

# Taxonomic and nitrogen-cycling microbial community functional profiles of sugarcane and adjacent forest soils in Southeast Brazil

## Abstract

Nowadays, due to the expansion of agricultural borders, it is highly desirable to increase the sustained productivity of sugarcane cultivars using the knowledge of soil microbial communities. In this study, twelve shotgun metagenomic datasets based on genomic DNA from soil were analyzed using the Metagenomics Rapid Annotation using Subsystem Technology (MG-RAST) and Statistical Analysis of Metagenomic Profiles (STAMP) to assess differential responses for the total soil bacterial community composition and nitrogen-cycling microbial community functional potential in soils from sugarcane field with pre-harvest burning and adjacent forest in dry and wet seasons in Southeast Brazil. The soil bacterial community revealed higher abundance for Actinobacteria in forest soil than sugarcane soil in dry and wet seasons, and an opposite pattern for Proteobacteria and Planctomycetes in these soils in both seasons. The results obtained in this study based on the KEEG map suggest that the forest soil has a higher nitrogen-cycling microbial community functional potential compared to the sugarcane soil, independently of the season. The gene sequences associated with carbohydrate metabolism were the most frequent in all soil metagenomes. Taken together, the results confirm previous findings regarding the effects of forest conversion to sugarcane production area, providing new insights regarding to this conversion through the prism of the seasonality and pre-harvesting method on microbially mediated nitrogen cycle in sugarcane production fields.

**Keywords:** soil bacterial community composition, shotgun metagenomic sequencing, nitrogen cycle, seasonality, sustainability

Volume 6 Issue 4 - 2021

Eliamar Aparecida Nascimbém Pedrinho,<sup>1</sup>  
Acacio Aparecido Navarrete,<sup>2</sup> Luciano  
Takeshi Kishi,<sup>1</sup> Camila Cesário Fernandes,<sup>1</sup>  
Victoria Romancini Toledo,<sup>2</sup> Rita de Cassia  
Félix Alvarez,<sup>2</sup> Elisângela de Souza Loureiro,<sup>2</sup>  
Leandro Nascimento Lemos,<sup>3</sup> Siu Mui Tsai,<sup>4</sup>  
Eliana Gertrudes de Macedo Lemos<sup>1</sup>

<sup>1</sup>São Paulo State University, Technology Department, Jaboticabal, São Paulo, Brazil

<sup>2</sup>Federal University of Mato Grosso do Sul, Chapadão do Sul, Mato Grosso do Sul, Brazil

<sup>3</sup>National Laboratory of Scientific Computing, Petrópolis, Rio de Janeiro, Brazil

<sup>4</sup>Center for Nuclear Energy in Agriculture, University of São Paulo, Piracicaba, São Paulo, Brazil

**Correspondence:** Acacio Aparecido Navarrete, Federal University of Mato Grosso do Sul, Rod. MS-306, km 105, Chapadão do Sul, MS, Brazil, Zip Code 79560-000, Tel +55-67-3562-6352, Email [acacionavarrete@gmail.com](mailto:acacionavarrete@gmail.com)

**Received:** June 24, 2021 | **Published:** July 05, 2021

## Introduction

Brazil is a major sugarcane producer and Southeast region cultivates 5.3 million hectares, close to 50% of Brazil's sugarcane area.<sup>1</sup> The rapid increase in production has brought into question the sustainability of biofuels, especially considering the sustainable use of the soil associated to its ecosystem services. The continuous cropping of sugarcane can result in the disruption of the soil microbial community and cause significant declines in yields.<sup>2</sup> Pre-harvesting burning of sugarcane straw is an agricultural practice that, despite the state legislation recommending it to be completely phased out in areas where mechanized harvesting is possible, it is usual in non-mechanized areas.<sup>3</sup> Despite the previous studies focusing the soil microbiota in sugarcane burnt soils,<sup>4-7</sup> there are a lack of conclusive data about the impact of this harvest method on the tropical soil ecosystem sustainability, mainly regarding to the microbe-mediated processes in soil.<sup>8</sup> Determination of the effect of agricultural management practices on soil sustainability is a challenge, particularly the aspect concerning the evaluation of the biological quality of the soil.<sup>9</sup>

