

Research Article





Identification of plants releasing isoprene causing smog

Abstract

The accessibility of water is most important component for plant productivity and growth. Water stress is a major threat for the agriculture system of Pakistan. The change in climate causes the change in the patterns of rainfall which may cause the extreme events. The main cause of climate change is the concentration of carbon dioxide in the air and global warming. A change in environment overall takes place by a number of causes but the most important cause of all these changes is isoprene. Water scarcity causes high isoprene emission from plants. The dominant volatile organic compound which is released from vegetation is Isoprene (2-methyl-1,3,-butadiene). Isoprene produced by few herbaceous plant species and many woody trees. The emission of these volatiles like isoprene has an indirect effect on climate change and cause smog formation. The examination found that isoprene responds with human made nitrogen oxide to make particulate issue. In light of the presence of nitrogen oxides, it is engaged with creating the negative impacts on climate and human wellbeing. The research investigates current knowledge about the presence of isoprene emissions within the plant kingdom under drought stress conditions. Eucalyptus, Kachnar, Mulberry and Conocarpus are few species which are able to produce isoprene. The pot experiment was designed to identify isoprene emission within these plants. The plants were exposed to decrease Fractional Transpirable Soil Water FTSW. Various VOCs emission was increase under stress conditions. The isoprene emission rate is high under mild drought stress but decline under severe drought stress. Other parameters like stomatal conductance, photosynthesis, internal CO2 were also observed.

Keywords: index terms-, isoprene, smog, GC-FID

Volume 7 Issue 2 - 2022

Tehreem Lutaf Ullah, Tariq Mahmood, Sanad Elhddad, Shagufta Usman, Sikandar M Zulgarnain, Reema Aslam, Mahnoor Zafar

¹Department of Environmental Sciences, Pir Mehr Ali Shah Arid Agriculture University Rawalpindi, Pakistan

²Department of Pharmaceutical Chemistry, Omar Almukhtar, University Libya

³Faculty of pharmaceutical sciences Riphah International University Islamabad. Lahore campus. Pakistan

⁴Department of Botany the Islamia University Bahawalpur,

⁵Department_ School of Public Health - Dow University of Health Sciences Karachi Pakistan

⁶Department of Environmental Science, COMSATS University, Islamabad, Abbottabad Campus Pakistan

Correspondence: Tehreem Lutaf Ullah, Department of Environmental Sciences, Pir Mehr Ali Shah Arid Agriculture University Rawalpindi, Punjab, Pakistan, Email angeljenni I 6@gmail.com

Received: March 15, 2022 | Published: April 08, 2022

Introduction

Given the potential impacts, climate change is always a concern for scientists. Increased environmental temperature and carbon dioxide are shown to be factors in the gradual climate change.\(^1\) The Increase in carbon dioxide concentration in atmosphere has been detected in the past due to human activities like urbanization. It is observed that carbon dioxide amount will increase up to 750 ppm at the end of this era. The earth's temperature has been increased to 0.4 - 0.660c in the present due to increase in CO2.\(^1\) Environmental changes have resulted in variations in rainfall patterns, magnitude and rate, resulting in irregular, low and unpredictable rainfall patterns that result in availability of water at native and geographical levels. Further, the current global change example promotes regional drought by predicting rising temperatures and prolonged drought with less rainfall.\(^2\)

Most significant components for the growth of plant and for their biochemical functions are water and CO2. Carbon uptake is affected by the enlarged amount of CO2 during the cellular gas exchange process, whereas a low availability of water prevents physiological functions such as the exchange of leaf gases, light synthesis and exchange of leaf gases, which eventually lead to prevent the growth and productivity of plants. The shortage of water is going to be high in the future. It will cause climate change and drought conditions in various parts of the world, which has become an important factor in limiting plant performance, leading to improved plant efficiency and production of biomass decreases.

During conditions of drought stress, the plants absorb smaller amount of water than transpiration and if this situation continues they target plant by damaging their growth, number of leaves, leaf area, fading, cell membranes and eventually lead to plant death. The main effect of limited supply of water to the plant trunk is partial or complete closing of the stomata, which generally reduces the conversation of water and carbon dioxide with the exterior environment. Less stomata conductivity decreases the level of internal carbon dioxide for photosynthesis. Low water availability also reduces photosynthesis process by decreasing in the area of leaf and photo unit rate unit per area.

Limited or high drought conditions affect the gaseous exchange and dynamics of carbon in plants in the ecosystem. To cope with stress situations, a large amount of volatile compounds are discharge by plants into the environment. During common circumstances, factories discharge about 3% saturated carbon in the manufacture of Biological Volatile Organic Compounds. Though, specifically in the case of drought stress, the plants are given carbon above 11% in the form of VOCs. **

Biological volatile organic compounds are a huge group of volatile compounds, with almost 1, 1,700 substances obtained from the plant identified. These compounds include a large variety of organic species, which includes isoprene, hemiterpenes, terpenes, monoterpenes, isoprenoids, and compounds of oxygen. There are many plants which released great amounts of isoprene. Eucalyptus, Wild Mulberry, Kachnar and Conocarpus are among the few species of plants that can release isoprene as well as monoterpenes. Monoterpenes are present in huge amount are usually kept in the tanks, while isoprene is present in free form and not in stored form, that's why it is openly released into the environment. The worldwide natural source rate is estimated per year is 1.1 to 1.6Pg Carbon¹¹ and this amount is much higher than human productions minutes. The worldwide natural source rate is estimated per year is 1.1 to 1.6Pg Carbon¹¹ and this amount is much higher than human productions minutes.

