitate identification of primitive and specialized aspects of α -Gls structure, function and regulation.

Materials and methods

Amino acid sequences of *E. coli*, *Sulfolobus solfataricus*, yeast (*Candida tsukubaensis*), barley (*Hordeum vulgare*), *Aspergillus Niger*, spinach (*Spinacia oleracea*), sugar beet (*Beta vulgaris*), mouse (*Mus musculus*), rat (*Rattus norvegicus*), chimpanzee (*Pan Troglodyte*) and human (*Homo sapiens*) α -glucosidase enzyme were retrieved from Uniprot (protein data base). The full sequence of each entity was used. The sequences were aligned and the Phylogenetic trees were constructed. Clustal X version 2.1¹⁴ was used for multiple alignment and Phylogenetic trees were constructed using Mega 6 software.¹⁵ Trees were created using maximum-likelihood approach, and 1000 bootstrap replicates were conducted.

Results and discussion

In the present study a comparative investigation on the divergence of α -glucosidase (α -Gls) enzyme was performed. The complete amino acid sequences of different species as stated in the materials and methods were aligned. The mammalian α -Gls enzymes are heterodimeric proteins with α -glucosidic activity. The enzymes contain a region responsible for the catalytic site for an enzyme termed glucoamylase. The comparative results from the present investigation showed that human and chimpanzee α -glucosidases had the closest similarity, and may have come from the same ancestor. There are two conserve P-type domains in human MGAM located in positions 88-134 and 954-1000,¹⁶ which is completely conserved in chimpanzee's enzyme. These domains have some synonymous amino acid alterations in rat and mouse MGAM. Interestingly, these domains are absent in plants and bacterial counterparts. There are two catalytic aspartic acid residues in mammalian enzymes which due to their nucleophilic nature has a major role in hydrolyzing glucosidic chains.¹⁶ One of them which are located in position 529 is conserved among all the species studied. This residue is in the maltase region of mammalian MGAM and is conserved in plants and bacterial enzymes, which improves the importance of this residue in catalytic site. The other aspartic acid residue is in position 1420 which is located in

glucoamylase region of mammalian MGAM. This is a remarkable feature of mammalian enzymes which is absent in plants and bacterial α -Gls. These characteristics make mammalian α -Gls a unique class which has different structure and function in comparison to plants and bacterial enzymes. Rat and mouse α -Gls showed a significant similarity with human enzyme, but obviously they had not been diverged from a common ancestor. It was noteworthy that five amino acid regions were found to be the conservative blocks of mammalian α -Gls which was summarized in (Table 1). These blocks were not fully conserved in plants, fungi and bacteria (Figure 1). *Aspergillus*

Niger, spinach, sugar beet, barley and yeast (*Candida tsukubaensis*) α -Gls exist in monomeric form. The plant enzyme proved to create a different subclass since sugar beet; barley and spinach shared more similar amino acid sequences with each other. On the other hand, *Aspergillus Niger* and yeast may be considered as the other subclass due to their sequence homology. According to Chiba's classification, plants, mammalian, yeast and *Aspergillus Niger* α -Gls are considered the family II of α -Gls as they all have a conservative block of GADICGF.^{1,13}

Table 1 Conservative blocks of mammalian α -Gls

Entry	Block 1	Block 2	Block 3	Block 4	Block 5
Mammalian α -Gls	185-LLTAEYQTSN RFHFKLTDQT	236- PFSIKVTRRSN NRVLFDSISGP	292- NRYGLGEHVHQQ YRHDMNWKTWP	854- LFCKTLCM DAVQHWG	900- KRSFILTRSTFAGSGK FAAHWLGDNATATW

