LEKTI, a physiological inhibitor of multiple serine proteinases, suppresses perineural and lymphovascular invasion of human head and neck cancer cells in a mouse orthotopic model

Abstract

Serine Protease Inhibitor Kazal-type 5 (SPINK5) gene encodes 3 different Lympho-Epithelial Kazal-Type-Inhibitor (LEKTI) isoforms. We identified and cloned LEKTI by its constitutive expression in normal oral mucosa and its loss of expression in matched tumor specimens of patients with head and neck squamous cell carcinoma (HNSCC). Stable re-expression of LEKTI in HNSCC OSC19 cells resulted in reduced migration and invasion and enhanced adhesion on variety of ECM substrates in vitro with a concomitant reduction inexpression of endogenous MMP-14, MMP-8, KLK5, and ADAM8. Here, we sought to determine the consequences of LEKTI re-expression on the in vivo changes in the tumor growth and invasion using an orthotopic model of tongue cancer. In the tongue tumors of mice, lymphovascular invasion or perineural spread was found in 100% of tumors derived from parental cell lines but was almost totally absent in all tumors derived from LEKTI-expressing clones. Our work suggests that loss of LEKTI expression in primary tumors might correlate with aggressive biologic behavior and restoration of LEKTI expression by pharmacologic means might be beneficial for patients with HNSCC.

Keywords: SPINK5, LEKTI, lymphovascular invasion, perineural spread, HNSCC

Abbreviations: LEKTI, lympho-epithelial kazal-type-inhibitor; SPINK5, serine protease inhibitor kazal-type 5; HNSCC, head and neck squamous cell carcinoma; MMP, matrix metalloproteinases; ECM, extracellular matrix; DMEM, dulbecco’s modified eagle’s medium

Introduction

Local tumor invasion and regional lymphatic metastasis occur by the attachment of tumor cells to components of the extracellular matrix (ECM) and by degradation of ECM by protease enzymes elaborated into the tumor microenvironment. These processes are regulated by such proteolytic enzymes as serine proteinases, cysteine proteinases, and matrix metalloproteinases (MMPs) tightly balanced by their endogenous inhibitors in the tumor microenvironment. Thus, inhibition of such proteinases can disrupt critical steps of invasion and metastasis. Indeed, several protease inhibitors have shown importance in a range of cancer types by the loss of expression correlating with advanced tumor progression. Moreover, proteinase inhibitors have been demonstrated to hold tumor suppressor functions in vitro including plasmin, trypsin, cathepsin G, human KLKs, and elastase, enzymes implicated in the activation of MMPs. We also showed that pro-LEKTI is processed into four cleavage products ranging in size from 37 to 60kDa in human oral keratinocytes.

In a recent study, we stably over expressed LEKTI in HNSCC cells and evaluated the effects of restored LEKTI on cellular proliferation, morphology, adhesion, invasion and expression of key MMPs involved in tumor progression. We demonstrated that stable expression of LEKTI in OSC-19 cells resulted in markedly decreased levels of expression of genes encoding MMP-9, MMP-14, KLK5, and ADAM8. Furthermore, LEKTI over expressing cells displayed striking morphological changes are more adhesive and less invasive. These results demonstrate a novel negative regulatory role for LEKTI in modulating the production of key MMPs involved in ECM degradation and suggest that loss of LEKTI in HNSCC tumor cells could have a pivotal role in HNSCC progression.

In the present work, we sought to determine the consequences of LEKTI re-expression on the in vivo changes in the tumor growth and invasion using an orthotopic model of tongue cancer. In the tongue tumors of mice, lymphovascular invasion or perineural spread was found in 100% of tumors derived from vector or parental cell lines but was almost totally absent in all tumors derived from LEKTI-expressing clones (p=0.008 by Fisher’s exact test).

An inhibitor of multiple serine proteinases, lymphoepithelial kazal-type inhibitor (LEKTI), was identified and cloned in our laboratory on the basis of its constitutive expression in normal oral mucosa and loss of its expression in matched head and neck squamous cell carcinoma (HNSCC) specimens and multiple HNSCC lines. It was also shown by several investigators that LEKTI protein was encoded by SPINK5 gene and mutations in SPINK5has been linked to the inherited disorder known as Netherton Syndrome.

We produced recombinant full length LEKTI and several of its fragments using baculovirus expression system and established that recombinant human LEKTI inhibits a battery of serine proteinases in vitro including plasmin, trypsin, cathepsin G, human KLKs, and elastase, enzymes implicated in the activation of MMPs.

