

Research Article





Simultaneous determination of paracetamol and codeine phosphate in pharmaceuticals using molecular absorption spectroscopy and classical least squares method

Summary

In this article, a combined method of molecular absorption spectroscopy and classical least squares is used to simultaneously determine paracetamol (PAR) and codeine phosphate (COP) in pharmaceutical samples without the need for prior separation or extraction steps. The absorption spectra of standard solutions and samples were measured in the wavelength range of 210 to 290 nm with a step size of 0.5 nm. The concentrations of PAR and COP in the sample solutions were calculated using a self-made program written in Microsoft Excel 2016 and Visual Basic for Applications (VBA). The reliability of the method was verified through the accuracy and reproducibility of the measurements when analyzing PAR and COP in Effer-Paralmax® Codeine tablets, and comparing the average values of their concentrations in the samples with the standard HPLC method.

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Introduction

Combining chemometrics with spectroscopic methods using fullspectrum data and various algorithms, statistics, and computers has been studied to simultaneously determine constituents in overlapping spectra mixtures. The advantages of these methods include a simple analysis process that does not require the separation or extraction of compounds from each other, and rapid execution that saves time, chemicals, and utilizes all data for calculations, resulting in increased accuracy in determination. These methods include classical least squares (CLS), partial least squares (PLS), principal component analysis (PCA), artificial neural networks (ANN), Kalman filtering, etc. Some authors have studied the application of these methods for analyzing real objects.¹⁻⁸

In this paper, we investigate the development of an analysis procedure for simultaneous determination of Paracetamol (PAR) and Codeine phosphate (COP) in Effer-Paralmax® Codeine effervescent tablets that are currently circulating in the Vietnamese market. PAR, with a molecular formula of $C_{s}H_{0}NO_{2}$, M = 151.17 g/mol, is a metabolite with activity of phenacetin, a pain-relieving and antipyretic medication that is most effective in reducing pain, reducing body temperature in feverish patients by acting on the lower hypothalamus, dissipating heat by dilating blood vessels and increasing peripheral blood flow. COP, with a molecular formula of $C_{18}H_{24}PNO_7$, M = 397.00 g/mol, is a phenanthrene derivative, also known as methylmorphine, that has pharmacological effects of pain relief, cough suppression, and excellent antidiarrheal and neuropathic pain management. Effer-Paralmax Codein tablets combine two main components, PAR and COP, which are used to treat mild to moderate pain, fever reduction, and gastric irritation.9 To analyze these components, different individual analytical methods such as spectroscopic methods,10-12 and HPLC methods¹³ can be used. The individual analytical procedures require much time and complexity, and HPLC method has a significant advantage but a high cost. According to,¹⁰⁻¹² the absorption spectra of PAR and COP overlap, so we chose the UV-VIS molecular absorption spectroscopy method with full-spectrum data and CLS algorithm to

simultaneously determine the amount of Paracetamol and Codeine phosphate in pharmaceuticals. The proposed method opens up the possibility of fast, cost-effective analysis that can be applied in practical analysis and pharmaceutical testing.

Experiment

Equipment and chemicals

Equipment

- Cary 60 UV-Vis spectrophotometer from Agilent Technologies, USA, equipped with quartz cuvettes from the Laboratory of Applied Chemistry, Department of Chemistry, Hue University of Science, capable of quickly scanning spectra in the range of wavelengths from 190 – 990 nm, connected to a computer and Cary WinUV software for storing spectral data in an excel spreadsheet format.
- 2) Other equipment and tools: Precisa XB 2204 analytical balance with an accuracy of 0.0001 g; Quartz double-distilled water stills, Fistreem Cyclon and Aquatron brands; HTL 100 μl and 1000 μl micropipettes; Glassware: pipettes, volumetric flasks, beakers, Erlenmeyer flasks, glass rods, filter paper, glass funnels, ...

Chemicals

- a) Paracetamol (PAR) standard substance with a purity of 100.0% from the Central Drug Testing Institute, Ministry of Health.
- b) Codeine phosphate (COP) standard substance with a purity of 97.5% from the Drug Testing Institute of Ho Chi Minh City, Ministry of Health.
- c) Acetonitrile CH₃CN (Merck), distilled water H₂O.
- d) Preparation of solvent: water/acetonitrile (9:1 v/v): Pour 100 ml of CH₃CN into a 1 L volumetric flask and add distilled water to the mark, shake well.

