

# Efficiency study of silver nanoparticles (AgNPs) supported on granular activated carbon against E.Coli

## Abstract

The antibacterial effects of Ag salts have been noticed since antiquity and currently are used to control bacterial growth in a variety of applications. Recent studies have revealed that the antimicrobial properties of silver are due to its ionized form, Ag<sup>+</sup>, and its ability to cause damage to cells by interacting with thiol-containing proteins and DNA. Silver nanoparticles are a form of silver of particular interest because of their easy production, high antimicrobial activity, and ability to be incorporated into a diverse range of products. We report the antibacterial properties of silver nanoparticles (AgNPs) supported on granular activated carbon (GAC). We prepare two types of GAC by using 60% (w/v) KOH and 85% (v/v) H<sub>3</sub>PO<sub>4</sub> solutions respectively, as activating agent. AgNPs were synthesized by GAC impregnation with 0.1M, 0.05M and 0.01M AgNO<sub>3</sub> solutions. Characterization was done using UV/V is spectrophotometry, Scanning Electron Microscopy (SEM) and adsorption isotherm data was used to determine the pore size distribution and surface area. Obtained materials showed nearly 100% antibacterial activities (elimination of microorganisms) against *Escherichia coli* because of the presence of the silver nanoparticles. Activity of the resulting material was related to the concentration of impregnated Ag and the surface area. Probably a high concentration of AgNPs on the GAC surface is more effective to attach to cell membrane and disrupt the permeability and respiration functions of the cell and thus kill the *Escherichia coli*.

**Keywords:** Silver nanoparticles, *Escherichia coli*, Activated carbon, Antimicrobial

**Abbreviations:** AC, Activated Carbon; GAC, Granulated Activated Carbon; AgNPs, Silver Nanoparticles; E. coli, *Escherichia coli*; SEM, Scanning Electron Microscope; EDX, Energy Dispersive X-ray; CFU, Colony Forming Units; TSA, Trypticase Soy Agar; EMB, Eosin Methylene Blue Agar; DNA, Deoxyribonucleic acid

## Introduction

Activated carbon (AC) is widely used in drinking water treatment to adsorb natural organic compounds, taste and odor compounds, and synthetic organic chemicals. Adsorbent property is because it is a highly porous material and provides a large surface area to which contaminants may adsorb. This is due to both the physical and chemical process of accumulating a substance at the interface between liquid and solids phases. Since AC filters will not remove microbial contaminants (such as bacteria and viruses), calcium and magnesium (hard water minerals), fluoride, nitrate, and many other compounds, combined methods are necessary to effectively treat the water. Previous studies have shown that antimicrobial formulations in the form of nanoparticles could be used as effective bactericidal materials.<sup>1-4</sup> Indeed, nanosized inorganic particles, of either simple or composite nature, display unique physical and chemical properties and represent an increasingly important material in the development of novel nano devices which can be used in numerous physical, biological, biomedical, and pharmaceutical applications.<sup>5-7</sup> A number of recent achievements offer the possibility of generating new types of nanostructure materials with designed surface and structural properties.<sup>7-10</sup>

It is well known that silver ions and silver-based compounds are highly toxic to microorganisms<sup>7-11</sup> showing strong biocidal effects on as many as 16 species of bacteria including E. coli.<sup>12-15</sup> Biocide mechanism is still under research. Some authors report that Ag

Volume 1 Issue 2 - 2014

Sabrina Acevedo,<sup>1</sup> Jose Arevalo-Fester,<sup>1</sup> Lissa Galicia,<sup>1</sup> Reinaldo Atencio,<sup>2</sup> Eric Plaza,<sup>3</sup> Eduardo Gonzalez<sup>4</sup>

<sup>1</sup>Unit of Characterization and Structure of Materials, Zulian Institute for Technological Research - INZIT, Venezuela

<sup>2</sup>Laboratory of Materials and Emerging Technologies, Venezuelan Institute of Scientific Research-IVIC, Venezuela