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Materials and methods

Materials

A human oral squamous cell carcinoma cell strain, OSC19, was a gift from Dr. Theresa Whiteside. Female athymic nude mice aged 6-8 weeks were purchased from Harlan Sprague-Dawley, Inc., Indianapolis, IN; SDS-polyacrylamide gels and pre-stained markers were procured from Bio-Rad Laboratories, Hercules, CA; nitrocellulose membrane from Schleicher & Schull BioScience, Keene, NH; horseradish peroxidase-conjugated goat-anti-mouse IgG (H+L) were obtained from Jackson Immuno Research Laboratories, West Grove, PA; lipofectamine 2000 and pLNA3.1 (-) were procured from Invitrogen, Carlsbad, CA; Kodak X-AR5 films were purchased from Eastman Kodak, Rochester, NY; hematoxylin and eosin (H&E) were purchased from Sigma-Aldrich, St. Louis, Mo.

Isolation of OSC-19 LEKTI stable clones

OSC19 cells were maintained in Dulbecco’s modified Eagle’s medium (DMEM) supplemented with 10% fetal bovine serum, 2mM glutamine, and antibiotics. The pro-LEKTI expression plasmid encodes the entire full-length LEKTI polypeptide and has a hexahistidine tag at its C-terminus. OSC-19 cells were stably transfected with the human LEKTI cDNA cloned into the pcDNA3.1 vector or a control vector without LEKTI insert after G418 (0.5 mg/ml) selection. OSC19, OSC19-LEKTI-expressing clone 11, or OSC19-LEKTI-expressing clone 17 were cultured at 37 °C in humidified incubator with 5% CO₂.

Orthotopic model of tongue cancer

Tongue tumors in mice were generated by modifications of orthotopic xenograph models previously described by Kawashir. Mice were cared for in accordance with the Institutional Animal Care and Use Committee and the Department of Veterinary Medicine of M. D. Anderson Cancer Center. Inoculation with OSC-19 parental, vector, LEKTI-expressing clone 11, or LEKTI-expressing clone 17 cells was randomly allocated in forty mice. In preparation for inoculation, cells were trypsinized and resuspended in DMEM at 8x10⁶ cells/mL. Animals were marked, weighed, and anesthetized by intraperitoneal injection of ketamine cocktail at 10µl/g of body weight. Inoculation was performed by injecting 25µl of cell suspension to deliver 200,000 cells into the submucosa of the dorsal tongue at the circumvallate line. Mice were maintained on a gel-consistency diet and monitored for weight loss. The animals were euthanized 21days after inoculation and the tongue, cervical lymph nodes, and lungs were removed by microsurgical dissection. Specimens were formalin-fixed, paraffin-embedded, serially sectioned at 20µm levels, and stained with hematoxylin and eosin (H&E). Histologic slides were independently reviewed by two investigators unaware of which inoculation group the specimens belonged.

Results and discussion

Summary of gross findings

Inoculating the tongue submucosa of athymic nude mice with OSC-19 cells predictably results in xenograft tumors with locally invasive growth. To examine the changes in invasive growth characteristics of tumors in which LEKTI expression was returned to the microenvironment, we inoculated mice with OSC-19 parental, vector LEKTI-expressing clone 11 or LEKTI-expressing clone 17 cells (Table 1). We reviewed serial sections of histologic specimens for pathologic features of tumor invasion (Table 2). An endophytic growth pattern with or without mucosal ulceration was noted. Grossly normal appearing regional lymph nodes were noted. Gross bone invasion in one of 40 animals was observed. No gross evidence of lung metastases was observed.

Table 1 Orthotopic tongue model

<table>
<thead>
<tr>
<th>40 Athymic Nude Mice</th>
<th>Inoculation of the dorsal tongue with 200,000 cells in 25µl</th>
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<tbody>
<tr>
<td>10 mice</td>
<td>10 mice</td>
</tr>
<tr>
<td>OSC-19 Parental</td>
<td>OSC-19 Vector</td>
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<tr>
<td>19 mice</td>
<td>19 mice</td>
</tr>
<tr>
<td>OSC-19 Clone 11</td>
<td>OSC-19 Clone 17</td>
</tr>
<tr>
<td>1 animal</td>
<td>1 animal</td>
</tr>
<tr>
<td>1 day post inoculation 14</td>
<td>1 day post inoculation 14</td>
</tr>
<tr>
<td>Euthanasia at 21 days. 6 specimens from each group formalin-fixed, paraffin-embedded, serially sectioned at 20 µm, and H&amp;E stained.</td>
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</tbody>
</table>