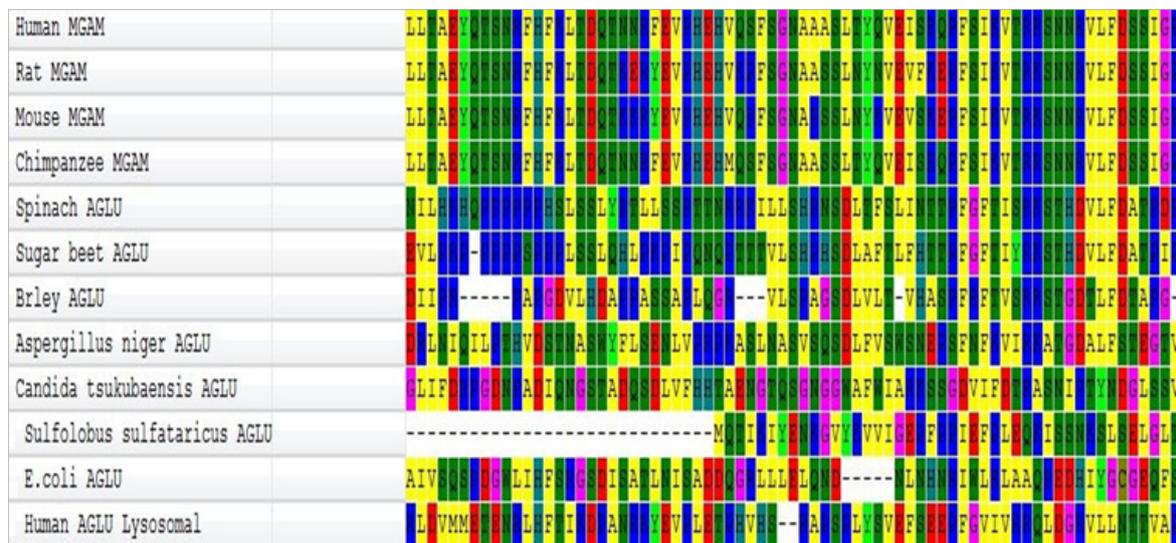


Figure 1 Multiple sequence alignment of α -Gls enzyme from collected amino acid sequences.

Our results were in favor of the above theory despite the Ile→Thr substitution in *Aspergillus Niger* and Ala→Pro substitution in chimpanzee and human α -Gls. However, due to the presence of glucoamylase site in mammalian enzymes, the classification could be changed in a way that the mammalian enzymes would be considered as an independent class. The results showed that *E. coli* and *Sulfolobus sulfataricus* α -Gls sequences were significantly different from plants and mammalian α -Gls. Interestingly it was proved that despite the mammalian α -Gls other enzyme counterparts do not contain the conservative blocks which was part of glucoamylase region of mammalian α -Gls. These conservative blocks were summarized in (Table 2). It might be concluded that during the evolutionary state of mammals, this region was inserted or had some *de novo* mutations which resulted in a perfect active glucoamylase that could produce β -glucose. The classification of α -Gls enzyme is of great importance to understand the structure and function of this crucial enzyme, and might help evolutionist understand the origin of this enzyme. As mentioned above, the inhibition of this enzyme had a major role in

treating diabetes type II and AIDS. In model organisms like yeast, mouse and *E. coli*, α -Gls has been used in many studies.¹⁷⁻²¹ As the concentration of human enzyme is scarce, the essential role of the enzyme of these model organisms has been bolded. Based on our results, the chimpanzee's enzyme showed the closest relation to human counterpart and might always be the best substitute. However, the yeast and *E. coli* α -Gls seemed to be structurally different from the mammals one and could not be considered as a suitable model. Based on the evolutionary point of view, it seemed that chimpanzees and humans share the most identical sequences and domains, and may have a common ancestor which a long time ago diverged from the mouse and rat ancestor. The plants α -Gls was significantly different from the mammals one since they lack the glucoamylase site. However, the homology of their maltase site was quite noticeable. Therefore, as depicted in (Figure 2), the eukaryotic enzyme had been diverged significantly from the prokaryotic α -Gls since the separation from the common ancestor.

Table 2 Conservative blocks of mammalian glucoamylase

Entry	Block 1	Block 2	Block 3	Block 4	Block 5
Mammalian Glucoamylase	1314- EKIDCYPDEN- GAS	1361- QYNHSGATA- DISLK	3250- IIFWDSQLLG- FTFSDMFIRISTRLP	3305- PPGYKKN- SYGVHPYYMGLEEDG- SAHG	3355- GGILDFYV- FLGPTPEIVTQQYTELI- GRPVMVPYWSLG- FQLCRYGY

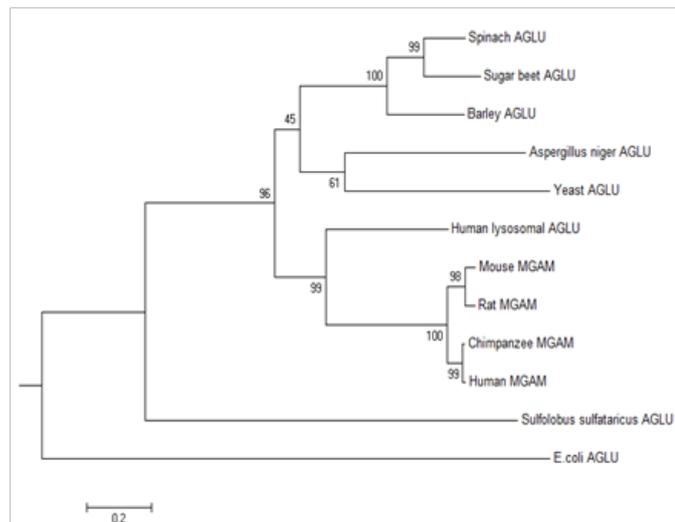


Figure 2 Phylogenetic tree of the α -Gls enzyme family. The numbers at each node are the bootstrap support values obtained by maximum likelihood.

