Fungal communities in *Macaca fascicularis* and *Macaca nemestrina* associates with forest matrix types in West Malaysia

**Abstract**

Genetic and geographical proximity between *Macaca fascicularis*, *Macaca nemestrina* and humans are posing a dangerous threat in terms of disease transmission to humans. The objective is to determine the fungal species in actively shedding anatomical sites of these macaques inhabiting forest fragment of different matrix types in Kemasul Forest Reserve, Pahang State, Malaysia. Matrix of Chemomoi (CM) had higher coverage of acacia (*Acacia mangium*), whereas JambuKias (JR) encompassed oil palm plantation. Swab samples were obtained from throat, anus and penis/vagina of three individual of macaques from each species and geographic location (N=12). The swabs were then streaked onto Potato Dextrose Agar plate containing 25 mg/ml chloramphenicol to obtain pure colony. The isolated fungal species were identified morphologically, and molecular identification was carried out in some of the isolates. The results reveal 18 fungal species from 10 genera identified, with JR showing 13 species compared to CM with only 7 species, but significant difference was observed between fungal distribution in different geographical locations and macaques species (p<0.05). Fungal species from both macaque species were higher in JR compared to CM but no significant difference in different anatomical sites. The most common fungi species found in JR is *Aspergillus fumigatus*, as it was found in all anatomical sites of both macaques but it was absent from samples in CM. Only two fungi species shared between both geographic locations which are *A. ruber* and *A. flavus*. This study reveals potential risk to both human-macaque, thus relocating the human settlements adjacent to the forests and proper waste management system is essential.

**Keywords:** actively shedding fungi, macaques, forest plantations, Malaysia

**Introduction**

Increasing human population has resulted in a higher number of human intrusion in wildlife habitat, leading to human–primates (NHPs) conflicts. Twig\(^1\) also stated that pest behavior among NHPs have been observed in urban environment as they raid trash for food. Dumping of rubbish from human food near their habitat is also one of the reason for increased human–primate infringement.\(^2\)

The declining habitats force macaques to migrate to near human settlements. Due to the lack of food source from the forest caused by deforestation, macaques quickly learn to raid crops or garden from agriculture land. These raid incidents increase the chances of human-macaque interactions that can lead to other incidents. *Macaca fascicularis* and *M. namestrina* are susceptible to different dangerous biological agents that are transmittable to human due to the close proximity in genetics and geography. According to Pederson & Davies,\(^2\) cross species disease transmission is caused by three major factors and one of them is overlapping geographical distribution. Besides that, biological factors such as genetic similarity highly determine a successful transmission and infection.\(^1\) Close genetic relationships among host species might translate into similar immunological responses.\(^3\) As a consequence macaques living in such close distance with humans, are serving as a main source of disease transmission. While *M. fascicularis* is not currently a species of concern on the International Union for Conservation of Nature (IUCN) list, whereas *M. nemestrina* is considered ‘Vulnerable’.\(^4\)

Current knowledge of fungal diseases in macaques in relation to habitat disturbance is somewhat patchy, as many studies focus on specific case study. The closest study by Carvalho et al.,\(^7\) on nasal, oral and rectal of black lion tamarins (*Leontopithecus chrysopygus*) found that mycelium fungi were the most frequently found fungal species from nasal and oral area while yeast fungus were found from anal samples. Mycelium fungi isolated include *Aspergillus*, *Candida*, *Penicillium* and *Trichoderma*. *Aspergillus conidia* can be easily inhaled into the human lungs causing various diseases such as invasive pulmonary aspergillosis, aspergillosima, and can lead to hypersensitivity diseases including allergic asthma, hypersensitivity pneumonitis, and allergic broncho pulmonary aspergillosis.\(^8\) Frequent interaction between humans and macaques can increase the risk of fungal spores’ transmission. Besides fungal infection diseases, humans are also at risk of toxin produced by *Aspergillus* genus known as mycotoxin and aflatoxin.\(^7\)