<sup>3</sup>Laboratory of Fluids and Plasma, Venezuelan Institute for Scientific Research - IVIC, Venezuela

<sup>4</sup>Institute of Surface and Catalysis, University of Zulia, Venezuela

**Correspondence:** Jose Arevalo-Fester, Zulian Institute for Technological Research - INZIT, Km 15 La Canada de Urdaneta, Zulia State, Venezuela, Tel 58-4160164864  
 Email: joseduardo27@gmail.com

**Received:** October 26, 2014 | **Published:** November 22, 2014

inactivates microbes by interacting with their enzymes, proteins or DNA to restrain cell proliferation or cell division. It also binds to the negatively-charged bacterial cells to change the functionality of the cell membrane, thereby preventing bacterial regeneration.<sup>13,16-17</sup> Thus, registration of biocidal silver and nanosilver products has increased dramatically over the last ten years, most likely as a result of improved capabilities in nanoscience and engineering that allow silver nanoparticles (AgNPs) to be formulated to confer increased durability and/or sustained antibacterial action, even under harsh environmental conditions.<sup>18</sup> Based on these facts, we prepare granular activated carbon impregnated with silver nanoparticles (AgNPs-GAC), in order to determinate their antibacterial activity (elimination of microorganisms) against *Escherichia coli*.

## Material and methods

### Material

Phosphoric acid (H<sub>3</sub>PO<sub>4</sub>, 85% v/v) silver nitrate (AgNO<sub>3</sub>) and Trypticase Soy Agar (TSA) were purchased from Merck. Potassium hydroxide (KOH) and ammonium hydroxide (NH<sub>4</sub>OH) were obtained from Riedel de Haen and Fluka respectively. Eosin Ethylene Blue Agar Medium (EMB) was purchased from Himedia. *Escherichia coli* (E. coli) used in this research was ATCC 35218. For microscopic characterization a Scanning Electron Microscope (SEM), FEI Quanta 200 was employed. Surface area SBET was determined using a Micromeritics ASAP 2020 system. UV/Vis spectra were obtained by using a Biochrom Light wave II Spectrophotometer. Bituminous mineral coal was obtained from Carbonesdel Guasare (Venezuela).

### GAC preparation

GAC adsorbents were prepared according to literature method, starting from mineral coal. For GAC-1, concentrated (85% v/v)

H<sub>3</sub>PO<sub>4</sub>, was used as activating agent, while GAC-2 was prepared using a (60% v/v) KOH solution. Pyrolysis was carried out for 2 hours at 500°C and 1 hour at 700°C, respectively.<sup>19</sup>

### Nanoparticles impregnation

For GAC impregnation with silver nanoparticles, aqueous solutions (0.1M, 0.05M and 0.01M) of AgNO<sub>3</sub> were prepared. In three 100 mL amber Erlenmeyers 1 g of GAC-1, was weighted and 20 mL solutions of AgNO<sub>3</sub> at given concentrations were added in each Erlenmeyer, followed by 2 mL of concentrated NH<sub>4</sub>OH as Ag reducing agent (Tollens reaction). All samples were stirred for 24 hours and then filtered and washed with distilled water and finally dried at 120°C for 4 hours. The same procedure was used for GAC-2.<sup>10,20-22</sup>