Biocompatible organic compouSnds (BVOCs) originated in marine and terrestrial ecosystems. About 80% of volatile compounds



are spread by trees and plants, in which the most important species are monoterpenes and isoprene. Deep farming programs started in 1979 to improve plants in India in 2005. In these programs different plant species were grown in different areas including rural and urban. The extensive cultivation of plant types offers much welfare, such as improving the quality of air, lower air temperatures, increasing soil fertility and isolating carbon dioxide. Still, several plant species have been emitting extremely volatile isoprene amount.¹³

Isoprene produces and disperses many types of wood and is one of the main reactors of tropospheric chemistry in continental regions. ¹⁴ Isoprene excretion cannot be detected in the dark, but starts faster when illuminated and causes fully stimulated genomic tissue interaction within 25 minutes. ¹⁵ All environmental dynamics (light, temperature, water stress, etc.) and the organism genetic structure have been studied. In dense vegetation areas, these volatile organic compounds direct the oxidizing atmosphere capacity which produces smog ¹⁶ and secondary organic aerosols.

There are many plants which released great amounts of isoprene. Eucalyptus, Wild Mulberry, Kachnar and Conocarpus are among the few species of plants that can release isoprene as well as monoterpenes. Monoterpenes are present in huge amount are usually kept in the tanks, while isoprene is present in free form and not in stored form, that's why it is openly released into the environment. Many people living in the semi-hilly areas of Pakistan, including Islamabad, are familiar with "Kachnar". It is from Pakistan and grows in large quantities in tropical mountainous areas. Kachnar (Bauhinia Variegate) can withstand very stressful situations. Therefore, minimal isoprene is emitted.

Eucalyptus also known as Blue Gum was brought to Pakistan from Australia a hundred years ago. This is an evergreen aromatic tree. Nowadays, it is widely cultivated in many areas of Punjab including Mansehra etc. The negative effects have recently been recognized in Pakistan. Eucalyptus spp. isoprene is one of the top plants that discharge isoprene. These plants usually grow along the coast and deal with the usual salt stress. Photosynthesis is inhibited under water pressure and dehydration, and transport traffic to carbon dioxide is gradually reduced. When performance of photosynthesis restored, then a wave of isoprene emission occurs.

Wild Mulberry (Morus Alba), a faster growing plant, is known as wild mulberry. Mulberry tree is grown in some parts of Pakistan. However, it does have some negative effects on the ecosystem. Like Eucalyptus in stressful situations, it emits isoprene. Severe drought effect carbon dynamics and plant gas exchange. Under dehydration conditions, mulberry release a large amount of VOC in the atmosphere. However, the plant gives above 11% of the same carbon in the usage of volatile compounds, especially in case of water stress.

The objective of this research was to identify and compare the isoprene emission rate in the Eucalyptus, Wild Mulberry, kachnar and Conocarpus under water stress conditions.

Materials and methods

This experimental study was directed to evaluate the emission of isoprene from different plants species (Eucalyptus, Wild Mulberry, Conocarpus, Kachnar). Six replications of each plant were taken. The isoprene emission was monitored under drought stress.

Plant selection

Plant species Eucalyptus, Conocarpous, Mulberry and Kachnar were selected for this experimental study because of their local

availability and wide abundance. The selected plants were obtained from local nurseries. Six plants from each specie have been selected for this study.

The plants of uniform height and several growth stages of leaves (12-15) were selected and then shifted them into ceramic pots (18.0cm diameter and 15.3cm deep). Pots were contained the fertile garden soil mixes with organic manure in the ecological garden.

Physiological analysis of plants

Effect of drought stress and gas exchange measurements were done to evaluate the physiological response of the plants Eucalyptus, Kachnar, Wild Mulberry and Conocarpus. Measurements of these plants were carried out on the daily basis.

Isoprene Sample Collection and analysis

Sample of Isoprene were taken from fresh fully expanded and mature leaves of all plants (Eucalyptus, Conocarpus, Wild Mulberry, Kachnar). Isoprene collection was performed three times in pot experiments (1. Full irrigated condition, 2. Under Mild stress condition when there was decrease in stomatal conductance 3. Under Severe stress when stomatal conductance reached zero). Two different experiments were performed.

Experiment (I)

The first experiment was carried out at the beginning of autumn. In this experiment all the plants (Eucalyptus, Conocarpus, Wild Mulberry, Kachnar) were well watered and a maximum amount of water was endorsed for drainage overnight. Initial and final pot weight was measured on a digital balance. Twelve plants were under water-stress and withholding water, while other 12 plants were fully irrigated up to the capacity of pot capacity which represented as control plants. Each plant was then enclosed in a plastic bag and slightly tightened with gardening wire (Figure 3.5). The sample was taken by using air tight syringe. These samples were collected in vials and mixed with methanol. Samples were collected for 1 h and 30 min. And they were analyzed by using GC-FID.