A widely accepted definition of soil quality is the capacity of a soil to function within ecosystems and land-use boundaries to sustain biological productivity, to maintain environmental quality, and to promote plant, animal, and human health.<sup>10</sup> Taxonomic and functional changes in the microbial community of soil can reflect changes in the soil quality.<sup>11</sup> Soil microorganisms are involved in ecological functions that are essential to the environment, such as decomposition of plant and animal residues, solubility of nutrients, formation of organic matter, soil structure, plant growth promoters, and biological control

agents. Thus, microbial processes are fundamentally important to the functioning of agro-ecological production systems, which perform functions directly related to their productivity and sustainability.<sup>12</sup>

The shotgun metagenomic sequencing and bioinformatic analysis are important tools to quantify the abundance and distribution of microbial taxa at the community ecological level, gene families involved in soil microbial functions and nutrient metabolism below-ground.<sup>13</sup> The size, composition, and function of these microbial communities are affected by land-use,<sup>14,15</sup> seasonality,<sup>16</sup> agricultural practices,<sup>17,18</sup> and other environmental changes. De Caceres, Legendre, and Moretti<sup>19</sup> consider that differential abundances in microbial taxa and functional genes can be used as potential bioindicators for environmental change. However, our knowledge on the soil microbial communities, and below-ground nitrogen metabolism in tropical soils under agricultural and forestry land-uses is limited and requires further elucidation of this ecological aspects related to soil quality concerning to vegetation cover, seasonality and agricultural practices.

In this study, we evaluated the seasonal patterns in taxonomic and functional profiles of the soil microbial communities in sugarcane and adjacent native forest areas located in the Southeast Brazil. We evaluated the following hypothesis: (1) forest soil presents higher abundance of the most bacterial phyla in wet and dry seasons in comparison with sugarcane soil, and (2) higher nitrogen-cycling microbial community functional potential is found in forest soil than in sugarcane soil in wet and dry seasons. To evaluate these expectations, we used whole genome shotgun metagenomic sequencing of DNA obtained from soil samples to generate raw data for subsequent bacterial profiling and functional analyses.

## Material and methods

### Sampling sites and soil sample collection

Two different field sites were chosen in Southeastern Brazil, a burnt sugarcane (SUG) harvesting site located in a private property (Fazenda Coqueiro) (21°42'30.67"S 47°34'56.89"W) and a native forest (FOR) site located in the Vassununga State Park (21°42'34.62"S 47°34'30.04"W). Both the sites are located in Santa Rita do Passa Quatro municipality in the state of São Paulo, Brazil. The climate in the region is classified as Cwa (Köppen's classification), with an annual precipitation varying from 1,300 to 1,700mm, and total precipitation of 18.6mm in the driest month (August). The average annual temperature in the region is 23.3°C, with an average maximum temperature (December to February) of 26°C and a minimum of 19.50°C (June to August).

Soil samples were collected from three different points at least 14m apart from each other along a transect at each sampling site in both the dry (D) (February 2011) and wet (W) (November 2011) seasons. Undeformed soil cores were taken from the 0- to 20-cm top-soil layer after the removal of the litter layer using a 5-cm diameter aseptic cylindrical corer. A total of 12 soil samples were collected from the field (two sampling sites × three sampling points × two sampling periods). The samples were transported to the laboratory in a cool box containing ice, stored at -20°C, and processed within 72h of sampling.

### Isolation of DNA from soil and high-throughput sequencing of soil metagenomes

Soil DNA was extracted from 0.5g (wet weight) of 24 soil samples using the FastDNA™ SPIN Kit for Soil (MP Biomedicals, Santa Ana, CA, USA) and the FastPrep® equipment (MP Biomedicals, Santa Ana, CA, USA) according to the manufacturer's instructions. Both purity and quality of the genomic DNA were assessed via spectrophotometry on a NanoDrop apparatus (NanoDrop ND-1000 NanoDrop Technologies, Inc., Wilmington, DE, USA) to determine the absorbance at the following wavelengths: 230, 260, 280, and 320nm. The DNA concentration was determined using a Qubit 2.0 fluorometer (Life technologies, Carlsbad, CA, USA). DNA was stored at -20°C until further use.

