

Distribution of extended spectrum beta-lactamase (ESBL) genes among *Acinetobacter baumannii* isolated from burn infections

Abstract

In this study, extended spectrum beta-lactamase (ESBL) genes among *Acinetobacter baumannii* isolated from hospitalized patients with burn infections were evaluated. From December 2016 to March 2017, sixty-five isolates (72.2%) of *A. baumannii* were obtained from 90 burn samples of patients which were hospitalized in two hospitals in Baghdad, Iraq. The identification of *A. baumannii* strains was conducted by API 20NE and the presence of bla_{OXA-51} gene. Kirby-Bauer disc diffusion method was used for antibiotic susceptibility test. The frequency of ESBL producers was evaluated by double disk synergy test (DDST) and prevalence of bla_{TEM} , bla_{SHV} , bla_{CTX-M} genes were assayed by PCR. The results revealed that bla_{OXA-51} gene was found in all *A. baumannii* isolates as a species-specific gene. Of sixty-five isolates, 40 (61.53%) *A. baumannii* were ESBL producers according to DDST. High rates of resistance to cefotaxime, ceftazidime, piperacillin, and cefepime antibiotics were detected among ESBL producing *A. baumannii* isolates. The rate of resistance to other antibiotics was as the follows: 29(72.5%) to meropenem, 32(80%) to imipenem, 30 (75%) to piperacillin/tazobactam, 26 (65%) to ampicillin/sulbactam. The prevalence of bla_{TEM} , bla_{CTX-M} , bla_{SHV} genes among ESBL producing *A. baumannii* isolates was 30 (75%), 18 (45%) and 11 (27.5%) respectively. Our results demonstrated the role of bla_{TEM} gene in the resistance of beta-lactam antibiotics in the local strains of burn infections. The high rate of resistance in ESBL producing *A. baumannii* strains detected in this study highlights the necessity for the quick detection of ESBL isolates and the antimicrobial management, particularly in burn units.

Keywords: *A. baumannii*, ESBL genes, burns

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Introduction

Acinetobacter baumannii is an important cause of nosocomial infections and a leading cause of mortality and morbidity among hospitalized patients and has been associated with a wide variety of diseases in the intensive care units (ICU). Widespread dissemination of *A. baumannii* infections, which are resistant to beta-lactams antibiotics particularly to the 3rd generation of cephalosporins and carbapenems, has become a globally significant problem.² These extended-spectrum β -lactamases (ESBLs) are enzymes, plasmid mediated and produced by gram negative bacilli that mediate a resistance to penicillins, cephalosporins, and monobactams.³ These ESBLs are commonly recognized in Enterobacteriaceae, *Pseudomonas aeruginosa*, and *A. baumannii* and are found worldwide.⁴ Extended-spectrum β -lactamases (ESBLs) of *A. baumannii* are encoded by TEM-type, SHV-type and CTX-M-type genes which are subjected to phenotypical resistance to penicillins and 3rd generation cephalosporins. These enzymes are mostly plasmid mediated and most are parts of the TEM and SHV families.^{2,5} Different microorganisms are related to contaminations in hospital environments, but the main pathogens include oxacillin-resistant *Staphylococcus aureus* (ORSA), vancomycin-resistant *Enterococcus* sp. (VRE), extended-spectrum beta-lactamases (ESBL), and carbapenem-resistant *Acinetobacter baumannii*.^{6,7} *A. baumannii* is able to survive on human skin, equipment, and objects in different sections, especially burn centers and ICUs for a long time.⁸ The major problem of concern with *A. baumannii* is the choice of suitable antibiotic for treatment, especially in nosocomial infections.⁹ The aim of this study was to investigate the detection and distribution of ESBLs encoding genes and its drug resistance against carbapenems

and cephalosporins among *A. baumannii* strains isolated from patients with burn infections.

Materials and methods

Sampling and isolation of bacteria

The study was initiated in December 2016 by collecting 90 samples from burn infections of burn units in 2 hospitals of Baghdad City in Iraq. The samples were collected by sterile swabs which used moistened with Trypticase Soy Broth (TSB) medium. [Based on what type of test characterization and identification of isolates were performed?] Immediately after the collection, they were again stored in the medium and incubated at 37°C for 24 hours. After the growth in TSB, the samples were cultured on Blood agar and MacConkey agar incubated at 37°C for 24 hours. The identification of the isolates was accomplished by API 20NE and confirmed by tracking the bla_{OXA-51} -like carbapenemase gene, which is intrinsic to this species, using single PCR.¹⁰