Acknowledgements

None.

Conflict of interest

The author declares no conflict of interest.

References

- Krasikov VV, Karelov DV, Firsov LM. Alpha-Glucosidases. *Biochemistry (Mosc)*. 2001;66(3):267–281.
- Bharatham K, Bharatham N, Park KH, et al. Binding mode analyses and pharmacophore model development for sulfonamide chalcone derivatives, a new class of α -glucosidase inhibitors. *J of Mol Graph Model*. 2008;26(8):1202–1212.
- Sun Z, Duke SH, Henson CA. The Role of Pea Chloroplast [alpha]-Glucosidase in Transitory Starch Degradation. *Plant physiol*. 1995;108(1):211–217.
- Sun Z, Henson CA. Degradation of native starch granules by barley α -glucosidases. *Plant physiol*. 1990;94(1):320–327.
- Sissons M, MacGregor A. Hydrolysis of barley starch granules by α -glucosidases from malt. *Journal of Cereal Science*. 1994;19(2):161–169.
- Mehta A, Zitzmann N, Rudd PM, et al. α -Glucosidase inhibitors as potential broad based anti-viral agents. *FEBS lett*. 1998;430(1–2):17–22.
- Gruters RA, Neeffjes JJ, Tersmette M, et al. Interference with HIV-induced syncytium formation and viral infectivity by inhibitors of trimming glucosidase. *Nature*. 1987;330:74–77.
- Yoshikawa M, Shimada H, Nishida N, et al. Antidiabetic principles of natural medicines. II. Aldose reductase and alpha-glucosidase inhibitors from Brazilian natural medicine, the leaves of *Myrcia multiflora* DC. (Myrtaceae): structures of myrciacitrins I and II and myrciaphenones A and B. *Chem Pharm Bull*. 1998;46(1):113–119.
- Yousefi R, Alavian Mehr MM, Mokhtari F, et al. Pyrimidine-fused heterocycle derivatives as a novel class of inhibitors for α -glucosidase. *J enzyme Inhib Med Chem*. 2013;28(6):1228–1235.
- Yu W, Gill T, Wang L, et al. Design, synthesis, and biological evaluation of N-alkylated deoxynojirimycin (DNJ) derivatives for the treatment of dengue virus infection. *Journal of medicinal chemistry*. 2012;55(13):6061–6075.
- Becq F, Norez C. *Use of glucosidase inhibitors for therapy of mucoviscidosis*. Google Patents, 2012.
- Nairn AV, Moremen KW. Glucosidase, alpha neutral AB; glucosidase II subunit beta (GANAB, PRKCSH, α -glucosidase II). In: Taniguchi N, editor. *Handbook of Glycosyltransferases & Related Genes*. 2nd ed. Tokyo: Springer-Verlag; 2014. p. 1283–1295.
- Chiba S. Molecular mechanism in alpha-glucosidase and glucoamylase. *Biosci Biotechnol Biochem*. 1997;61(8):1233–1239.
- Larkin MA, Blackshields G, Brown NP, et al. Clustal W and Clustal X version 2.0. *Bioinformatics*. 2007;23(21):2947–2948.
- Tamura K, Stecher G, Peterson D, et al. MEGA6: Molecular Evolutionary Genetics Analysis Version 6.0. *Mol Biol Evol*. 2013;30(12):2725–2729.
- Sim L, Quezada-Calvillo R, Sterchi EE, et al. Human intestinal maltase-glucoamylase: crystal structure of the N-terminal catalytic subunit and basis of inhibition and substrate specificity. *J Mol Biol*. 2008;375(3):782–792.
- Rengasamy KR, Aderogba MA, Amoo SO, et al. Potential antiradical and alpha-glucosidase inhibitors from *Ecklonia maxima* (Osbeck) Papenfuss. *Food chem*. 2013;141(2):1412–1415.
- Kim KT, Rioux LE, Turgeon SL. Alpha-amylase and alpha-glucosidase inhibition is differentially modulated by fucoidan obtained from *Fucus vesiculosus* and *Ascophyllum nodosum*. *Phytochemistry*. 2014;98:27–33.
- Nair SS, Kavrekar V, Mishra A. *In vitro* studies on alpha amylase and alpha glucosidase inhibitory activities of selected plant extracts. *Eur J Exp Biol*. 2013;3(1):128–131.
- Phan MAT, Wang J, Tang J, et al. Evaluation of α -glucosidase inhibition potential of some flavonoids from *Epimedium brevicornum*. *LWT-Food Science and Technology*. 2013;53(2):492–498.
- Thanakosai W, Phuwapraisirisan P. First identification of alpha-glucosidase inhibitors from okra (*Abelmoschus esculentus*) seeds. *Natural product communications*. 2013;8(8):1085–1088.