Experiment (2)

The experiment was carried out at the start of winter. For this experiment, the isoprene concentration was measured and expressed on the basis of leaf area. A leaf meter was used to measure a leaf area. The reagents are given below for this experiment:

- a. Liquid nitrogen
- b. Solid CaCl2.2H2O
- c. 100Mm EDTA (ethylenediaminetetraacetic acid)
- d. 15 mL volume glass vials and caps
- e. $10 \, \mathrm{mL}$ headspace screw cap vials and $10 \, \mathrm{mm}$ magnetic caps with silicon

After measuring the area, the 4-5 leaves per plant were separately dried in oven at 500oC and then with the addition of nitrogen they were ground to powder and consequently transferred into simple vials (15mL). Then we put the vials in water bath and incubate for 10 min at 370C. Add 1.1 g solid calcium chloride (CaCl2) and 0.5 ml EDTA (100mM, pH 7.5). Then close the vial and check vigorously. Now shake for 5 min in shaker incubator to facilitate CaCl2. Once this is done, transfer 900µl of the homogenize mixture to the bottom of 10 headspace vials. Now the sample is stable for up to 12 hours.

Isoprene analysis

The vials were analyzed by using GC-FID. The GC has been prepared by a split less injector and a HP-5MS capillary column (30m in length, 200m i.d. and 0.4m film thickness) and coupled with a mass selective detector. The oven temperature of column has been kept at 30°C for the first 1 min, after that the temperature was increased to 60°C to 150°C in the next min and finally it was sustained at 250°C for 7 min. A carrier gas He (Helium) was used for this purpose. Standards of gas at a concentration of 1 mg α - Pinene and Lamonene at a concentration of 100ppb were being used for the calibration of GC system. Then the isoprene concentration was estimated by direct comparison with the gaseous standards peak area. The identification of compound was seen by using the GC Chem Station software (Agilent). By analyzing the main fragments on spectra and parent ions, the retention time of GC peak was substantiated.

Statistical analysis

Statistical analysis (Univariate Analysis of Variance) was used for the analysis of all data set treatments. By using least significant differences (LSD) the mean values were calculated to determine the significant differences among all treatments.

Results

The water stress is the main reason of decrease in leaf conductance and in the plant biochemical procedures which shows that isoprene emission is actually an inhibitory factor for plant physiological responses. Different physiological responses a stomatal conductance, rate of photosynthesis and transpiration, an internal carbon dioxide and chlorophyll content were observed in selected plants during this experiment.

Physiological analysis of plants

The physiological responses of plants were analyzed under drought stress. The rate of photosynthesis, transpiration rate, stomatal conductance and internal carbon dioxide was similar at the middle of drought stress. A sharp decline was observed at severe stress condition.

Isoprene emission

Isoprene emission was identified under drought stress condition (Figures 1–4). Isoprene emission was identified at early, middle and final stage of drought stress period (FTSW 100%, FTSW 50% and FTSW<10%). With the increase in drought stress the isoprene emission is also increased. In this experiment the full irrigated plants released a small amount of isoprene as compared to stressed plants. Different plants showed different emission rates at different stress periods.

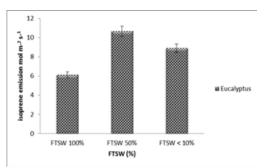


Figure 1 Isoprene emission from leaves of Eucalyptus spp. under drought stress (n=3) \pm SD. Three levels 100%, 50%, < 10 of Fractional of Transpirable Soil Water (FTSW) were made.

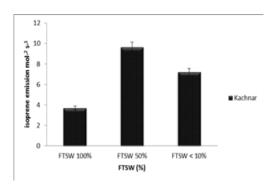


Figure 2 Isoprene emission from Kachnar (Bauhinia Variegate) under drought stress (n=3) \pm SD.

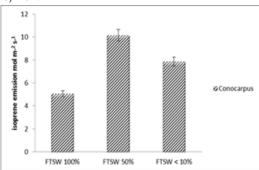


Figure 3 Isoprene emission from Conocarpus spp. under drought stress (n=3) \pm SD.

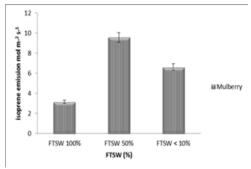


Figure 4 Isoprene emission from Mulberry (Morus Alba). under drought stress (n=3) \pm SD.

The plants released small number of isoprene in the beginning of drought stress period. In the middle of the drought stress a large number of isoprene emission rates were measured. But a slight decline at the end of this experiment when stress rate was on peak (FTSW<10) was observed. However, higher emission rate was observed in Eucalyptus spp. under drought stress period.

At FTSW 50% large number of isoprene emission was observed in all plants. At FTSW<10% the isoprene was still emitted in many stressed leaves of plants. A minor significant difference (P<0.05) was detected in all treatments. Other major and minor compounds were also identified during this experiment i.e. Limonen and α -pinene increased with decreasing FTSW.