Table 2 Histological evaluation

<table>
<thead>
<tr>
<th>Blinded review</th>
<th>Regional metastasis</th>
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<tbody>
<tr>
<td><strong>Primary tumor</strong></td>
<td></td>
</tr>
<tr>
<td>i. Presence or absence</td>
<td>i. Presence or absence</td>
</tr>
<tr>
<td>ii. Axial, Transverse, AP Dimensions to calculate Tumor volume</td>
<td>ii. Number of nodes identified</td>
</tr>
<tr>
<td>iii. Pattern of Invasion</td>
<td>iii. Number of nodes positive</td>
</tr>
<tr>
<td>iv. Differentiation</td>
<td>iv. Metastatic grade</td>
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<tr>
<td>v. Presence or absence of</td>
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<tr>
<td>• Ulceration</td>
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<tr>
<td>• Lymphovascular invasion</td>
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<tr>
<td>• Perineural invasion</td>
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<tr>
<td>• Bone invasion</td>
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**Statistical analysis**

i. Comparisons between
• Vector and Parental
• Clones and vector

Chi Squared Test or Wilcoxon Rank Sum Test

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OSC-19 parental and vector lines showed 70% and stable clones showed 60% tumorigenicity (Figure 1, bottom left histogram). Moreover, re-expression of LEKTI had no effect on tumor volume (Figure 1, bottom right histogram). However, tumors derived from OSC-19-LEKTI-expressing clone 17 demonstrated no perineural invasion compared to perineural invasion in 100% of tumors derived from parental cells (Figure 2, top left histogram). Representative micrographs of typical tumors from parental cells show multiple tumor cells invading into the perineurium of small nerves (Figure 2, top left panel, arrow head), whereas tumors from OSC-19-LEKTI-expressing clone 17 show tumor nests in the proximity of nerves without invasion (Figure 2, top right panel, arrow head).

Similarly, tumors derived from OSC-19-LEKTI-expressing clone 17 demonstrated 17% lymphovascular invasion compared to lymphovascular invasion in 100% tumors derived from parental cells (Figure 2, bottom left histogram). Representative micrographs of typical tumors from parental cells show multiple tumors cells invading beyond the basement membrane of vascular and lymphatic vessels (Figure 2, bottom left panel, arrow head), whereas tumors from OSC-19-LEKTI-expressing clone 17 show tumor nests in the proximity of vessels without invasion (Figure 2, bottom right panel, arrow head).

Together these findings support a role for LEKTI as a tumor suppressor gene in head and neck squamous cell carcinoma. Indeed, several studies have reported the differential expression of proteinase inhibitors as markers for diagnosis and prognosis in various primary cancer types. For instance, the differential expression of maspin has been associated with good prognostic factors in breast cancer, increased survival and longer remission after resection of lung cancer, and low malignant potential in ovarian cancer. Such studies have proven vital to generating hypotheses of functional implications for further study.

Our findings in a mouse xenograph model of tongue cancer suggest that LEKTI expression results reduce tumor invasion potential in vivo. The prognostic significance of perineural invasion in head and neck squamous cell carcinoma warrants the investigation of mechanisms underlying this process. To date, the understanding of specific mechanisms has been limited by the difficulty in establishing appropriate models. However, analysis of human squamous cell carcinoma specimen by immunohistochemistry has found correlation between Laminin-5 staining and the presence of perineural invasion in head and neck squamous cell carcinoma, suggesting that expression of Laminin-5 is an important step in the process of perineural invasion. Thus, mechanisms of disrupting the processing of Laminin-5 in relation to tumor invasion may underlie the effect of LEKTI expression in mouse tumors. Our recent published result showing stable LEKTI re-expression blocks migration and invasion of HNSCC OSC-19 cells and immunohistochemical analysis of the graded loss of LEKTI expression along the spectrum of epithelial dysplasia suggests a role for the protein as a marker of differentiation that is lost early in tumor progression. Moreover, similar studies to correlate levels of LEKTI expression with perineural invasion in head and neck squamous cell carcinoma tumor specimen are ongoing.

Figure 1 Orthotopic tongue model showing the technique of injection of cancer cells and a resected tongue specimen from a mouse.

Figure 2 LEKTI re-expression reduces the frequency of perineural and lymphovascular invasion in the orthotopic tongue model.

Together, lymphovascular invasion or perineural spread was found in 100% of tumors derived from vector or parental cell lines but was almost totally absent in all tumors derived from LEKTI-expressing clones (p=0.008 by Fisher’s exact test).

In summary, in a xenograph model of tongue cancer, tumors derived from LEKTI-expressing clones of an invasive head and neck cell line demonstrated limited pathologic features of lymphovascular and perineural invasion. Our findings suggest an important negative regulatory role for LEKTI in modulating extracellular matrix degradation. The loss of LEKTI expression in head and neck squamous cell carcinoma may represent a critical event in tumor progression to an invasive cellular phenotype. Restoration of LEKTI expression by pharmacologic means might be beneficial for patients with HNSCC.

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Author contributions

A.J, T.D.S, and G.L.C provided intellectual input into the experimental design and execution of the study. A.J and T.D.S wrote the manuscript. K.B. generated LEKTI stable clones. H.K.W. and T.D.S performed most of the experiments. V.R. helped to organize the data.

Conflict of interest

The author declares no conflict of interest.

References


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