Humans who feed and interact with the macaques can either be exposed to zoonotic diseases after being bitten or scratched such as simian foamy virus.\(^6\) According to Jones et al.,\(^6\) there is potential for previously unknown diseases transferred from the monkeys to humans who were feeding them, posing a serious health risk to the surrounding communities.\(^8\)

Chikungunya Virus (CHIKV) has caused epidemics worldwide of rash, fever and althralgia in the last decade.\(^9\) Recent studies have revealed the presence of this virus in macaques in Malaysia, especially in Perak and Kuala Lumpur.\(^10\) In Northern Thailand, a number of viruses were found among *M. namestrina*.\(^11\) The viruses include Dengue virus, Japanese Encephalitis and Chikungunya virus making *M. namestrina* a very harmful virus reservoir to human as they live in close proximity.
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According to Demanche et al.,\(^{12}\) *Pneumocystis carinii* is one of the most regular fungal pathogen infecting primates. They also stated that the same agent can cause *Pneumocystis carinii* pneumonia (PCP), which is one of the most critical fungal respiratory infections that can develop in immune compromised patients. A case report was written regarding an infection of *Coccidioides* spp. resulting in disseminated coccidioidomycosis that was later identified to be the cause of impaired locomotion ability in a 5-year, 10-month-old male *M. fascicularis*.\(^{13}\) Symptoms found in the infected macaque include Pyogranulomatous inflammation and spherules in the tissue. *Coccidioides* spp. is a pathogenic fungus usually found in the soil of some region in United States, Central and South America and also Mexico. This fungal species naturally infects wild rodents and they can be isolated in high concentration from the rodents’ habitat.\(^{13}\) *Coccidioides* spp. will grow into vegetative state after some periods of rain, and dispersed under dry and windy conditions.\(^{14}\)

We want to determine the fungal species distribution in different anatomical sites of macaques, in relation to habitat condition. The macaques have been reported encroaching near human settlements in Kemasul Forest Reserve, Pahang. The surrounding forest area was demolished and substituted with acacia, oil palm and rubber plantation. Chemomoi (CM) and Jambu Rias (JR) was an extended forest fragment, with different surrounding matrix types. CM were surrounded by Acacia plantation (*Acacia mangium*), which is a rapidly growing plant species, whereas JR were surrounded with oil palm plantation. Oil palm plantations were known as disturbed habitat due to the manipulation of natural forest with the use of pesticides, human interference and invasion of foreign species, making it a incongruous habitat for wildlife. We hypothesize forest with oil palm matrix would yield a higher fungal infection due to its more disturbed nature as more fungal spores were produce from the soil and the plant itself. *M. fascicularis* is more adapted and lives close to human settlements, therefore we anticipate that this species exhibit higher fungal distribution, compared to *M. namestrina*. We also expect different fungal isolates in different anatomical sites.

### Materials and methods

#### Study areas

Kemasul Forest Reserve is located between three districts in Pahang State of Peninsular (West) Malaysia (Figure 1) which are Bera, Temerloh and Bentong. Intense deforestation for commercial plantations has taken place in the forest reserve, leaving forest fragments scattered in the landscape. Two geographic locations within Kemasul Forest Reserve (FR) were chosen for sampling were JambuRias (JR) and Chemomoi (CM) as shown in Figure 2. Both locations differ in matrix types, where JR is surrounded by mixture of oil palm and rubber plantation, whereas CM is surrounded by acacia plantation (*Acacia mangium*).
Sampling of macaques

Capture-release method for *M. fascicularis* and *M. nemestrina* was employed in this study between August and November 2015 at both JR and CM. Macaques were captured using 30 large wire mesh live cage traps (38x38x106 cm). Traps were deployed in 6 transect lines of 50 m length which located at different location in the forest. Jackfruit was used as bait due to its strong aromatic smell. The captured macaques were then anesthetized using 2-3ml of Zoletil to collect the samples. Sterile swabs were used to collect samples in three anatomical sites: anus, throat and penile-vaginal. The swabs were then stored in a tube containing lysis buffer for preservation and kept within liquid nitrogen before being transported back to the Environmental Microbiology Laboratory in Universiti Kebangsaan Malaysia (UKM), Bangi. List of individuals captured is listed in Table 1.