Twelve DNA sequencing libraries (three FOR-D libraries, three FOR-W libraries, three SUG-D libraries, and three SUG-W libraries) were prepared using the Illumina Nextera sample preparation kit (Illumina, San Diego, CA, USA) according to the manufacturer's instructions. The libraries were evaluated on the 2100 Bioanalyzer using a High Sensitivity DNA kit (Agilent, Santa Clara, CA, USA) to estimate the library size. The libraries were quantified using the Qubit dsDNA HS kit on a Qubit 2.0 fluorometer (Life Technologies, Carlsbad, CA, USA) and quantitative PCR (qPCR) was performed using the KAPA SYBR FAST qPCR Master mix, Illumina standards, and primer premix (KAPA Biosystems, Wilmington, MA, USA) according to the instructions provided by Illumina. The resulting DNA libraries were denatured with NaOH, diluted to 8pM using Illumina's HT1 buffer, and spiked with 1% PhiX. Equal concentrations of the libraries were loaded on the Illumina's HiScanSQ System (Illumina, San Diego, CA, USA) operated in a Rapid Run Mode to generate 2×100bp paired-end reads. After sequencing, the libraries were separated, and the BCL files were converted into FASTQ format using the Casava 1.8.3 software (Illumina, San Diego, CA, USA). The raw data of the sequencing were deposited in the National Center for Biotechnology Information (NCBI) server (<https://www.ncbi.nlm.nih.gov>) under accession number PRJNA665014 'Taxonomic and functional microbial profiles of sugarcane and forest soils in the tropical region'.

### Shotgun metagenomic data processing and taxonomic identification of bacterial groups

Statistical analysis was conducted using the Statistical Analysis of Metagenomic Profiles (STAMP).<sup>20</sup> The significance of the relative proportion difference in the taxonomic distribution of metagenomic samples was performed using the two-sided Fisher's exact test, with the Newcombe–Wilson confidence interval method. Since the *P*-values were not uniformly distributed using the Storey's false discovery rate (FDR), the Benjamini–Hochberg FDR was used for the correction.

### Nitrogen-associated genes and -metabolic pathway analysis

Nitrogen-associated cycle genes were identified using the SEED subsystems annotation source of MG-RAST, with a maximum e-value of 1e<sup>05</sup>, minimum identity of 60%, and minimum alignment length of 15. Subsets of the nitrogen-associated cycle genes in a functional subsystem for sequences belonging to *Bacteria* domain were generated using the workbench tool from the MG-RAST server. The nitrogen metabolic pathway analysis was performed using KEGG mapper from the MG-RAST server, with a maximum e-value of 1e<sup>05</sup>, minimum identity of 60%, and minimal alignment length of 15.

A functional analysis using the SEED<sup>21</sup> and KEGG<sup>22</sup> databases was performed using the STAMP software. Each sequence was related to its SEED functional role using the best BLAST score to protein sequences without known functional roles. A similar procedure was used to match each sequence to a KEGG orthology (KO) accession number.

## Results and discussion

### Metagenomic profiles

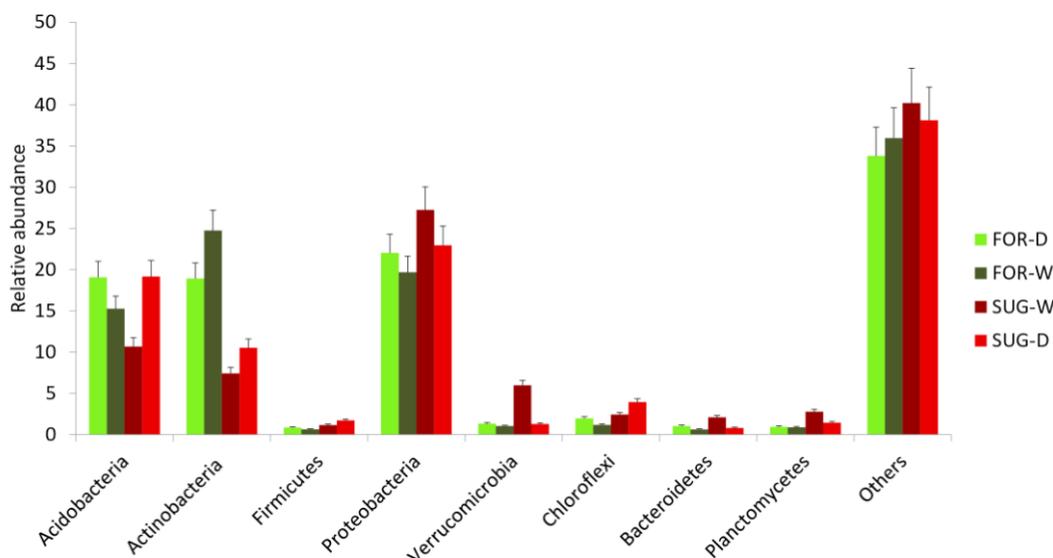
The twelve libraries yielded over 15,000 reads. The readings identified as artificial duplicates were removed using MG-RAST. After quality control, an average of 76,607, 74,426, 63,005, and 166,676 proteins were predicted for FOR-D, FOR-W, SUG-W, and SUG-D, respectively, and 71.7%, 73.9%, 73.1%, and 78.5% of the total reads were annotated as functionally assigned proteins. The taxa richness estimated for FOR-D, FOR-W, SUG-W, and SUG-D revealed the following values: 284.599, 252.993, 381.042, and 385.731, respectively.