Discussion

Water is an important factor which takes part in biochemical procedures and therefore it is necessary for growth of plant. The water stress effects the plant development and growth and brings normal plant into stress. Under drought conditions the physiological changes have been identified in plants. We found that plants produce secondary volatile compounds under drought stress.

Under drought stress conditions, the plant physiological responses were identified. Plants were observed for their responses to different stress conditions. We found that water scarcity causes a decline in leaf stomatal conductance, photosynthesis, water use efficiency and transpiration rate, which causes to low carbon assimilation and plant growth.

Stomatal conductance is indicator of water scarcity and that's why as compared to other parameters it is quickly impacted by the water shortage. ¹⁹ In our study the stomatal conductance was decreased due to increase in drought stress. The stomatal conductance rate of Eucalyptus and Kachnar was high as compared to other two plants.

Photosynthesis is important for plant yield. Photosynthesis is inhibited under drought stress.²⁰ Its rate can be affected by the biochemical limitations. Under water stress conditions the photosynthesis rate is inhibited by closing of stomata. However, due to closing of stomata the reduction takes place in internal CO2 which decreases the carbon assimilation rate.²¹ In this study the photosynthesis reduction takes place in all plants due to drought stress. But the photosynthesis rate was high and stable in all controlled plants. So, the past study showed that the closing of stomata does not limit the photosynthesis rate under stress condition.

The study also indicated that the rate of transpiration reduced due to decrease in stomatal conductance under drought stress. The Conocarpus and Mulberry showed a high transpiration rate over other plants. The transpiration is also affected by the regulation in stomatal conductance under water stress.²² The closing of stomata decrease the transpiration rate but increase the water use efficiency.

The reduction in chlorophyll content in plants has been observed under drought stress. The chlorophyll content of Kachnar and Conocarpus decreased slowly relative to other plants for the first 12 days.

Under drought situations, large amount of assimilated carbon is released in the form of BVOCs by plants. A major emitting group which is the sum of volatile emission is isoprenoid (a major emittors in plants). Isoprene acts as a defensive compound of plant against high stress situation. Earlier studies of biogenic volatile compounds revealed that the effect of drought stress on isoprene emission is directly proportional to the stress intensity. The minor rates of drought stress do not stimulate the monoterpene and isoprene emissions.

In our experimental studies, the isoprene emission rate was too low to identify in our first pot experiment during drought condition, whereas in our second pot experiment isoprene emission rate become high at middle of an experiment and uncoupled from photosynthesis. Isoprene emission rate continued to increase even when the photosynthesis rate declined. Probably in the case of un-coupling a huge amount of carbon from alternative sources subsidized in maintaining the high emission rate of isoprene as a rate of photosynthesis declined. Relation among high rate of isoprene emission and a declined in photosynthesis rate showed that the isoprene emission is less delicate to water shortage as related to photosynthesis and stomatal conductance, is unaffected during water scarcity and therefore, it becomes disengaged from rate of photosynthesis during drought stress. In the experiment, the production rate of isoprene constantly increased during drought stressed situation as a very low quantity of energy require for a leaf ontogeny and production of isoprene. Mature leaves released large amount of isoprene. This increase in the emission rate of isoprene during drought stress conditions can be recognized as the increased in

transport of electron - net assimilation rate which increased availability by decreasing the methylerythritol phosphate (MEP) corridor in other sinks of non-photosynthetic carbon reduction.

Emission rate of Isoprene declined and lastly stopped during persist high drought situations.²⁴ The emission rate of isoprene was reserved and decreased under severe water stress condition, which possibly cause a photochemistry failure in plants. The failure would be happened only during prolonged drought stress conditions over the endpoint of FTSW, at this point stomata become totally closed for the prevention of plant dehydration.^{25,26–103}

Conclusion

In the present study we concluded that in the first pot experiment (branch enclosure method) for isoprene measurement, only a small concentration of isoprene was observed that was non-significant in all treatments. In second experiment the isoprene emission was high in drought stressed plants as related to control plants, but the other VOCs compounds did not affect significantly. High isoprene emission was observed at middle of the stress and later showed decrease at the end (FTSW<10%) of experiment. The stress and later showed decrease at the end (FTSW<10%) of experiment.

Acknowledgments

None.

Funding

None.

Conflicts of interest

Authors declare that there is no conflict of interest.