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- e) Preparation of PAR, COP standard stock solutions:
- f) Prepare a PAR solution of 25 μ g/mL: Accurately weigh 12.5 mg of Paracetamol standard substance, add to a 250 mL volumetric flask, add solvent to about 150 mL, shake well until dissolved and add solvent to the mark, resulting in a PAR solution of 50 μ g/mL. Take 100 mL of the PAR 50 μ g/mL solution (using a 100 mL volumetric flask) and 100 mL of solvent (using a 100 mL volumetric flask) into a 250 mL glass beaker, stir well to obtain a PAR working solution of 25 μ g/mL.
- g) Prepare a COP solution of 25 μ g/mL: Accurately weigh 12.8 mg of Codeine phosphate standard substance, add to a 250 mL volumetric flask, add solvent to about 150 mL, shake well until dissolved and add solvent to the mark, resulting in a COP solution of 50 μ g/mL. Take 100 mL of the COP 50 μ g/mL solution (using a 100 mL volumetric flask) and 100 mL of solvent (using a 100 mL volumetric flask) into a 250 mL glass beaker, stir well to obtain a COP working solution of 25 μ g/mL.

Analysis method

For determination of PAR and COP simultaneously in a mixed solution, we used the UV-VIS molecular absorption spectroscopy method with a full-spectrum combined with the classic least squares method (CLS) and a calculation program written in Microsoft Excel 2016 and Visual Basic for Applications (VBA). The essence of the method is clearly presented in.⁸ The process of measuring and calculating the concentration of PAR and COP is as follows:

Step 1: Prepare standard solutions of each component to be determined and their mixed solutions.

Step 2: Scan the solutions in the appropriate wavelength range on the UV-Vis spectrophotometer, save and retrieve the measured data (Export data) in "*.CSV" file format.

Step 3: Run the CLS-Excel program⁸ to calculate the concentration of components in the mixed solution.

Statistical quantities

Relative error

The relative error between the determined concentration and the prepared concentration (RE %) is calculated as:

$$RE\% = (C - C_0) \cdot 100/C_0 \tag{1}$$

Where C is the determined concentration (μ g/mL) and C₀ is the known standard solution concentration (μ g/mL).

Repeatability

Repeatability is evaluated by the relative standard deviation value RSD%:

$$RSD\% = SD.100/C_{mean}$$
(2)

Here, SD is the standard deviation and $C_{_{mean}}$ is the average concentration after n measurements ($\mu g/mL).$

For internal laboratory quality control, the repeatability of the

method is achieved when the RSD% values obtained are less than $1/2RSD_{Horwitz}$

$$RSD_{H} = RSD_{Horwitz} = 2^{(1-0.5 \cdot lgC)}$$
(3)

where C is the concentration expressed as a fraction. (for example, C= 5 μ g/mL = 5.10⁻⁶)

Method accuracy

a) Recovery

The recovery of the method is calculated based on the standard addition method using the following formula:

$$Rev\% = (C_2 - C_1) \cdot 100/C_{add}$$
(4)

where $C_2 (\mu g/mL)$ is the determined concentration after adding the standard, $C_1 (\mu g/mL)$ is the determined concentration before adding the standard, and $C_{add} (\mu g/mL)$ is the concentration of the added standard.

b) Comparison of the average sample values of the research method with the standard method

According to,¹⁴ to determine the accuracy of the method, the same sample can be analyzed repeatedly using both the research method and the standard method, and the two average sample values can be compared using the student's t-test.

$$t_{exp} = \frac{\left| \overline{X}_A - \overline{X}_B \right|}{\sqrt{\left(S_A^2 / n_A \right) + \left(S_B^2 / n_B \right)}}$$
(5)

Where: t_{exp} : experimental student's t-value, $\overline{X}_A, \overline{X}_B$: the average values of method A and B; n_A, n_B : numbers of experiment of method A and B. Compare the value of t_{exp} with the value of $t(\alpha, \nu)$, in which α is the chosen significance level, ν is degrees of freedom of two methods. If $t_{exp} < t(\alpha, \nu)$, then the difference in the average values of the two methods is not significant.