**Table 1** Surface areas and pore volumes

GAC	[Ag] (mol/L)	S <sub>BET</sub> (m <sup>2</sup> /gr)	V <sub>tpore</sub> (cm <sup>3</sup> /gr)	V <sub>micro</sub> (cm <sup>3</sup> /gr)	V <sub>meso</sub> (cm <sup>3</sup> /gr)	Ag <sup>2</sup> % (w/w)
<b>GAC-1<sup>a</sup></b>	0.00	528.25±6.123	0.373	0.086	0.286	ND
<b>(AgNPs)GAC-1<sup>a</sup></b>	0.01	423.22±8.717	0.214	0.125	0.089	7.49
<b>(AgNPs)GAC-2<sup>a</sup></b>	0.05	369.27±10.826	0.205	0.111	0.093	23.62
<b>(AgNPs)GAC-3<sup>a</sup></b>	0.10	309.04±6.573	0.155	0.093	0.061	29.56
<b>GAC-2<sup>b</sup></b>	0.00	1386.10±19.001	0.729	0.395	0.395	ND
<b>(AgNPs)GAC-4<sup>b</sup></b>	0.01	698.62±15.777	0.364	0.238	0.125	8.86
<b>(AgNPs)GAC-5<sup>b</sup></b>	0.05	673.74±15.142	0.378	0.225	0.143	27.16
<b>(AgNPs)GAC-6<sup>b</sup></b>	0.10	615.16±16.048	0.305	0.240	0.064	42.37

SBET1 :Values are means ± SD of triplicate determinations; Ag2: Elemental analysis was determined by Energy-Dispersive X-ray Spectroscopy (EDX); aActivated with H<sub>3</sub>PO<sub>4</sub>; bActivated with KOH, ND: Not Detected

### Minimum contact time test

The minimum contact time was determined as follows: a 250 mL Erlenmeyer containing 100 mL of TSA was inoculated with pure culture of *E. coli* and incubated at 37°C for 18 hours, until reaching a concentration of 10<sup>8</sup> CFU/mL, as determined by UV/Vis spectrophotometry at 600 nm wavelength. After that, 1 gram of (AgNPs) GAC was added and kept under stirring for 60 min. These assays were performed in triplicate for each type of (AgNPs) GAC. Each 5 min, 1 mL of extract was taken and grown in EMB agar medium and incubated at 37°C for 24 hours. CFU was calculated using equation 1.<sup>24</sup>

$$\frac{(\text{No.ofcolonies}) \times (\text{dilutionfactor})}{(\text{volumeofcultureplate})} = \frac{\text{CFU}}{\text{ml}}$$

## Results and discussion

### GAC characterization

Surface areas and pore volumes of the GAC materials activated with H<sub>3</sub>PO<sub>4</sub> and KOH and their corresponding AgNPs are shown in (Table 1). The Tollen's synthesis method gives silver nanoparticles with a controlled size in a one-step process.<sup>25</sup> The basic Tollen's method involves reaction 2, where Ag<sup>+</sup> ions are reduced by aldehydes or carboxylates which are present on the GAC surface, in the presence of ammonia, yielding AgNPs.

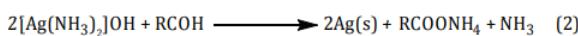


Table 1, shows a relationship between Ag concentration in the impregnation solution and surface area SBET. The BET surface area diminishes when increasing the Ag content in the impregnation

### Characterization

Electron microscopy characterization was performed and obtained images were analyzed in order to observe the presence of AgNPs on the GAC surface. Elemental analysis was done by using Energy Dispersive X-ray technique (EDX), included in the SEM configuration. Nitrogen adsorption isotherms were obtained at 77K and surface area S<sub>BET</sub> was determined by mean of Brunauer, Emmert and Teller method.<sup>23</sup>

### Bacteriological analysis

Efficiency of (AgNPs)GACs was analyzed using serial dilution technique in Petri dish. Trypticase Soy Agar (TSA) was used for inoculation of pure culture of *Escherichia coli*.

solution until the maximum saturation level is reached. A decreasing of pore volume is also observed due to Ag dispersion along all pore range without any specific size limitation.<sup>25-27</sup>

### SEM characterization

SEM micrographs presented in (Figures 1 & 2) corroborate that the AgNPs content supported on the carbon matrix is related with the concentration of the impregnation solution and the surface area of the GACs. This fact was also corroborated by elemental analysis as shown in Table 1. Since nature and concentration of the reductant must play a major role in controlling the silver nanoparticle size, it was assumed that the higher surface area determined in GACs activated with KOH (higher concentration of reductant groups) favors the primary formation of a large number of small clusters with a low coagulation barrier; their further irreversible aggregation was responsible for the presence of coarser agglomerates as observed in (Figures 1A-1D & 2A-2D).<sup>28-29</sup>