References

- Solomon S, Qin D, Manning M, et al. Climate change 2007–the physical science basis: Working group I contribution to the fourth assessment report of the IPCC (Vol. 4). Cambridge University Press; 2007.
- Jentsch A, Kreyling J, Beierkuhnlein C. A new generation of climatechange experiments: events, not trends. Frontiers in Ecology and the Environment. 2007;5(7):365–374.
- Iqbal J, Ronggui H, Lijun D, et al. Differences in soil CO₂ flux between different land use types in mid–subtropical China. Soil Biology and Biochemistry. 2008;40(9):2324–2333.
- Centritto M, Lauteri M, Monteverdi MC, et al. Leaf gas exchange, carbon isotope discrimination, and grain yield in contrasting rice genotypes subjected to water deficits during the reproductive stage. *Journal of Experimental Botany*. 2009;60(8):2325–2339.
- Brauman KA, Siebert S, Foley JA. Improvements in crop water productivity increase water sustainability and food security—a global analysis. *Environmental Research Letters*. 2013;8(2):024030.
- Weltzin JF, Loik ME, Schwinning S, et al. Assessing the response of terrestrial ecosystems to potential changes in precipitation. *Bioscience*. 2003;53(10):941–952.
- Peñuelas J, Llusià J. BVOCs: plant defense against climate warming? Trends in Plant Science. 2003;8(3):105–109.
- Šimpraga M, Verbeeck H, Demarcke M, et al. Comparing monoterpenoid emissions and net photosynthesis of beech (Fagus Sylvatica L.) in controlled and natural conditions. *Atmospheric Environment*. 2011;45(17):2922–2928.
- 9. Loreto F, Schnitzler JP. Abiotic stresses and induced BVOCs. *Trends in Plant Science*. 2010;15(3):154–166.

- Zemankova K, Brechler J. Emissions of biogenic VOC from forest ecosystems in central Europe: Estimation and comparison with anthropogenic emission inventory. *Environmental Pollution*. 2010;158(2):462–469.
- Kreuzwieser J, Rennenberg H, Steinbrecher R. Impact of short–term and long–term elevated CO₂ on emission of carbonyls from adult Quercus petraea and Carpinus betulus trees. *Environmental Pollution*. 2006;142(2):246–253.
- Kesselmeier J, Staudt M. Biogenic volatile organic compounds (VOC): an overview on emission, physiology and ecology. *Journal of Atmospheric Chemistry*. 1999;33(1):23–88.
- 13. Tingey DT, Manning M, Grothaus LC, et al. The influence of light and temperature on isoprene emission rates from live oak. *Physiologia Plantarum*. 1979;47(2):112–118.
- Fehsenfeld F, Calvert J, Fall R, et al. Emissions of volatile organic compounds from vegetation and the implications for atmospheric chemistry. Global Biogeochemical Cycles. 1992;6(4):389–430.
- Monson RK, Hills AJ, Zimmerman PR, et al. Studies of the relationship between isoprene emission rate and CO2 or photon-flux density using a real-time isoprene analyser. *Plant, Cell & Environment*. 1991;14(5):517– 523
- Simpson D, Guenther A, Hewitt CN, et al. Biogenic emissions in Europe:
 Estimates and uncertainties. *Journal of Geophysical Research: Atmospheres*. 1995;100(D11):22875–22890.
- Centritto M, Tognetti R, Leitgeb E, et al. Above ground processes: anticipating climate change influences. *IForest Management and the Water Cycle*. Springer, Dordrecht; 2010:31–64.
- Wilkinson S, Davies WJ. Drought, ozone, ABA and ethylene: new insights from cell to plant to community. *Plant, Cell & Environment*. 2010;33(4):510–525.
- Chartzoulakis K, Patakas A, Kofidis G, et al. Water stress affects leaf anatomy, gas exchange, water relations and growth of two avocado cultivars. Scientia Horticulturae. 2002;95(1–2):39–50.
- Misra AN, Biswal AK, Misra M. Physiological, biochemical and molecular aspects of water stress responses in plants, and the biotechnological applications. *Proceedings–National Academy of Sciences India Section B*. 2002;72(2):115–134.
- Centritto M, Loreto F, Chartzoulakis K. The use of low [CO2] to estimate diffusional and non-diffusional limitations of photosynthetic capacity of salt-stressed olive saplings. *Plant, Cell & Environment*. 2003;26(4):585–594.
- Wahbi S, Wakrim R, Aganchich B, et al. Effects of partial root zone drying (PRD) on adult olive tree (Olea europaea) in field conditions under arid climate: I. Physiological and agronomic responses. *Agriculture, Ecosystems & Environment*. 2005;106(2–3):289–301.
- 23. Niinemets Ü, Arneth A, Kuhn U, et al. The emission factor of volatile isoprenoids: stress, acclimation, and developmental responses. *Biogeosciences*. 2010;7(7):2203–2223.
- Potosnak MJ, LeStourgeon L, Nunez O. Increasing leaf temperature reduces the suppression of isoprene emission by elevated CO₂ concentration. Science of the Total Environment. 2014;481:352–359.
- Brilli F, Barta C, Fortunati A, et al. Response of isoprene emission and carbon metabolism to drought in white poplar (Populus Alba) saplings. New Phytologist. 2007;175(2):244–254.
- Akıncı Ş, Lösel DM. Plant water–stress response mechanisms. Water Stress. 2012:15–42.
- 27. Barathi P, Sundar D, Reddy AR. Changes in mulberry leaf metabolism in response to water stress. *Biologia Plantarum*. 2001;44(1):83–87.