Results and discussion

Accuracy and precision of the analytical method

Standard working solutions of PAR 25 μ g/mL and COP 25 μ g/mL were added to 12 volumetric flasks (VF) numbered from 1 to 12, each containing 25 mL, and the volume was adjusted to the mark with H₂O-ACN solvent mixture (9:1, V/V). In VF 13 containing 25 mL, 13.5 mL of PAR standard solution of 50 μ g/mL and 3.0 mL of COP standard solution of 25 μ g/mL were added and diluted to the mark with the solvent mixture. The solutions were scanned in the wavelength range of 210 – 290 nm with a step size of 0.5 nm. The preparation and measurement were repeated 3 times. The absorption spectra of the solutions are shown in Figure 1. The concentration of PAR and COP in the mixed solutions were calculated using the CLS-Excel program, and the relative error RE% and relative standard deviation RSD% of the analytical results were determined. The analytical results of PAR and COP in the prepared mixed solutions in the laboratory along with the corresponding statistical parameters are presented in Table 1.

Table I Analytical result	of PAR and COP in laborator	y mixtures by CLS-Excel m	nethod with statistical quantities
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Sample Conc. ratio	Meas. time	PAR		СОР				
N⁰ (µg/mL) of PAR/COP		C _{PAR} (µg/mL)	RE (%)	Statistical quantities	C _{cop} (μg/mL)	RE (%)	Statistical quantities	
		I	18.961	-0.21	C - 10.010	0.988	-1.20	C = 0.000
MI	19:1	2	19.056	0.29	$C_{tb} = 19.019$ RSD(%) = 0.267 I/2RSD = 5.136	0.992	-0.80	$R_{tb}^{-} = 0.999$ RSD(%)= 1.630 1/2RSD = 8.000
		3	19.040	0.21	$RE_{tb}(\%) = 0.10$	1.018	1.80	$RE_{tb}(\%) = -0.07$
		I	18.077	0.43		1.975	-1.25	
M2	18:2	2	17.941	-0.33	C _{tb} = 17.991 RSD(%)= 0.414	2.004	0.20	$C_{\pm} = 1.997$ RSD(%)= 0.996
		3	17.956	-0.24	$RE_{tb}(\%) = -0.05$	2.013	0.65	$RE_{tb}(\%) = -0.13$
		L	17.070	0.41		2.970	-1.00	
M3	17:3	2	17.080	0.47	C, = 17.075 RSD(%)= 0.029	2.996	-0.13	C_ = 2.990 RSD(%)= 0.595
		3	17.076	0.45	$RE_{tb}(\%) = 0.44$	3.004	0.13	$RE_{tb}(\%) = -0.33$
		I	15.998	-0.01		3.951	-1.23	
M4	16:4	2	16.000	0.00	C_ = 16.008 RSD(%)= 0.097 1/2RSD = 5.271	4.005	0.12	C, = 3.985 RSD(%)= 0.743 L/2RSD = 6.493
		3	16.026	0.16	RE_{tb} (%)= 0.05	3.999	-0.02	$RE_{tb}(\%) = -0.38$
		I	15.027	0.18	C _ 15 007	4.978	-0.44	C - C 012
M5	15:5	2	15.026	0.17	$C_{\rm c} = 15.027$ RSD(%)= 0.010 I/2RSD_ = 5.322	5.028	0.56	$C_{1} = 5.013$ RSD(%)= 0.612 I/2RSD_ = 6.279
		3	15.029	0.19	$RE_{tb}(\%) = 0.18$	5.034	0.68	$RE_{tb}(\%) = 0.27$
		I	14.123	0.88		6.034	0.57	
M6	14:6	2	14.135	0.98	C, = 14.128 RSD(%)= 0.046	6.085	1.42	C, = 6.072 RSD(%)= 0.545
		3	14.125	0.89	RE_{tb} (%) = 0.92	6.096	1.60	$RE_{tb}(\%) = 1.20$
		I	13.068	0.52	0 10.070	7.082	1.17	0 7 0 7 0
M7	13:7	2	12.948	-0.40	C ₁ = 12.973 RSD(%)= 0.654	7.090	1.29	C, = 7.078 RSD(%)= 0.196
		3	12.904	-0.74	$RE_{tb}(\%) = -0.21$	7.063	0.90	$RE_{tb}(\%) = 1.12$
		I	12.029	0.24		8.092	1.15	
M8	12:8	2	11.911	-0.74	C_ = 11.953 RSD(%)= 0.551	8.088	1.10	$C_{tb} = 8.096$ RSD(%)= 0.131
		3	11.919	-0.67	RE_{tb} (%) = -0.39	8.108	1.35	$RE_{tb}(\%) = 1.20$
		1	10.984	-0.15		9.088	0.98	
M9	11:9	2	10.889	-1.01	C, = 10.915 RSD(%)= 0.550	9.102	1.13	С _{tb} = 9.095 RSD(%)= 0.077
		3	10.873	-1.15	$1/2RSD_{H} = 5.576$ RE _{tb} (%) = -0.77	9.096	1.07	1/2RSDH = 5.747 RE _{tb} (%)= 1.06
		1	10.039	0.39		10.083	0.83	
		2	10.043	0.43	$C_{\mu} = 10.042$	10.185	1.85	$C_{\pm} = 10.152$
M10	10:10	3	10.043	0.43	$I/2RSD_{H} = 5.657$ BE (%) = 0.42	10.187	1.87	RSD(%)= 0.366 I/2RSDH = 5.657 RF (%)= 1.52
		I	26.990	-0.04		2.970	-1.00	
		2	26.988	-0.06	$C_{tb} = 26.990$	2.993	-0.23	$C_{tb} = 2.998$
MII	27:3	3	26.992	-0.03	$I/2RSD_{H} = 4.871$ RE (%) = -0.04	3.030	1.00	$1/2RSD_{H} = 6.781$ RE (%) = -0.08
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Figure I Molecular absorption spectra of PAR10 and COP10 standard solutions and sample solutions at different concentration ratios in laboratory-prepared mixtures:

- (1) PAR10 standard solution at 10 $\mu\text{g/mL}$
- (2) COP10 standard solution at 10 µg/mL
- (3) M1: Solution containing PAR 19 $\mu g/mL$ and COP 1 $\mu g/mL$
- (4) M2: Solution containing PAR 18 $\mu g/mL$ and COP 2 $\mu g/mL$
- (5) M3: Solution containing PAR 17 $\mu g/mL$ and COP 3 $\mu g/mL$
- (6) M4: Solution containing PAR 16 $\mu g/mL$ and COP 4 $\mu g/mL$
- (7) M5: Solution containing PAR 15 $\mu g/mL$ and COP 5 $\mu g/mL$
- (8) M6: Solution containing PAR 14 $\mu g/mL$ and COP 6 $\mu g/mL$
- (9) M7: Solution containing PAR 13 $\mu g/mL$ and COP 7 $\mu g/mL$
- (10) M8: Solution containing PAR 12 $\mu g/mL$ and COP 8 $\mu g/mL$
- (11) M9: Solution containing PAR 11 $\mu g/mL$ and COP 9 $\mu g/mL$
- (12) M10: Solution containing PAR 10 $\mu\text{g/mL}$ and COP 10 $\mu\text{g/mL}$
- (13) M11: Solution containing PAR 27 $\mu g/mL$ and COP 3 $\mu g/mL$

Table 1 shows that at different concentration ratios of PAR and COP, all RE% values are very small ($\leq 1.6\%$), therefore, within the concentration range of PAR (10-27 µg/mL) and COP (1-10 µg/mL), with different concentration ratios, the method error for both compounds is small and acceptable. On the other hand, all calculated RSD% values are much smaller than 1/2RSD_H, so it can be concluded that the repeatability of the method is very good for laboratory mixed solutions with the investigated concentration ranges and concentration ratios of the compounds given above.

Development of actual sample analysis procedure

Sample preparation

Weigh 20 tablets, determine the average weight of each tablet (M_{mean}) , grind into fine powder and mix well. Weigh an amount of the drug powder equal to the average weight of one tablet on an analytical balance, put it into a 250 mL flask, add about 150 mL of solvent H₂O-ACN 9:1 (V/V), then put it in an ultrasonic machine for about 30 minutes, volumetrically adjust with the solvent to the mark (solution 1). Then filter solution 1 through a blue ribbon filter paper and take 10 mL of the filtered solution into a 100 mL volumetric flask and add the solvent to the mark (solution 2), take 10 mL of solution 2 into a 100 mL volumetric flask and add the solvent to the mark (solution 3), shake well to get the sample solution. To compare the results, take 20 tablets (same batch number, manufacturer, and expiration date) to the Testing Center for Drugs, Food and Cosmetics of Thua Thien Hue to analyze by standard HPLC method.

$$H(mg/tablet) = (M/m).(C.100.10.25)/1000 = (M_{mean}/m).25.C$$
 (6)

Where: C (μ g/mL): concentration of each substance determined in the sample solution. m: mass of the sample weighed for analysis (mg), M_{mean} : average weight of one tablet.