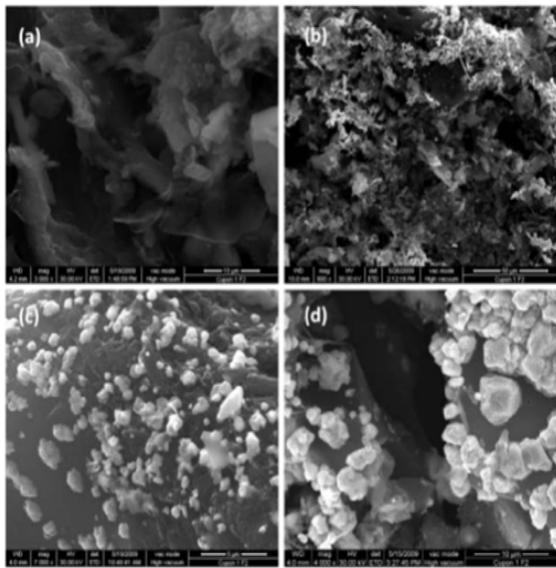
### Antibacterial efficacy results

Antibacterial inhibition test is shown in Figure 3. GACs used in this study were found to have various degrees of antimicrobial effects against microorganisms tested. GAC activated with H<sub>3</sub>PO<sub>4</sub> and KOH (GAC-1 (Figure 3A-3D) and GAC-2 (Figure 3E-3H), respectively) were not effective as antibacterial material as is also observed in Figure 4 & 5. These results also demonstrated that these materials were not capable to reduce significantly *E. coli*, without aid of AgNPs.

In Figure 4, *E. coli* concentration decrease when Ag nanoparticles in GAC are increased. For (AgNPs)GAC-3 (%Ag: 29.56 w/w), total *E. coli* was eliminated after 25 min of stirring. The same behavior was observed for (AgNPs) GAC-2 (%Ag: 23.62 w/w), while the GAC with the less quantity of Ag nanoparticles, (AgNPs) GAC-1 (%Ag: 7.49 w/w) was able to reduce *E. coli* after 45 min from t0: 16×10<sup>7</sup> UFC/

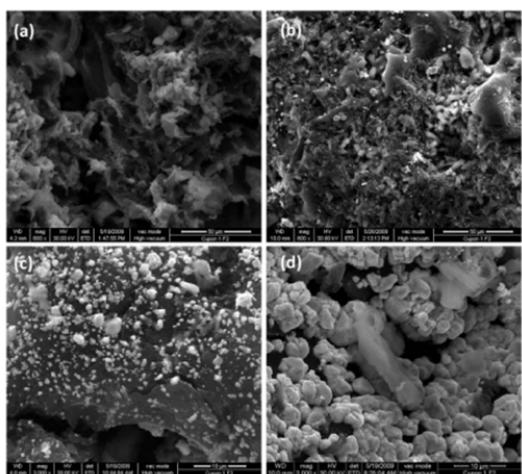
mL to  $96 \times 10^5$  UFC/mL. After that, bacterial concentration remains constant until the end of the assay (60 min).

All (AgNPs) GACs prepared from GAC activated with KOH were in general, more time-efficient to eliminate *E. coli*, as shown in (Figure 5). For (AgNPs)GAC-6 (%Ag: 42.37 w/w), *E. coli* concentration decreased dramatically after 10 min with a final minimum contact time of 15 min. The same behavior was observed for (AgNPs)GAC-5 (%Ag: 27.16 w/w), with a minimum contact time of 25 min. Finally, after 30 min of stirring, (AgNPs)GAC-4 (%Ag: 8.86 w/w), was able to eliminate *E. coli*.