Antibiotic susceptibility testing

The susceptibility of *A. baumannii* isolates to carbapenems and cephalosporins (Mast, UK) was tested by Kirby-Bauer disk-diffusion method. The antibiogram procedure was performed as the manufacturer which manufacture constructed. In brief, 1.5×10^8 CFU/ml of bacterial suspension, equivalent to McFarland Turbidity Standard No. 0.5, was transferred on Muller-Hinton agar medium (Merck, Germany) and antibiogram disks containing: cefipime (FEP:30 μ g), cefotaxime (CTX:30 μ g), ceftazidime (CAZ:30 μ g), imipenem (IPM:10 μ g),

meropenem (MEM: 10µg), piperacillin/tazobactam (PTZ: 100/10µg) and ampicillin/sulbactam (10/10µg) were placed on the medium. Then, the media were incubated for 18 hours at 37°C. The results were interpreted according to the Clinical and Laboratory Standards Institute (CLSI) recommendations.¹¹ A control strain of *Pseudomonas aeruginosa* ATCC 27853 was used for quality control of susceptibility testing.

Phenotypic identification of ESBL producing isolates

Phenotypic identification of ESBL producing isolates have been carried out using disk potentiating test and confirmed by DDST screening method. Antibiogram disks containing CAZ (30µg), CTX (30µg), ceftazidime (30µg) + clavulanic acid (10µg) and cefotaxime (30µg) + clavulanic (10µg) were used. Pairs of disks (ceftazidime with ceftazidime/clavulanic acid and cefotaxime with cefotaxime/clavulanic) were placed on Muller-Hinton agar medium with 20 mm space between them. According to the CLSI criteria and manufacturer

instruction, an increase in zone diameter of ≥5 mm in the presence of clavulanic acid indicated the presence of ESBL in the test organism. *Klebsiella pneumoniae* ATCC700603 was used as a positive control for ESBL production.¹¹

Amplification of ESBL genes

The specific primers including CTX-M, SHV and TEM (Table 1) were used for PCR amplification of the genes. The PCR mixture contained the DNA template, the forward/reverse primers, and the master mix (Promega, USA). Amplification was carried out with the thermal cycling conditions according to the relevant studies and mentioned in Table 1. The PCR amplification was conducted in the thermal cycler device (Applied Biosystems, USA). Agarose gel electrophoresis of the amplified PCR products with 100bp size marker (Promega, USA) were carried out in a 2% agarose gel for 2h at 80V and stained with ethidium bromide. The PCR product bands were then checked under UV irradiation.

Table 1 Primers used for PCR amplification of the ESBL genes

Primer name	Primer sequence (5' to 3')	Annealing temp. (°C)	Product size (bp)	Reference
TEM	F:ATGAGTATTCAACATTTCCG	53	931	12
	R: CCAATGCTTAATCAGTGAGC			
SHV	F:AAGATCCACTATCGCCACAG	60	231	13
	R:ATTGAGTTCCGTTTCCCAGCGG			
CTX-M	F: CGATGTGCAGTACCAGTAA	60	585	14
	R:TTAGTGACCAGAATAAGCGG			

Results and discussion

In the current study 65 *A. baumannii* (72.2 %) were isolated out of 90 samples which collected from patients with burn infections in 2 hospitals of Baghdad City in Iraq. The isolates of *A. baumannii* were identified by using API 20NE system (bioMérieux) and the identification was confirmed by PCR amplification of the specific gene *bla*_{OXA-51} (Figure 1). *A. baumannii* isolates were identified by molecular detection of the carbapenemase gene *bla*_{OXA-51} in all 75 isolates, which then used to identify *A. baumannii*-mediated infections. Many previous studies indicated to the identification of *A. baumannii* at the species level by PCR detection of *bla*_{OXA-51}-like genes [10, 15]. This study revealed that *A. baumannii* was one of the most important sources of nosocomial infections in ICUs and the prevalence of this species in ICU equipment in four hospitals in Baghdad city was 22.05%. ICUs are one of the units that have the highest occurrence rates of infections and the prevalence of infection is two to five times higher in ICUs than in the general inpatient population.^{16,17} *Acinetobacter* spp. are more frequently colonized on inanimate objects and hands of staff in the ICU than *Pseudomonas* spp and *Staphylococcus aureus*.¹⁸ The study conducted by Mehraban et al.¹⁸ revealed the high frequency of genus *Acinetobacter* among Gram negative bacteria isolated from environmental surfaces of ICUs of four hospitals located in the city of Qom, Iran.¹⁹ In another study in an adult ICU tertiary care Hospital in Riyadh, Saudi Arabia, *A. baumannii* was recognized as the most frequently isolated bacteria.²⁰ Multidrug resistant *A. baumannii* is