Simultaneous quantification of PAR and COP in drug samples

We conducted a determination of the amounts of PAR and COP in Effer-Paralmax® Codein tablets manufactured by BOSTON Vietnam Pharmaceutical Joint-Stock Company, located at No. 43 Street 8, Vietnam-Singapore Industrial Park, Thuan An, Binh Duong, Vietnam, with the label indicating 500 mg PAR and 30 mg COP per tablet. Batch number: 9003, date of manufacture: 25/12/2021, expiration date: 25/12/2024. The average weight of one tablet was M = 3215.0 mg. The sample was processed as described in Section 3.2.1. Accurately weighed 3215.0 mg of the drug powder and processed it into a sample solution. The sample solution was scanned in the wavelength range of 210-290 nm with a step of 0.5 nm. The CLS-Excel program was used to determine the concentrations of PAR and COP in the sample solutions. The amounts of PAR and COP in Effer-Paralmax® Codein were calculated according to equation (6). The absorption spectra of the Effer-Paralmax® Codein drug sample solutions are shown in Figure 2. The results of the analysis of Effer-Paralmax® Codein tablets are presented in Table 2.



Figure 2 Molecular absorption spectra of standard solutions of PAR and COP, and the sample solution of Effer-Paralmax® Codein.

- (1) Standard solution of PAR at 10 µg/mL
- (2) Standard solution of COP at 10 µg/mL

(3) Sample solution of Effer-Paralmax® Codein

Table 2 Quantities of PAR and COP in Effer-Paralmax® Codein tablets

Sample N ⁰	H _(PAR) (mg/tablet)	H _(COP) (mg/tablet)
MI	514.03	31.15
M2	523.90	30.30
M3	504.93	31.83
Average	514.28	31.09
Announced	500.00	30.00
RE%	2.86	3.64
RSD(%)	1.845	2.458
*1/2RSD _H	5.075	7.739

(* The values of $I/2RSD_{\rm H}$ were calculated based on the average concentration of the measured sample solutions).

The results presented in Table 2 from the simultaneous quantification of PAR and COP in Effer-Paralmax® Codeine effervescent tablets showed that the analytical method had a relative error (RE%) of less than 5%, which is in accordance with the Vietnamese Pharmacopoeia standard.¹⁵ The experimental relative standard deviation (RSD%)

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was also less than 1/2 of the Horwitz RSD (RSD_H), indicating good repeatability of the method.

Reliability of the analytical procedure

To demonstrate the reliability of the analytical procedure, its repeatability and accuracy were evaluated.

Repeatability

The results from Table 2 showed that the method had good repeatability, with RSD% values for both analytes below 2.5% and below 1/2 of RSD_H.

Accuracy

The accuracy was determined by calculating the recovery and comparing the results obtained from the study method with those obtained from the standard HPLC method.

Recovery

To determine the recovery, the sample powder mass was weighed using the average mass of one tablet. The first sample did not contain any standards, while the remaining samples contained increasing amounts of both PAR and COP standards. The samples were processed according to the procedure described in 3.2.1 to obtain the sample solution without standards and three sample solutions with added standards. The spectra of the PAR 10 µg/mL and COP 10 µg/ mL standard solutions, as well as the spectra of the sample solution without standards (S0) and the sample solutions with added standards (S1, S2, and S3), were recorded. The concentrations of PAR and COP in the sample solutions and the sample solutions with added standards were calculated using the CLS-Excel filter program. The spectra of the standard and sample solutions with added standards are presented in Figure 3, while the concentrations of the added standards and the calculated concentrations of the sample solutions without and with added standards are shown in Table 3. The recovery of the two analyte concentrations in the samples with added standards was calculated using Equation (4).



Figure 3 The molecular absorption spectra of standard solutions, sample solutions, and standard-spiked sample solutions.