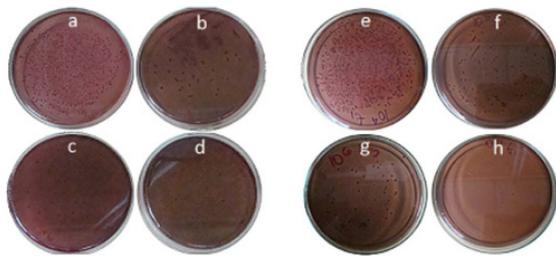
**Figure 1** SEM micrographs of  $H_3PO_4$  activated GACs.

- a. GAC-1
- b. (AgNPs) GAC-1
- c. (AgNPs) GAC-2
- d. (AgNPs) GAC-3



**Figure 2** SEM micrographs of KOH activated GACs.

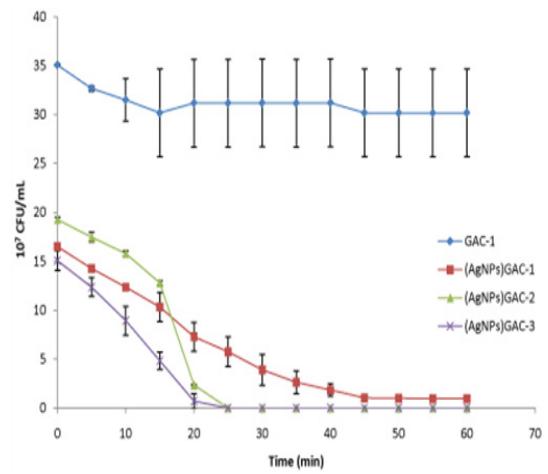
- a. GAC-2
- b. (AgNPs) GAC-4
- c. (AgNPs) GAC-5
- d. (AgNPs) GAC-6



**Figure 3** Antibacterial inhibition test of GACs and AgNPs GACs against *E. Coli*

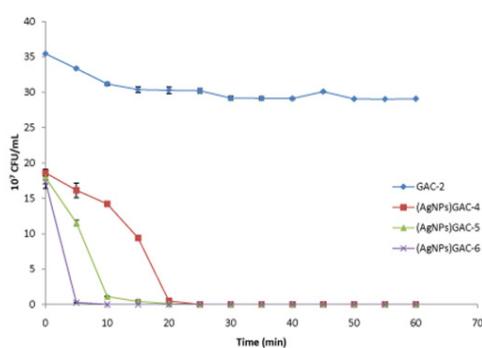
- a. GAC-0
- b. AgNPsGAC-1
- c. AgNPsGAC-2
- d. AgNPsGAC-3
- e. GAC-1
- f. AgNPsGAC-4
- g. AgNPsGAC-5
- h. AgNPsGAC-6

The results of this study showed that (AgNPs) GACs were able to eliminate *E. coli*. The elimination rate depends on the quantity of Ag adsorbed on GAC.<sup>3</sup> Obviously, the adsorption phenomenon is related to surface area of the material. In this sense, GAC activated with KOH and impregnated with 0.1M  $AgNO_3$  solution, were more effective material for *E. coli* elimination, under employed experimental conditions. This results are in agreement with the above-mentioned antimicrobial properties of Ag, which is believed that the interaction between positively charged Ag ions and negatively charged bacterial membranes plays a major role in causing the membrane rupture and cell lysis. Researchers also reported that  $Ag^+$  prevents DNA replication or involves its interaction with thiol groups of proteins, blocking and inactivating of respiratory enzymes.<sup>30-32</sup>



**Figure 4** Activity of GAC activated with  $H_3PO_4$  and their (AgNPs) GACs at different Ag concentrations against *E. coli*

*Coli*. Ag content in (%(w/w)): GAC-1: 0; (AgNPs)GAC-1: 7.49; (AgNPs)GAC-2: 23.62; (AgNPs)GAC-3: 29.56. Values are the average of three individual replicates and error bars represent standard deviation.