- Barnabás B, Jäger K, Fehér A. The effect of drought and heat stress on reproductive processes in cereals. *Plant, Cell & Environment*. 2008;31(1):11–38.
- Blanch JS, Penuelas J, Sardans J, et al. Drought, warming and soil fertilization effects on leaf volatile terpene concentrations in Pinus halepensis and Quercus ilex. Acta Physiologiae Plantarum. 2009;31(1):207.
- Calfapietra C, Wiberley AE, Falbel TG, et al. Isoprene synthase expression and protein levels are reduced under elevated O3 but not under elevated CO₂ (FACE) in field-grown aspen trees. *Plant, Cell & Environment*. 2007;30(5):654–661.
- Campos H, Trejo C, Peña–Valdivia CB, et al. Stomatal and non–stomatal limitations of bell pepper (Capsicum Annuum L.) plants under water stress and rewatering: Delayed restoration of photosynthesis during recovery. *Environmental and Experimental Botany*. 2014;98:56–64.
- 32. Carter WP. Development of ozone reactivity scales for volatile organic compounds. *Air & waste*. 1994;44(7):881–899.
- 33. Centritto M, Loreto F. Photosynthesis in a changing world: photosynthesis and abiotic stresses. *Agriculture, Ecosystems & Environment.* 2005;106:2–3.
- Chaves MM, Flexas J, Pinheiro C. Photosynthesis under drought and salt stress: regulation mechanisms from whole plant to cell. *Annals of Botany*. 2009;103(4):551–560.
- Chaves MM, Maroco JP, Pereira JS. Understanding plant responses to drought—from genes to the whole plant. Functional Plant Biology. 2003;30(3):239–264.
- Chen LIN, Dodd IC, Davies WJ, et al. Ethylene limits abscisic acid-or soil drying-induced stomatal closure in aged wheat leaves. *Plant, Cell & Environment*. 2013;36(10):1850–1859.
- Chen L, Dodd IC, Theobald JC, et al. The rhizobacterium Variovorax paradoxes 5C–2, containing ACC deaminase, promotes growth and development of Arabidopsis Thaliana via an ethylene–dependent pathway. *Journal of Experimental Botany*. 2013;64(6):1565–1573.
- Cinege G, Louis S, Hänsch R, et al. Regulation of isoprene synthase promoter by environmental and internal factors. *Plant Molecular Biology*. 2008;69(5):593–604.
- Dani KS, Jamie IM, Prentice IC, et al. Evolution of isoprene emission capacity in plants. *Trends in Plant Science*. 2014;19(7):439–446.
- Delfine S, Loreto F, Pinelli P, et al. Isoprenoids content and photosynthetic limitations in rosemary and spearmint plants under water stress. Agriculture, Ecosystems & Environment. 2005;106(2–3):243–252.
- 41. Dicke M, Loreto F. Induced plant volatiles: from genes to climate change. *Trends in Plant Science*. 2010;15(3):115–117.
- 42. Dodd IC, Whalley WR, Ober ES, et al. Genetic and management approaches to boost UK wheat yields by ameliorating water deficits. *Journal of Experimental Botany*. 2011;62(15):5241–5248.
- 43. Dume B, Mosissa T, Nebiyu A. Effect of biochar on soil properties and lead (Pb) availability in a military camp in South West Ethiopia. *African Journal of Environmental Science and Technology*. 2016;10(3):77–85.
- 44. Ercisli S. A short review of the fruit germplasm resources of Turkey. *Genetic Resources and Crop Evolution*. 2004;51(4):419–435.
- Evans RC, Tingey DT, Gumpertz ML, et al. Estimates of isoprene and monoterpene emission rates in plants. *Botanical Gazette*. 1982;143(3):304–310.
- Fares S, Gentner DR, Park JH, et al. Biogenic emissions from Citrus species in California. *Atmospheric Environment*. 2011;45(27):4557– 4568.