### Bacterial community composition in sugarcane and forest soils in both dry and wet seasons

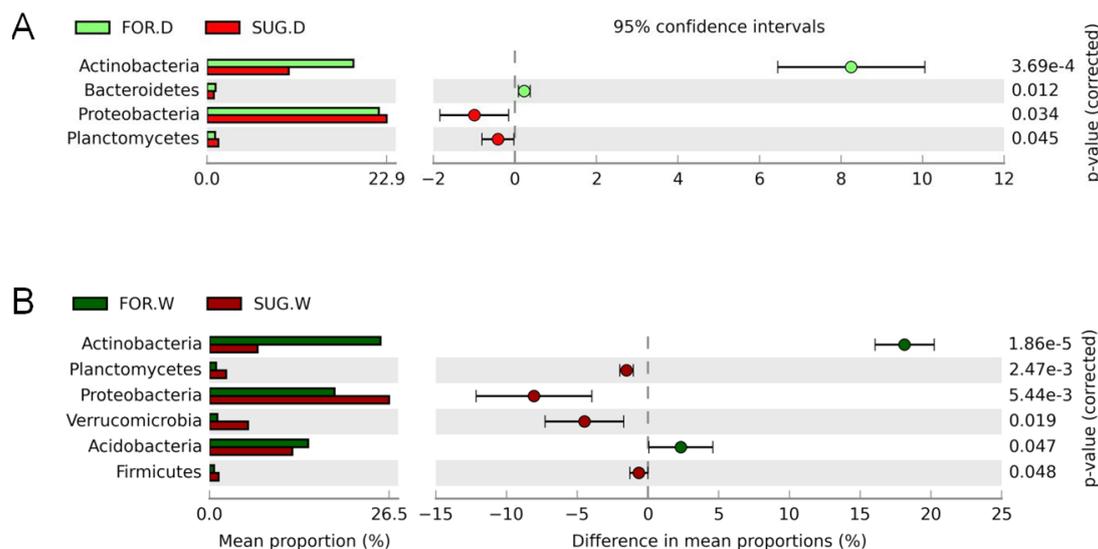
Proteobacteria, Acidobacteria, and Actinobacteria were the most abundant bacterial phyla found in all soil metagenomes (Figure 1). Higher abundance was showed for Actinobacteria in FOR-D and FOR-W than SUG-D and SUG-W, and an opposite pattern was revealed for Proteobacteria in these soils in both seasons (Figure

2a and 2b). These results corroborate findings of previous studies, which also reported that these phyla were the most abundant in soil samples.<sup>23–28</sup> The predominance of Proteobacteria in such environments can be explained by the fact that this phylum comprises

the most phylogenetically diverse group in the *Bacteria* domain, known to be metabolically versatile, including aerobic and facultative bacteria.<sup>29</sup>



**Figure 1** Relative abundance of the most abundant bacterial phyla as revealed by metagenomic sequencing and measured by percentage of total *Bacteria*.



**Figure 2** Differences in abundance of bacterial phyla revealed using the Statistical Analysis of Metagenomic Profiles (STAMP) software based on abundance of metagenomic sequences. (a) Differences between soil samples from native forest (FOR-D) and sugarcane (SUG-D) both collected during the dry season. (b) Differences between soil samples from native forest (FOR-W) and sugarcane (SUG-W) both collected during the wet season.