**Figure 5** Activity of GAC activated with KOH and their (AgNPs) GACs at different Ag concentrations against *E. coli*.

Coli. Ag content in % (w/w): GAC-2: 0; (AgNPs)GAC-4: 8.86; (AgNPs)GAC-5: 27.16; (AgNPs)GAC-6: 42.37. Values are the average of three individual replicates and error bars represent standard deviation.

## Conclusion

The fact that the number of researches devoted to antibacterial and antiviral activity of AgNPs increases every year confirms the high interest in both synthesis and properties of silver nanoparticles. In this research we found that antimicrobial activity might be dependent on the surface area of GACs, AgNPs concentration and nanoparticles size as well as bacterial colony forming units (CFU). The obtained results from the (AgNPs) GACs assays demonstrate that they were effective as antibacterial against *E. coli* over a short period of time. In this sense, GAC activated with KOH and impregnated with 0.1M AgNO<sub>3</sub> solution AgNPsGAC-6 showed the higher AgNPs content (42.37 wt %). For this reason, it was able kill completely *E. coli* strain (from 10<sup>7</sup> CFU/mL to 0 CFU/mL) after 15 min of stirring. Nevertheless, in this work, experimental conditions employed are rarely found in real systems, it is possible to predict based on physical, chemical and biological properties of both GACs and AgNPs, that they could be combined to produce nanotechnology based devices for practical applications such as water treatment and recycling.

## Acknowledgements

We acknowledge the financial support of FONACIT. (Project: 201100139).

## Conflicts of interest

None.