- Flexas J, Barón M, Bota J, et al. Photosynthesis limitations during water stress acclimation and recovery in the drought-adapted Vitis hybrid Richter-110 (V. berlandieri× V. rupestris). *Journal of Experimental Botany*. 2009;60(8):2361–2377.
- 48. Flexas J, Bota J, Cifre J, et al. Understanding down-regulation of photosynthesis under water stress: future prospects and searching for physiological tools for irrigation management. *Annals of Applied Biology*. 2004;144(3):273–283.
- Fortunati A, Barta C, Brilli F, et al. Isoprene emission is not temperaturedependent during and after severe drought-stress: a physiological and biochemical analysis. *The Plant Journal*. 2008;55(4):687–697.
- Fuentes JD, Wang D. On the seasonality of isoprene emissions from a mixed temperate forest. *Ecological Applications*. 1999;9(4):1118–1131.
- Grassi G, Magnani F. Stomatal, mesophyll conductance and biochemical limitations to photosynthesis as affected by drought and leaf ontogeny in ash and oak trees. *Plant, Cell & Environment*. 2005;28(7):834–849.
- Guenther A, Karl T, Harley P, et al. Estimates of global terrestrial isoprene emissions using MEGAN (Model of Emissions of Gases and Aerosols from Nature). Atmospheric Chemistry and Physics. 2006;6(11):3181–3210.
- Hui D, Luo Y, Cheng W, et al. Canopy radiation-and water-use efficiencies as affected by elevated [CO₂]. Global Change Biology. 2001;7(1):75–91.
- Kirstine W, Galbally I, Ye Y, et al. Emissions of volatile organic compounds (primarily oxygenated species) from pasture. *Journal of Geophysical Research: Atmospheres*. 1998;103:10605–10619.
- Laothawornkitkul J, Paul ND, Vickers CE, et al. Isoprene emissions influence herbivore feeding decisions. *Plant, Cell & Environment*. 2008;31(10):1410–1415.
- Lawlor DW, Tezara W. Causes of decreased photosynthetic rate and metabolic capacity in water-deficient leaf cells: a critical evaluation of mechanisms and integration of processes. *Annals of Botany*. 2009;103(4):561–579.
- Lenton TM, Huntingford C. Global terrestrial carbon storage and uncertainties in its temperature sensitivity examined with a simple model. Global Change Biology. 2003;9(10):1333–1352.
- Llusià J, Peñuelas J. Changes in terpene content and emission in potted Mediterranean woody plants under severe drought. *Canadian Journal of Botany*. 1998;76(8):1366–1373.
- Loivamäki M, Louis S, Cinege G, et al.. Circadian rhythms of isoprene biosynthesis in grey poplar leaves. *Plant Physiology*. 2007;143(1):540– 551.
- Loreto F, Centritto M. Leaf carbon assimilation in a water-limited world. *Plant Biosystems*. 2008;142(1):154–161.
- 61. Loreto F, Delfine S. Emission of isoprene from salt–stressed Eucalyptus globulus leaves. *Plant Physiology.* 2000;123(4):1605–1610.
- Loreto F, Velikova V. Isoprene produced by leaves protects the photosynthetic apparatus against ozone damage, quenches ozone products, and reduces lipid peroxidation of cellular membranes. *Plant Physiology*. 2001;127(4):1781–1787.
- 63. Masinde PW, Stützel H, Agong SG, et al. Plant growth, water relations and transpiration of two species of African nightshade (Solanum villosum Mill. ssp. miniatum (Bernh. ex Willd.) Edmonds and S. sarrachoides Sendtn.) under water-limited conditions. *Scientia Horticulturae*. 2006;110(1):7–15.
- Mayrhofer S, Teuber M, Zimmer I, et al. Diurnal and seasonal variation of isoprene biosynthesis-related genes in grey poplar leaves. *Plant Physiology*. 2005;139(1):474

 –484.

- McGarvey DJ, Croteau R. Terpenoid metabolism. The Plant Cell. 1995;7(7):1015.
- Miller B, Oschinski C, Zimmer W. First isolation of an isoprene synthase gene from poplar and successful expression of the gene in Escherichia coli. *Planta*. 2001;213(3):483–487.
- Monson RK, Holland EA. Biospheric trace gas fluxes and their control over tropospheric chemistry. *Annual Review of Ecology and Systematics*. 2001;32(1):547–576.
- Monteiro JA, Nell TA, Barrett JE. Postproduction of potted miniature rose: Flower respiration and single flower longevity. *Journal of the American Society for Horticultural Science*. 2001;126(1):134–139.
- Muhammad S, Müller T, Joergensen RG. Relationships between soil biological and other soil properties in saline and alkaline arable soils from the Pakistani Punjab. *Journal of Arid Environments*. 2008;72(4):448–457.
- Ormeno E, Mevy JP, Vila B, et al. Water deficit stress induces different monoterpene and sesquiterpene emission changes in Mediterranean species. Relationship between terpene emissions and plant water potential. *Chemosphere*. 2007;67(2):276–284.
- 71. Pichersky E, Gershenzon J. The formation and function of plant volatiles: perfumes for pollinator attraction and defense. *Current Opinion in Plant Biology.* 2002;5(3):237–243.
- 72. Rayment GE, Higginson FR. Australian laboratory handbook of soil and water chemical methods. Inkata Press Pty Ltd. 1992.
- Sanadze GA. Biogenic isoprene (a review). Russian Journal of Plant Physiology. 2004;51(6):729–741.
- Sánchez–Blanco MJ, Ferrández T, Morales MA, et al. Variations in water status, gas exchange, and growth in Rosmarinus officinalis plants infected with Glomus deserticola under drought conditions. *Journal of Plant Physiology*. 2004;161(6):675–682.
- Sasaki K, Ohara K, Yazaki K. Gene expression and characterization of isoprene synthase from Populus alba. FEBS letters. 2005;579(11):2514– 2518.
- Schnitzler JP, Zimmer I, Bachl A, et al. Biochemical properties of isoprene synthase in poplar (Populus× canescens). *Planta*. 2005;222(5):777–786.
- Shao HB, Chu LY, Jaleel CA, et al. Water-deficit stress-induced anatomical changes in higher plants. *Comptes Rendus Biologies*. 2008;331(3):215–225.
- 78. Sharkey TD, Chen X, Yeh S. Isoprene increases thermo tolerance of fosmidomycin–fed leaves. *Plant Physiology*. 2001;125(4).
- 79. Sharkey TD, Wiberley AE, Donohue AR. Isoprene emission from plants: why and how. *Annals of Botany*. 2008;101(1):5–18.
- Sharkey TD, Yeh S, Wiberley AE, et al. Evolution of the isoprene biosynthetic pathway in kudzu. *Plant Physiology*. 2005;137(2):700–712.
- Sharma N, Waterer DR, Abrams SR. Evaluation of abscisic acid analogs as holding agents for bedding plant seedlings. *HortTechnology*. 2006;16(1):71-77.
- Silver GM, Fall R. Enzymatic synthesis of isoprene from dimethylallyl diphosphate in aspen leaf extracts. *Plant Physiology*. 1991;97(4):1588– 1591
- Silver GM, Fall R. Characterization of aspen isoprene synthase, an enzyme responsible for leaf isoprene emission to the atmosphere. *Journal of Biological Chemistry*. 1995;270(22):13010–13016.
- 84. Sindelarova K, Granier C, Bouarar I, et al. Global data set of biogenic VOC emissions calculated by the MEGAN model over the last 30 years. *Atmospheric Chemistry and Physics*. 2014;14(17):9317–9341.