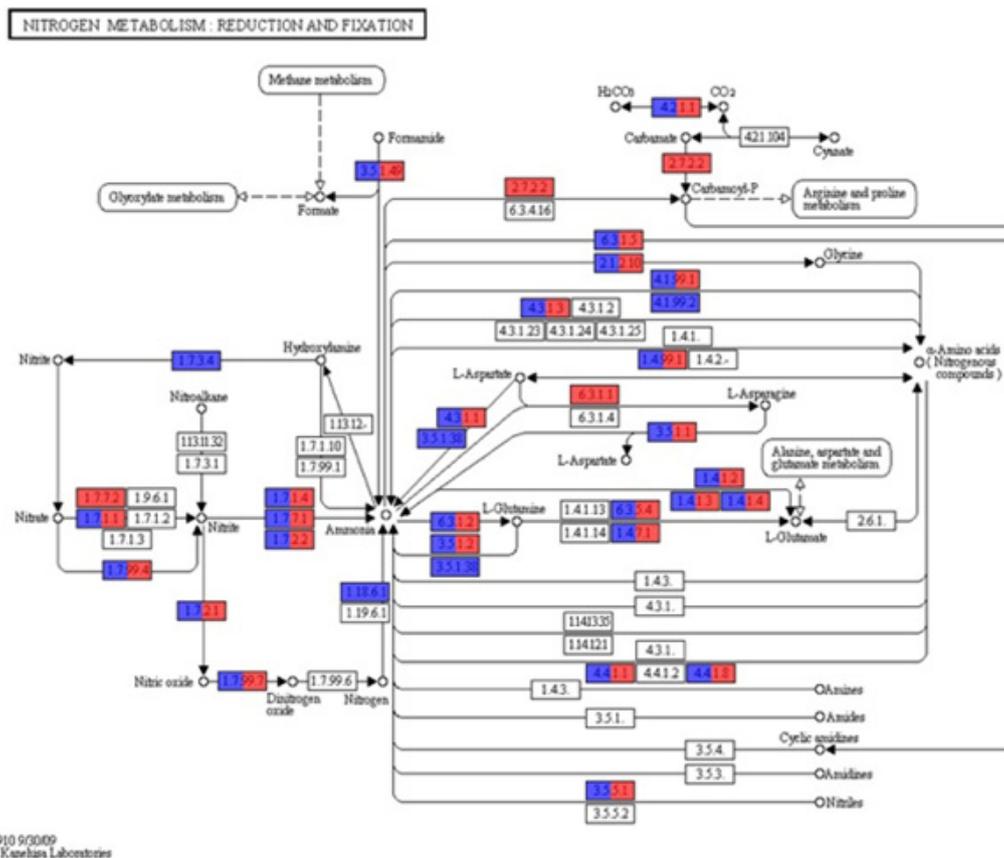
The prevalence of Acidobacteria ranged from 2.4% to 78.5% in different soil samples. Acidobacteria form one of the main groups found in tropical soils, with prevalence in tropical forest soil.<sup>14,17</sup> This corroborates the results found in FOR-D (26.8%) and FOR-W (26.2%), in contrast to the SUG-W sample, where only 13.5% of the sequences corresponded to the phylum Acidobacteria. According to Ward et al.<sup>30</sup> and de Chaves et al.,<sup>13</sup> the occurrence of Acidobacteria in soil may indicate the presence of genes related to the synthesis of cellulose and proteins, and carbon degradation, suggesting characteristics of resistance to desiccation and metabolic potential for C degradation in sugarcane soil.

The relative abundance of Actinobacteria was lower in sugarcane soil, SUG-D (11.8%) and SUG-W (5.9%), while it was higher in native

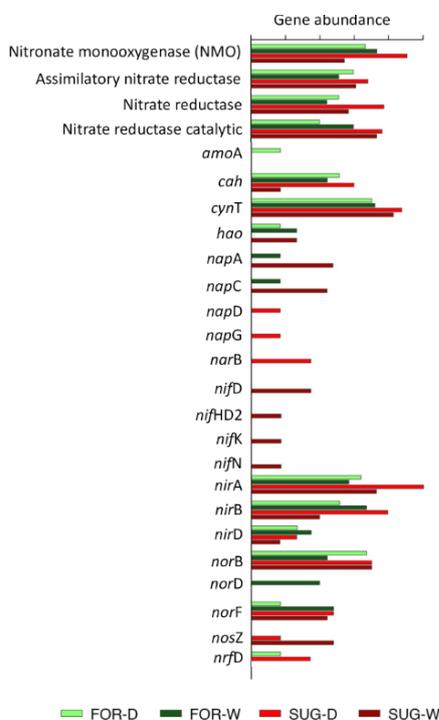
forest soils, FOR-D (18.3%) and FOR-W (27.1%) (Figure 2a and 2b). The data also showed that Actinobacteria was the dominant phylum in the native forest samples in dry and wet seasons. Russo et al.<sup>31</sup> have reported that Actinobacteria is a dominant class in sandy soil, and moisture is a determining factor for abundance of this phylum, which increases in arid soil due to the resistance of several species to drought stress.<sup>32,33</sup>