the most common cause of nosocomial infection in burn patients in Iran.²¹ *Acinetobacter* spp. can survive dry conditions for long periods, thus *A. baumannii* is frequently isolated from medical equipment and the contamination of hospital equipment caused by *Acinetobacter* outbreaks in the hospital environment is prevalent.²² Phenotypic detection of ESBL producing isolates of *A. baumannii* have been carried out using DDST screening method (Figure 2). From total of 65 isolates, 40 (61.53%) *A. baumannii* isolates were identified to be produced by ESBL enzymes. Susceptibility of 40 ESBL producing *A. baumannii* isolates to 8 of beta-lactam an antibiotic was evaluated by the Kirby-Bauer disk diffusion method (Table 2).The results showed high resistance to all antibiotics used in this study. Ninety-five percent, 92.5 %, 92.5 % and 90 % out of 40 *A. baumannii* isolates were resistant to Cefotaxime, Ceftazidime, Cefepime, and Piperacillin respectively. Also high resistance to Imipenem and moderate resistance to Meropenem, Piperacillin/tazobactam, and Ampicillin/sulbactam. Multidrug-resistant (MDR) *A. baumannii* became prevalent in many hospitals all over the world and has been recognized as one of the most important nosocomial pathogens especially in burn units.²³ Our results showed high resistance of *A. baumannii* isolates to beta-lactam antibiotics such as penicillins, cephalosporins, and carbapenems used in the susceptibility test. Production of carbapenem-hydrolyzing beta-lactamases is one of the significant mechanisms of carbapenem resistance.²⁴ The extended-spectrum beta-lactamases, ESBLs, play an important role in resistance against third-generation cephalosporins such as cefotaxime, ceftazidime, and cefepime.¹⁰ Specific primers

were used to detect ESBL genes by PCR assay. Of all 40 *A. baumannii* isolates, 30 (75%), 18 (45%) and 11 (27.5%) isolates have been confirmed to harbor the bla_{TEM} , bla_{CTX-M} and bla_{SHV} genes, respectively. It was obvious that bla_{TEM} genes were the most frequent ESBL types among *A. baumannii* strains in burn infections. PCR products of ESBL genes were analyzed by gel electrophoresis to detect the prevalence of these genes (Figures 3-5). In this study, the prevalence of ESBLs encoding genes among *A. baumannii* isolates has been investigated showing a high frequency of bla_{TEM} genes among the isolates (75%). ESBL positive strains of *A. baumannii* and *P. aeruginosa* are increasingly found in hospital isolates. These strains revealed resistant to available antibiotics and these genes can transfer to other clinical strains. Molecular detection would determine which types of ESBL are present in each isolate and the identification of beta-lactamases would be essential for an epidemiological investigation of antimicrobial resistance.¹² Ting et al.²⁴ investigated the drug resistance genes in 7 strains of Imipenem-resistant *A. baumannii* including TEM, SHV, CTX-M, DHA, CIT, IMP, VIM, KPC, OXA-

23, the results found TEM (100%) and OXA-23 (100%) genes among the isolates, but the other genes such as SHV, CTX-M, DHA, CIT, IMP, VIM, KPC could not be detected.²⁵ The present findings was in contrast to the results of Khalilzadegan et al.²⁵ who detected that the frequencies of bla_{TEM} and bla_{CTX} genes were 3.2% and 19.4%, respectively, while the level of resistance to various antibiotics was in the range of 69.4% to 100%.²⁶ Another research reported that 40% of the isolates carried bla_{CTX} and 12.8% carried bla_{TEM} .²⁶ The study of Odewale et al. in Ladoke Akintola University teaching hospital, Osogba, Nigeria, detected that 72.7% of patients in the ICU were most infected with *A. baumannii* and the isolates showed 90.9% resistance to both Ceftriaxone and Ceftazidime. The PCR results showed that bla_{TEM} and bla_{CTX-M} genes were positive in some isolates, while bla_{SHV} was not detected in any of the isolates. ESBL genes are plasmid-borne and are easily transmitted from one strain to another; this explains why these bacteria have firmly established themselves as MDR nosocomial pathogens whose infections no longer respond to treatment by commonly used antibiotics.¹⁴

Table 2 The antibiotic resistance patterns of *A. baumannii*

Antibiotic	Number of sensitive (%)	Number of intermediate (%)	Number of resistant (%)
Cefipime	3 (7.5)	0 (0.0)	37(92.5)
Cefotaxime	1 (2.5)	1 (2.5)	38(95)
Ceftazidime	1 (2.5)	2 (5)	37(92.5)
Imipenem	6 (15)	2 (5)	32(80)
Meropenem	7 (6.6)	4 (10)	29(72.5)
Piperacillin/tazobactam	7 (17.5)	3 (7.5)	30(75)
Ampicillin/sulbactam	10 (25)	4 (10)	26(65)
Piperacillin	2 (5)	2 (5)	36(90)



Figure 1 Electrophoresis of the amplified products of bla_{OXA51} (353bp) genes by PCR in a 2% agarose gel. Lane 2-11: positive result of gene detection in *A. baumannii* isolates; Lane 12: negative result for *A. Iwoffii*; Lane 1: negative control (PCR product without the DNA template); Lane M:100bp DNA ladder.