Table 1  Samples from individuals captured according to geographic location, sex and age

<table>
<thead>
<tr>
<th>Site</th>
<th>Species</th>
<th>ID</th>
<th>Sex</th>
<th>Age</th>
</tr>
</thead>
<tbody>
<tr>
<td>JR</td>
<td><em>M. fascicularis</em></td>
<td>WRG 0008</td>
<td>F</td>
<td>A</td>
</tr>
<tr>
<td></td>
<td></td>
<td>WRG 0009</td>
<td>F</td>
<td>A</td>
</tr>
<tr>
<td></td>
<td></td>
<td>WRG 0013</td>
<td>M</td>
<td>A</td>
</tr>
<tr>
<td></td>
<td><em>M. nemestrina</em></td>
<td>WRG 0001</td>
<td>M</td>
<td>A</td>
</tr>
<tr>
<td></td>
<td></td>
<td>WRG 0002</td>
<td>M</td>
<td>A</td>
</tr>
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<td></td>
<td></td>
<td>WRG 0006</td>
<td>F</td>
<td>SA</td>
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<tr>
<td></td>
<td></td>
<td>WRG 0016</td>
<td>F</td>
<td>J</td>
</tr>
<tr>
<td>CM</td>
<td><em>M. fascicularis</em></td>
<td>WRG 0018</td>
<td>M</td>
<td>A</td>
</tr>
<tr>
<td></td>
<td></td>
<td>WRG 0027</td>
<td>F</td>
<td>SA</td>
</tr>
<tr>
<td></td>
<td><em>M. nemestrina</em></td>
<td>WRG 0017</td>
<td>F</td>
<td>A</td>
</tr>
<tr>
<td></td>
<td></td>
<td>WRG 0028</td>
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<tr>
<td></td>
<td></td>
<td>WRG 0030</td>
<td>F</td>
<td>A</td>
</tr>
</tbody>
</table>

(Sex: M=male, F=female; Age: A=adult, SA=sub-adult, J=juvenile)

Ethical note

All procedures were approved by UKM Animal Ethic Committee with approval number FST/2016/SUKOR/18-MAY/750-MAY-2016-SEPT.-2018-AR-CATS. All captured individuals were released at the point of capture after they gained consciousness.

Determination of fungal species

Three individuals were chosen for *M. fascicularis* and *M. nemestrina* sampled at both study areas (N=12). Swabs from all three anatomical sites of each individual were streaked on Potato Dextrose (PDA) agars infused with Chloramphenicol 25mg/mL then poured onto 9 cm petri dish, before incubation at 30°C under both aerobic and anaerobic condition for 30 days. Sub-cultured fungal isolates were incubated for 14 days or until full sporulation. Control plates were incubated along with the swabbed plates to ensure aseptic condition with no fungal contamination from media/air/incubators. Lactophenol cotton blue staining method was used for morphological identification of fungal species under light microscope at 100× magnification. Identifications of fungal species were based on Howard et al. and Germain & Summerbell. For presumed Basidiomycete isolates, molecular identification was carried out due to inadequate sporulation. The DNA of fungal isolates were extracted and the fragments of ITS region of tRNA gene was amplified by PCR before sequencing followed by fungal barcoding protocol according to Raja et al. Results from the nucleotide sequences were compared with the reference sequences available in National Centre for Biotechnology Information (NCBI) database using the Basic Local Alignment Search Tool (BLAST) algorithm.