Other bacterial phyla with differential responses in abundance regarding to sugarcane and forest soils in the dry and wet seasons were Bacteroidetes, Planctomycetes, Verrucomicrobia, and Firmicutes (Figure 2a and 2b). Verrucomicrobia was more abundant in SUG-W (5.98%) than in SUG-D (1.30%), FOR-D (1.34%), and FOR-W (1.04%) samples. The comparison of the abundance of bacterial phyla





**Figure 4** Nitrogen metabolic pathway analysis performed using KEGG mapper (<http://www.genome.jp/kegg/mapper>). The red box indicates the enzyme that had a significantly high relative abundance in sugarcane site sampled in the dry season. The blue box indicates the enzyme that had a significantly high relative abundance in sugarcane site sampled in the wet season.



**Figure 5** Absolute sequence abundance of microbial functional genes associated with nitrogen metabolism based on metagenomic sequencing data for each of three libraries per sampling point in native forest and sugarcane sites in both dry and wet seasons.

The SEED and KEGG classification results showed high abundance of genes involved in energy metabolism. These genes were classified as participants of nitrogen metabolism (Figure 3) (Figure 4). Thus, the functional analyses of the metagenomic datasets demonstrated that the key genes involved in the N metabolism were related to the energy cycles. For example, in the subcategory of oxidative phosphorylation, most of the sequences showed similarity to enzymes related to energy storage (respiratory chain) such as *coxA* (cytochrome C oxidase subunit I), PPA (inorganic pyrophosphatase), NDH (NADH dehydrogenase), NADH quinone N oxidoreductase subunits L, M, F, H, and D. Additionally, the following sequences associated with enzymatic functions were also observed: methane metabolism (carbon), enzymes that catalyze oxidation-reduction reactions (*cutL/coxL*, *coxS*, and *cutM/coxM*), sulfur (sulfate adenylyltransferase), nitrogen (*nirA*-ferredoxin-reductase nitrite, *nirB*-nitrite reductase large subunit, and nitronate monooxygenase), and carbon sequestration. However, the enzymes involved in the synthesis of carbohydrates by photosynthesis (fructose-6-phosphate phosphoketolase) were lower than those involved in other metabolic pathways.

The results obtained in this study based on the KEEG map suggest that the FOR-D and FOR-W samples have a higher nitrogen-cycling microbial community functional potential due to the higher abundance of genes involved in nitrogen metabolism compared to that of SUG-W and SUG-D samples. Assuming that the functional structure of the community changes with land use conversion, similar to the results observed for the bacterial community composition, this fact justifies the changes in the C and N microbially-mediated processes in the soil.

Comparative metagenomic analyses have indicated that the substrate (i.e., soil or water) plays a fundamental role in determination of the taxonomic and functional profiles of microbial communities. Therefore, soil samples tend to be more similar to each other with respect to taxonomy and the presence of environment-specific genes than the similarity observed in samples from other environments.<sup>34</sup> In FOR-D and FOR-W, nutrient contents and vegetation may be the most important factors affecting the soil bacterial community, while in SUG-D and SUG-W, in addition to these factors, stress conditions (caused by temperature, UV, and drought) might also affect the microbial functionality in soil.

Among the genes restricted to the denitrification process, we found genes belonging to groups *norB*, *norC*, *norD*, and *norF* involved in the reduction of nitric oxide; however, only *norB* and *norF* were present in all the analyzed metagenomes. The presence of groups of genes that encode nitrate reductase, which are responsible for the reduction of nitrate to nitrite, was not found except for *naiD* in FOR-W. The nitrite reductase group of genes, *nirA*, *nirB*, and *nirD*, which are responsible for reducing nitrite to ammonia, were present in all the metagenomes and with the highest prevalence in the metagenome FOR-D. This occurrence of genes involved in nitrite and ammonia assimilation suggests the retention of N and consequently contribute with lower loss by denitrification.

The genes *nifH*, *nifK*, *nifN*, and *nifW*, which encode the nitrogenase reductase enzymes, and the gene *nifB*, which encodes the nitrogen binding protein, were found only in the SUG-D metagenome, probably because the lack of these genes in other metagenomes is due to the low copy numbers which compromises their identification (Figure 5). Another subunit of the *nif* gene (*nifD*) was identified in the SUG-D and SUG-W metagenomic samples, possibly due to soil handling.