- 85. Street RA, Hewitt CN, Mennicken S. Isoprene and monoterpene emissions from a Eucalyptus plantation in Portugal. Journal of Geophysical Research: Atmospheres. 1997;102:15875–15887.
- 86. Sumanta N, Haque CI, Nishika J, et al. Spectrophotometric analysis of chlorophylls and carotenoids from commonly grown fern species by using various extracting solvents. Research Journal of Chemical Sciences. 2014.
- 87. Thomas DS, Loreto F. Water stress, temperature, and light effects on the capacity for isoprene emission and photosynthesis of kudzu leaves. Oecologia. 1993;95(3):328-333.
- 88. Vickers CE, Possell M, Cojocariu CI, et al. Isoprene synthesis protects transgenic tobacco plants from oxidative stress. Plant, Cell & Environment. 2009;32(5):520-531.
- 89. Vickers CE, Possell M, Hewitt CN, et al. Genetic structure and regulation of isoprene synthase in Poplar (Populus spp.). Plant Molecular Biology. 2010;73(4-5):547-558.
- 90. Vuorinen T, Reddy GVP, Nerg AM, et al. Monoterpene and herbivoreinduced emissions from cabbage plants grown at elevated atmospheric CO2 concentration. Atmospheric Environment. 2004;38(5):675-682.
- 91. Walkley A. A critical examination of a rapid method for determining organic carbon in soils-effect of variations in digestion conditions and of inorganic soil constituents. Soil Science. 1947;63(4):251-264.
- 92. Wiberley AE, Donohue AR, Meier ME, et al. Regulation of isoprene emission in Populus trichocarpa leaves subjected to changing growth temperature. Plant, Cell & Environment. 2008;31(2):258-267.
- 93. Wiberley AE, Donohue AR, Westphal MM, et al. Regulation of isoprene emission from poplar leaves throughout a day. Plant, Cell & Environment. 2009;32(7):939-947.
- 94. Wiberley AE, Linskey AR, Falbel TG, et al. Development of the capacity for isoprene emission in kudzu. Plant, Cell & Environment. 2005;28(7):898–905.

- 95. Wildermuth MC, Fall R. Light-dependent isoprene emission (characterization of a thylakoid-bound isoprene synthase in Salix discolor chloroplasts). Plant Physiology. 1996;112(1):171-182.
- 96. Wildermuth MC, Fall R. Biochemical characterization of stromal and thylakoid-bound isoforms of isoprene synthase in willow leaves. Plant Physiology. 1998;116(3):1111-1123.
- 97. Wilkinson MJ, Owen SM, Possell M, et al. Circadian control of isoprene emissions from oil palm (Elaeis guineensis). The Plant Journal. 2006;47(6):960-968.
- 98. Wilkinson S, Davies WJ. ABA-based chemical signaling: the coordination of responses to stress in plants. Plant, Cell & Environment. 2002;25(2):195-210.
- 99. Wilkinson S, Kudoyarova GR, Veselov DS, et al. Plant hormone interactions: innovative targets for crop breeding and management. Journal of Experimental Botany. 2012;63(9):3499-3509.
- 100. Winters AJ, Adams MA, Bleby TM, et al. Emissions of isoprene, monoterpene and shortchained carbonyl compounds from Eucalyptus spp. in southern Australia. Atmospheric Environment. 2009;43(19):3035-3043.
- 101. Wong SC, Cowan IR, Farquhar GD. Stomatal conductance correlates with photosynthetic capacity. Nature. 1979;282(5737):424.
- 102. Yordanov I, Velikova V, Tsonev T. Plant responses to drought and stress tolerance. Bulgarian Journal of Plant Physiology. 2003:187-206.
- 103. Zhang M, Duan L, Zhai Z, et al. Effects of plant growth regulators on water deficit-induced yield loss in soybean. In Proceedings of the 4th International Crop Science Congress, Brisbane, Australia; 2004:252-