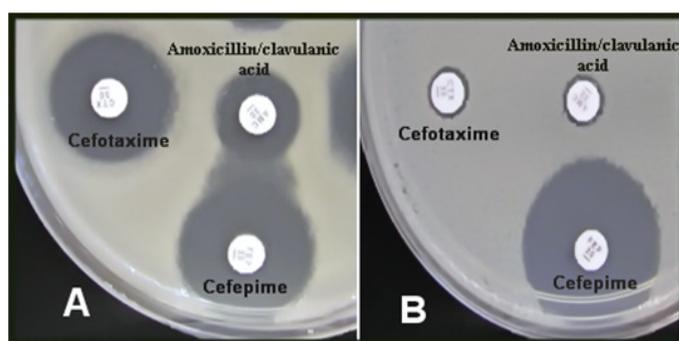


Figure 2 Double-disk synergy test for extended spectrum beta-lactamases in *A. baumannii*, A (positive result), B (negative result). AMC – Amoxycillin + Clavulanate, CTX-Cefotaxime, FEP–Cefepime (A clear extension of the edge (Synergy) of the FEP inhibition zone towards the disc containing clavulanate).

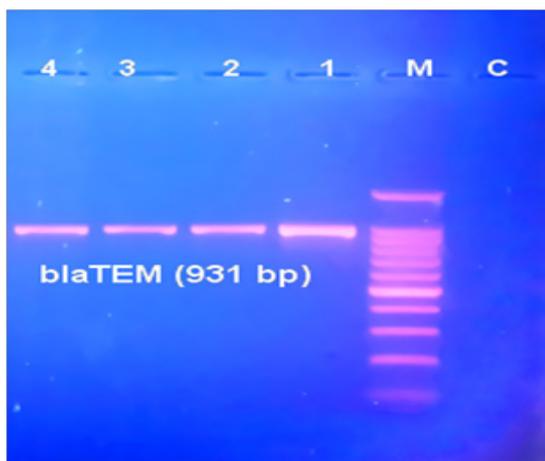


Figure 3 Electrophoresis of the amplified products of *bla_{TEM}* (931bp) genes by a PCR in a 2 % agarose gel. Lane 1-4: positive result of gene detection in *A. baumannii* isolates; Lane C: negative control (PCR product without the DNA template); Lane M: 100bp DNA ladder.



Figure 4 Electrophoresis of the amplified products of *bla_{CTX-M}* (585bp) genes by a PCR in a 2 % agarose gel. Lane 5 and 10: positive result of gene detection in *A. baumannii* isolates; Lane 1: negative control (PCR product without the DNA template); Lane M: 100bp DNA ladder.

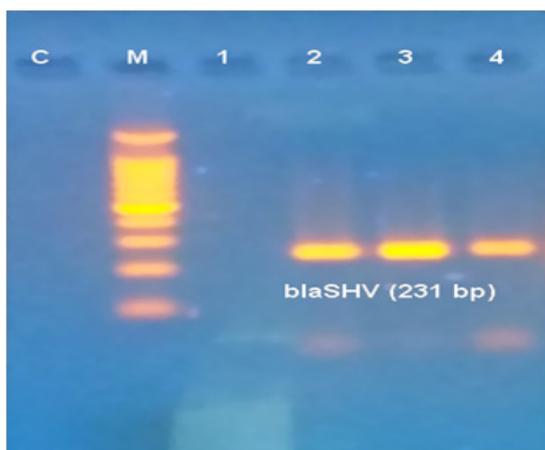


Figure 5 Electrophoresis of the amplified products of *bla_{SHV}* (231bp) genes by a PCR in a 2 % agarose gel. Lane 2-4: positive result of gene detection in *A. baumannii* isolates; Lane C: negative control (PCR product without the DNA template); Lane M: 100bp DNA ladder

Conclusion

The presence of *A. baumannii* in ICU increases the risk of transmission to patients leading to nosocomial infections. The prevention of the nosocomial infections needs the quick detection of the sources of contamination, especially on the equipment. Detection of ESBL genes by PCR is known as an effective and sensitive method for epidemiological analysis and tracing MDR *A.baumannii*. ESBL producing bacteria should be identified quickly so that appropriate antibiotic usage can be conducted.

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Conflict of interest

The author declares there is no conflict of interest.

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