The presence of these functional genes associated to N fixation in the metagenomes of soils from sugarcane field with pre-harvest burning and adjacent forest in dry and wet seasons can be explained by the abundance of organisms belonging to the Rhizobiales order (data not shown), as determined by Ribosomal Database Project (RDP) library compare tool using Naive Bayesian classifier.<sup>35</sup> The *Bradyrhizobium* genus has an important role in N and C fixation and biodegradation of aromatic compounds.<sup>36</sup>

## Conclusion

Taken together, the results of the current study showed higher abundance of the most bacterial phyla and nitrogen-cycling microbial community functional potential in wet and dry seasons in forest soil in comparison with sugarcane soil. Hence, both hypothesis of this study are acceptable based on these results. The differential distribution of bacterial taxa across the soil profile in sugarcane field with pre-harvest burning and adjacent forest site in dry and wet seasons indicates that land use, harvesting management and seasonality is having an effect on soil bacterial community composition affecting the integration of carbon and nitrogen pathways related to the energy metabolism. These findings therefore tighten the importance of policies for extinguishing pre-harvest sugarcane straw burning in an effort to improve sugarcane ethanol sustainability based on the nitrogen-cycling microbial community functional potential.

## Acknowledgments

We thank Gabriel Henrique Sant'Ana Pereira, manager of Vassununga State Park, for support in field work.

## Funding

This work was supported by Fundação de Amparo à Pesquisa do Estado de São Paulo (FAPESP, Proc. 2009/54274-9), Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq), and Coordenação de Aperfeiçoamento de Pessoal de Nível Superior-Brasil (CAPES) — Finance Code 001 and 23038.006927/2014-35/Premium 116/2017. E.G.M.L. Thanks CNPq for the Research Productivity Grant (305032/2019-4).

## Conflicts of interest

The authors declare that there is no conflict of interest.

## References

1. CONAB. Brazilian National company of provision. 2020.
2. Pang Z, Dong F, Liu Q, et al. Soil metagenomics reveals effects of continuous sugarcane cropping on the structure and functional pathway of rhizospheric microbial community. *Frontiers in Microbiology*. 2021.
3. Agro-environmental protocol. Secretaria do Meio Ambiente. 2020.
4. Rachid CTCC, Santos AL, Piccolo MC, et al. Effect of sugarcane burning or green harvest methods on the Brazilian Cerrado soil bacterial community structure. *PLoS One*. 2013;8(3):e59342.
5. Rachid CTCC, Piccolo MC, Leite DCA, et al. Physical-chemical and microbiological changes in Cerrado Soil under differing sugarcane harvest management systems. *BMC Microbiology*. 2012;12(1):170.
6. Souza RA, Telles TS, Machado W, et al. Effects of sugarcane harvesting with burning on the chemical and microbiological properties of the soil. *Agriculture, Ecosystems & Environment*. 2012;155:1–6.
7. Val-Moraes SP, Macedo HS, Kishi LT, et al. Liming in the sugarcane burnt system and the green harvest practice affect soil bacterial community in north-eastern São Paulo, Brazil. *Antonie van Leeuwenhoek*. 2016;109:1643–1654.
8. Navarrete AA, Pitombo LM, Brandani CB, et al. Multi-analytical interactions in support of sugarcane agroecosystems sustainability in tropical soils. In: Alexandre de Oliveira, editor. *Sugarcane - Technology and Research*. IntechOpen; 2018:3–28.
9. DE-Polli H, Pimentel MS. Soil quality indicators. In: Aquino AM, Assis RL, editors. *Biological processes in the soil-plant system: tools for sustainable agriculture*. Brasília: Embrapa-SCT; 2005:17–28.
10. Doran JW, Parkin TB. Defining soil quality. In: Doran JW, Coleman DC, Bezdicek DF, editors. *Defining soil quality for a sustainable environment*. Madison: Soil Science Society of America Special Publications; 1994:3–21.
11. Mendes LW, Tsai SM, Navarrete AA, et al. Soil-borne microbiome: Linking diversity to function. *Microbial Ecology*. 2015;70:255–265.
12. Kennedy AC, Smith KL. Soil microbial diversity and the sustainability of agricultural soils. *Plant Soil*. 1995;170:75–86.
13. De Chaves MG, Silva GGZ, Rossetto R, et al. Acidobacteria subgroups and their metabolic potential for carbon degradation in sugarcane soil amended with vinasse and nitrogen fertilizers. *Frontiers in Microbiology*. 2019;10:1680.
14. Jesus EC, Marsh TL, Tiedje JM, et al. Changes in land use alter the structure of bacterial communities in Western Amazon soils. *The ISME Journal*. 2009;3:1004–1011.
15. Navarrete AA, Diniz TR, Braga LPP, et al. Multi-analytical approach reveals potential microbial indicators in soil for sugarcane model systems. *PLoS ONE*. 2015;10(6):e0129765.