Data analysis

Mean and standard error of number of species isolated from different anatomical sites were calculated for each geographic location, according to macaques’ species. Normality test performed showed a non-normal distribution between number of fungal species and each variables (Shapiro-Wilk p<0.05), therefore, we used non-parametric test. Mann-Whitney U test was used to determine the differences in number of fungi species between the two macaques species, as well as between two geographical locations. Kruskal-Wallis H test was used to determine the differences in number of fungal species between different anatomical sites.

Results

Determination of fungal species

A total of 18 species of fungal including both yeast and mycelium fungi were identified from all individuals of *M. fascicularis* and *M. nemestrina* from JR (N=6) and CM (N=6) areas. All 18 isolates were identified under genera *Aspergillus* (6 species), *Penicillium* (3 species), *Paeclomyces* (2 species), *Sporothrix* (1 species), *Geotrichum* (1 species), *Curvularia* (1 species), *Candida* (1 species), *Phellinus* (1 species), *Basidiomycota* (1 species) and *Phebia* (1 species) as seen in Table 2 and Table 3. Figure 3(a-s) illustrate photos of each fungus isolates.

Table 2  Fungal species identified in Jambu Rias (JR)

<table>
<thead>
<tr>
<th>Macaca species</th>
<th>Macacofascicularis</th>
<th>Macacanemestrina</th>
</tr>
</thead>
<tbody>
<tr>
<td>Fungi/Anatomical sites</td>
<td>Anus</td>
<td>Throat</td>
</tr>
<tr>
<td>Aspergillus clavatus</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>Aspergillus niger</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Aspergillus fumigatus</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>Aspergillus candidus</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Aspergillus ruber</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Aspergillus flavus</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Curvularia lunata</td>
<td>+</td>
<td>-</td>
</tr>
<tr>
<td>Sporothrix schencki</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Candida parapsilosis</td>
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</tr>
</tbody>
</table>
Table continue

<table>
<thead>
<tr>
<th>Macaca species</th>
<th>Macaca fascicularis</th>
<th>Macaca nemestrina</th>
</tr>
</thead>
<tbody>
<tr>
<td>Fungi/Anatomical sites</td>
<td>Anus</td>
<td>Throat</td>
</tr>
<tr>
<td>Penicillium jenseni</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Penicillium frequentans</td>
<td>-</td>
<td>+</td>
</tr>
<tr>
<td>Phellinus sp.</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Basidiomycote sp.</td>
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</tr>
</tbody>
</table>

Table 3: Fungal species identified in Chemomoi (CM)

<table>
<thead>
<tr>
<th>Macaca species</th>
<th>Macaca fascicularis</th>
<th>Macaca nemestrina</th>
</tr>
</thead>
<tbody>
<tr>
<td>Fungi/Anatomical sites</td>
<td>Anus</td>
<td>Throat</td>
</tr>
<tr>
<td>Aspergillus ruber</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Aspergillus flavus</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Paecilomyces lilacinus</td>
<td>+</td>
<td>-</td>
</tr>
<tr>
<td>Paecilomyces variotii</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Geotrichum candidum</td>
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<td>-</td>
</tr>
</tbody>
</table>

Comparisons of fungal species distribution in different macaques’ species and geographical locations

Figure 4 illustrates fungal species distribution in different geographic locations, in macaque species. Fungal species isolates from *M. fascicularis* and *M. nemestrina* was generally higher in JR (N=13), compared to CM (N=7). However, Man-Whitney U test revealed no significant differences between number of fungal species and geographic locations (U = 1470, z = -1.262, p = 0.207). Fungal distribution in *M. nemestrina* was equally high in both geographic locations, compared to *M. fascicularis* which was significantly higher in JR compared to CM. *M. nemestrina* on the other hand, shows significantly higher fungal species compared to *M. fascicularis* at CM, but Man-Whitney U test revealed no significant differences between the number of fungal species (U = 1404, z = -0.437, p = 0.662) in both macaques species. Thus, this study reveal no significant differences in number of fungal species on macaques’ species, as well as geographic locations, although JR shows slightly higher fungal distribution compared to CM.