## References

- Pal S, Tak YK, Song JM. Does the antibacterial activity of silver nanoparticles depend on the shape of the nanoparticle? A study of the Gram-negative bacterium *Escherichia coli*. *Appl Environ Microbiol*. 2007;73(6):1712–1720.
- Choi O, Deng KK, Kim NJ, et al. The inhibitory effects of silver nanoparticles, silver ions, and silver chloride colloids on microbial growth. *Water Res*. 2008;42(12):3066–3074.
- Park HJ, Kim JY, Kim J, et al. Silver-ion-mediated reactive oxygen species generation affecting bactericidal activity. *Water Res*. 2009;43(4):1027–1032.
- Silver S, Phung LT, Silver G. Silver as biocides in burn and wound dressings and bacterial resistance to silver compounds. *J Ind Microbiol Biotechnol*. 2006;33(7):627–634.
- Balashanmugam P, Caral Dinesh R, Manivasagan V, et al. Extracellular biosynthesis of silver nanoparticles using *Cassia fistula* extract and in-vitro antimicrobial studies. *J Pharm Res*. 2014;8(2):187–191.
- Patel D, Patel M, Krishnamurthy R. Silver nanoparticles: biosynthesis and its antimicrobial activity. *Cibtech J Bio-Protocols*. 2013;2(1):50–57.
- Sondi I, Salopek-Sondi B. Silver nanoparticles as antimicrobial agent: a case study on *E. coli* as a model for Gram-negative bacteria. *J Colloid Interface Sci*. 2004;275(1):177–182.
- Srivastava R, Yadav BC. Nanostructured ZnO, ZnO-TiO<sub>2</sub> and ZnO-Nb<sub>2</sub>O<sub>5</sub> as solid state humidity sensor. *Adv Mater Lett*. 2012;3(3):197–203.
- Mattoussi H, Matthew M, Goldman R, et al. Self-Assembly of CdSe – ZnS quantum dot bioconjugates using an engineered recombinant protein. *J Am Chem Soc*. 2000;122(49):12142–12150.
- Jibril B, Al-maamari RS, Houache O, et al. Effects of H<sub>3</sub>PO<sub>4</sub> and KOH on pyrolysis of bituminous coal in preparation of activated carbon. *J Appl Sci Res*. 2007;3(11):1343–1351.
- Sondi I, Fedynyshyn TH, Sinta R, et al. Encapsulation of nanosized silica by in situ polymerization of tert –butyl acrylate monomer. *Langmuir*. 2000;16(23):9031–9034.
- Rai M, Yadav A, Gade A. Silver nanoparticles as a new generation of antimicrobials. *Biotechnol Adv*. 2009;27(1):76–83.
- Bae E, Park HJ, Lee J, et al. Bacterial cytotoxicity of the silver nanoparticle related to physicochemical metrics and agglomeration properties. *Environ Toxicol Chem*. 2010;29(10):2154–2160.
- Maillard J, Hartemann P. Silver as an antimicrobial: facts and gaps in knowledge. *Crit Rev Microbiol*. 2012;39(4):373–383.
- Fayaz AM, Balaji K, Girilal M, et al. Biogenic synthesis of silver nanoparticles and their synergistic effect with antibiotics: a study against Gram-positive and Gram-negative bacteria. *Nanomedicine*. 2010;6(1):103–109.
- You J, Zhang Y, Hu Z. Bacteria and bacteriophage inactivation by silver and zinc oxide nanoparticles. *Colloids Surf B Biointerfaces*. 2011;85(2):161–167.
- Gogoi SK, Gopinath P, Paul A, et al. Green fluorescent protein-expressing *Escherichia coli* as a model system for investigating the antimicrobial activities of silver nanoparticles. *Langmuir*. 2006;22(22):9322–9328.
- Hong KH. Preparation and properties of electrospun poly(vinyl alcohol)/silver fiber web as wound dressings. *Polym Eng Sci*. 2007;47:43–49.
- Murty BS, Shankar P, Baldev R, et al. Textbook of Nanoscience and Nanotechnology. Springer, Berlin, Germany. 2013;pp.256.
- Le Pape H, Solano-Serena F, Contini P, et al. Evaluation of the antimicrobial properties of an activated carbon fiber supporting silver using a dynamic method. *Carbon*. 2001;40(15):2947–2954.
- Bandyopadhyaya R, Venkata S, Shankar P. Silver-embedded granular activated carbon as an antibacterial medium for water purification. *J Chem Technol Biotechnol*. 2008;83(8):1177–1180.
- Le Pape H, Solano-Serena F, Contini P, et al. Involvement of reactive oxygen species in the bactericidal activity of activated carbon fiber supporting silver bactericidal activity of ACF(Ag) mediated by ROS. *J Inorg Biochem*. 2004;98(6):1054–1060.
- Fagerlund G. Determination of specific surface by the BET method. *Materiaux et Construction*. 1973;6(33):239–245.
- Gamazo C, Lopez-Gani I, Diaz R. Manual practico de microbiologia. Elsevier, Spain. 2005;pp.231.
- Landage SM, Wasif A I. Nanosilver – an effective antimicrobial agent for Finishing of textiles. *Int J Eng Sci Emerg Technol*. 2012;4(1):66–78.

26. Marsh H, Rodriguez F. Activated Carbon. Elsevier Science & Technology Books. London, UK, 2006;536.
27. Webb P, Orr C. Analytical Methods in fine particle technology. Micrometrics Instrument Corp, USA. 2007.
28. Kvitek L, Panacek A, Soukupova J, et al. Effect of surfactants and polymers on stability and antibacterial activity of silver nanoparticles (NPs). *J Phys Chem.* 2008;112(15):5825–5834.
29. Le AT, Huy PT, Tam PD, et al. Green synthesis of finely-dispersed highly bactericidal silver nanoparticles via modified Tollens technique. *Curr Appl Phys.* 2010;10(3):910–916.
30. Zhang S, Fu R, Wu D, et al. Preparation and characterization of antibacterial silver-dispersed activated carbon aerogels. *Carbon.* 2004;42(15):3209–3216.
31. Feng QL, Wu J, Chen GQ, et al. A mechanistic study of antibacterial effects of silver ions on *Escherichia Coli* and *Staphylococcus aureus*. *J Biomed Mater Res.* 2000;52(4):662–668.
32. Garcia-Barrasa J, Lopez J, Mongue M. Silver nanoparticles: synthesis through chemical methods in solution and biomedical applications. *Cent Eur J Chem.* 2001;9(1):7–19.