16. Nishisaka CS, Youngerman C, Meredith LK, et al. Differences in N<sub>2</sub>O fluxes and denitrification gene abundance in the wet and dry seasons through soil and plant residue characteristics of tropical tree crops. *Frontiers in Environmental Science*. 2019;7:11.
17. Navarrete AA, Kuramae EE, de Hollander M, et al. Acidobacterial community responses to agricultural management of soybean in Amazon forest soils. *FEMS Microbiology Ecology*. 2013;83:607–621.
18. Navarrete AA, Tsai SM, Mendes LW, et al. Soil microbiome responses to the short-term effects of Amazonian deforestation. *Molecular Ecology*. 2015;24:2433–2448.
19. De Caceres M, Legendre P, Moretti M. Improving indicator species analysis by combining groups of sites. *Oikos*. 2010;119:1674–1684.
20. Parks DH, Tyson GW, Hugenholtz P, et al. STAMP: Statistical analysis of taxonomic and functional profiles. *Bioinformatics*. 2014;30:3123–3124.
21. Overbeek R, Begley T, Butler RM, et al. The subsystems approach to genome annotation and its use in the project to annotate 1000 genomes. *Nucleic Acids Research*. 2005;33:5691–5702.
22. Kanehisa M, Goto S, Sato Y, et al. KEGG for integration and interpretation of large-scale molecular data sets. *Nucleic Acids Research*. 2012;40:D109–D114.
23. Janssen PH. Identifying the dominant soil bacterial taxa in libraries of 16S rRNA and 16S rRNA genes. *Applied Environmental Microbiology*. 2006;72:1719–1728.
24. Roesch LFW, Fulthorpe RR, Riva A, et al. 2007. Pyrosequencing enumerates and contrasts soil microbial diversity. *The ISME Journal*. 2007;1:283–290.
25. Quirino BF, Pappas GJ, Tagliaferro AC, et al. Molecular phylogenetic diversity of bacteria associated with soil of the savanna-like Cerrado vegetation. *Microbiological Research*. 2009;164:59–70.
26. Bruce T, Martinez IB, Maia Neto O, et al. Bacterial community diversity in the Brazilian Atlantic forest soils. *Microbiology Ecology*. 2010;60:840–849.
27. Faoro H, Alves AC, Souza EM, et al. Influence of soil characteristics on the diversity of bacteria in the Southern Brazilian Atlantic Forest. *Applied Environmental Microbiology*. 2010;76:4744–4749.
28. Araujo JF, De Castro AP, Costa MMC, et al. Characterization of soil bacterial assemblages in Brazilian savanna-like vegetation reveals acidobacteria dominance. *Microbial Ecology*. 2012;64:760–770.
29. Marín I. Proteobacteria. In: Amils R, editor. *Encyclopedia of Astrobiology*. Berlin, Heidelberg: Springer; 2014.
30. Ward NL, Challacombe JF, Janssen PH, et al. Three genomes from the phylum acidobacteria provide insight into the lifestyles of these microorganisms in soils. *Applied and Environmental Microbiology*. 2009;75:2046–2056.
31. Russo SE, Legge R, Weber K, et al. Bacterial community structure of contrasting soils underlying Bornean rain forests: inferences from microarray and next-generation sequencing methods. *Soil Biology and Biochemistry*. 2012;55:48–59.
32. Cannon SA, Lester ED, Shafaat HS, et al. Bacterial diversity in hyperarid Atacama desert soils. *Journal of Geophysical Research*. 2007;112:G04S17.
33. Jeffries TC, Seymour JR, Gilbert JA, et al. Substrate type determines metagenomic profiles from diverse chemical habitats. *PLoS ONE*. 2011;6:e25173.
34. Tringe SG, von Mering C, Kobayashi A, et al. Comparative metagenomics of microbial communities. *Science*. 2005;308:554–557.
35. Wang Q, Garrity GM, Tiedje JM, et al. Naive Bayesian classifier for rapid assignment of rRNA sequences into the new bacterial taxonomy. *Applied and Environmental Microbiology*. 2007;73:5261–5267.
36. Yuan H, Ge T, Chen C, et al. Significant role for microbial autotrophy in the sequestration of soil carbon. *Applied and Environmental Microbiology*. 2012;78:2328–2336.