I. PAR10: standard solution PAR 10 $\mu g/mL;$ 2. COP10: standard solution COP 10 $\mu g/mL$

S0: sample without added; S1: sample added standard the first time (PAR = 2,0 $\mu g/mL,$ COP = 0,5 $\mu g/mL)$

S2: sample added standard the 2sd time (PAR = 4,0 μ g/mL, COP = 1,0 μ g/mL)

S3: sample added standard 3rd time (PAR = 6,0 μ g/mL, COP = 1,5 μ g/mL)

From the results of the Rev% recovery calculation presented in Table 3, it can be seen that the recovery of the method is approximately

100% for both substances. This confirms that the analytical method has good accuracy.

 $\label{eq:stables} \begin{array}{l} \textbf{Table 3} \mbox{ Recovery of the method for Effer-Paralmax} \mbox{ Codeine effervescent tablets} \end{array}$

Sample	Substance	C _ı (µg/mL)	C ₂ (µg/mL)	C _{add} (µg/mL)	Rev (%)
<u> </u>	PAR	20.461	22.514	2.0	102.65
51	COP	1.246	1.739	0.5	98.60
62	PAR	20.461	24.568	4.0	102.68
52	COP	1.246	2.240	1.0	99.40
S3	PAR	20.461	26.550	6.0	101.48
	COP	1.246	2.730	1.5	98.93

Comparison of the analysis results of the research method with HPLC method

To continue the objective evaluation of the accuracy of the current research method, we compared the results of the determination of the amount of substance in the Effer-Paralmax® Codeine drug sample using the analytical method with the results of the standard HPLC method performed by the Drug, Food, and Cosmetics Testing Center of Thua Thien Hue province. We evaluated the results of the two methods statistically.¹⁴ The results are presented in Table 4.

 Table 4 Comparison of the analysis results of PAR and COP content

 determination in the Effer-Paralmax® Codeine drug sample using the CLS

 method and the HPLC method

H _{PAR} (mg/tablet)		H _{COP} (mg/tablet)		
CLS	HPLC	CLS	HPLC	
514.03	520.10	31.15	32.87	
523.90	527.68	30.30	32.08	
504.93	514.14	31.83	32.12	
$H_{1(mean)} = 514.28$ $t_{exp} = 0.944$ $t_{(0.05;4)} = 2.78$	H _{2(mean)} = 520.64	$H_{1(mean)} = 31.09$ $t_{exp} = 2.478$ $t_{(0.05; 4)} = 2.78$	H _{2(mean)} = 32.36	

The results obtained in Table 4 show that all t_{exp} values are smaller than t(0.05; 4), therefore, the results of the analysis of the two active ingredients PAR and COP in Effer-Paralmax® Codein tablets by the CLS method and the HPLC standard method are statistically equivalent with a significance level of $\alpha = 0.05$, and it can be said that the analysis results of the CLS method are not different from the results of the standard HPLC method.

Conclusion

With the goal of applying the UV-Vis molecular absorption spectroscopy method combined with the classic least squares (CLS) method to simultaneously determine the content of Paracetamol and Codeine phosphate in Effer-Paralmax® Codein effervescent tablets, we have achieved the following results:

Suitable conditions were studied and selected to simultaneously determine PAR and COP in laboratory mixed solutions containing the two substances with different concentration ratios using the molecular absorption spectroscopy method combined with the CLS method, specifically: Water: Acetonitrile (9:1 v/v) was chosen as the solvent for dissolution; The appropriate wavelength range for spectrum scanning is from 210 nm to 290 nm with a step of 0.5 nm. With different concentration ratios of PAR and COP in the mixed solution, the method errors were less than or equal to 2% for both PAR and COP, and the RSD% values were all small (<1.7). Therefore, the method ensures accuracy and repeatability.

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The analytical procedure for the tablet samples containing PAR and COP was established using the UV-Vis molecular absorption spectroscopy method combined with the CLS method, and the reliability of the analysis procedure was demonstrated through the analysis of Effer-Paralmax® Codeine tablets. The results of the analysis drug content had a relative error compared to the label RE% < 5%, which is comply with the quality standards of the Vietnamese Ministry of Health.¹⁵ The experimental RSD% was less than 1/2RSD_H, so the method has good repeatability. Comparing the results of determining the content of PAR and COP in Effer-Paralmax® Codeine analyzed by the research method and the standard HPLC method showed that the results of the two methods are identical at a significance level of $\alpha = 0.05$.

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None.

Data availability

The data used to support the finding of this study are available from the corresponding author upon request.

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Conflicts of interest

The authors declare that they have no conflicts of interest.

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