Figure 3(a) Aspergillus clavatus, a) clavate conidial head of *A. clavatus*, b) Septate hyphae and smooth-walled conidia (x100), c) Greyish *A. clavatus* on PDA.

Figure 3(b) Aspergillus niger, a) Globose conidial head of *A. niger*, b) Rough-walled conidia (x100), c) Powdery, black colony of *A. niger* on PDA.
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Figure 3(c) *Aspergillus fumigatus*. a) Conidial head of *A. fumigatus*, b) Smooth-walled conidia (x100), c) Thin, bluish green colony of *A. fumigatus* on PDA.

Figure 3(d) *Aspergillus candidus*. a) Globose conidial head of *A. candidus*, b) Smooth-walled conidia (x100), c) Creamy white colony of *A. candidus* on PDA.

Figure 3(e) *Aspergillus ruber*. a) Conidial head of *A. ruber* (x100), b) Colony of *A. ruber* on PDA.

Figure 3(f) *Aspergillus flavus*. a) Radiating conidial head, b) Hyaline, rough walled conidiophore with conspicuously echinulate conidias (x100), c) Yellow green colonies with dark brown sclerotia.

Figure 3(g) Paecilomyces lilacinus a) P. lilacinus phialides, b) Fusiform conidia (x100), c) Vinaceous colonies on PDA.

Figure 3(h) Paecilomyces variotii a) P. variotii phialides and uneven sizes conidia, b) Chlamydospores of P. variotii (x100), c) Olivaceous colonies on PDA.

Figure 3(i) Curvularia lunata a) Conidiophores and conidia of C. lunata (x100), b) Dark brown, velvety C. lunata colony on PDA.

Figure 3(j) Sporothrix schenckii a) Conidiferous denticles and blastoconidia of S. schenckii (x100), b) White, glabrous and folded S. schenkii colony on PDA.

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Figure 3(k) *Geotrichum candidum*. a) Dichotomous branching hyphae with conidial chains (x100), b) White and butyrous colony on PDA.

Figure 3(l) *Candida parapsilosis*. a) Globose to ovoid yeast cells (x100), b) Cream and lacy colonies on PDA.

Figure 3(m) *Penicillium jensenii*. a) *P. jensenii* phialides and conidia (x100), b) Folded, greyish green colony on PDA.

Figure 3(n) *Penicillium frequentans*. a) Conidiophores of *P. frequentans* (x100), b) Conidia in long compact columns, c) Dark green and velvety colonies on PDA.
Figure 3(o) Penicillium canescen. a) Divergent metulae of P. canescens, b) Ovate, smooth-walled conidia (x100), c) Grey-green and powdery colonies on PDA.

Figure 3(p) Phellinus sp., a) Hyaline, aseptate hyphae of Phellinus sp. (x100), b) Brown, cottony colony with ring structure on PDA.

Figure 3(q) Basidiomycete sp., a) Hyaline, septate hyphae of Basidiomycete sp. (x100), b) White and cottony colony on PDA.

Figure 3(r) Phlebia sp., a) Hyaline, septate hyphae of Phlebia sp. (x100), b) Orange-colored colony on PDA.

Comparisons of fungal species distribution in different anatomical sites

Figure 5(a & b) illustrate distribution of fungal species according to anatomical and geographical locations for each individual macaques' species (N=12). Fungal species isolates from *M. fascicularis* was higher in JR with mean ranges from 2.7 to 3.7 species in all anatomical sites. *M. fascicularis* in CM only recorded 1.3±0.67 fungi species in anus only. Similarly, fungal species isolated from *M. nemestrina* was higher in JR especially in throat with 3.7±0.3, followed by penis/vagina and anus. *M. nemestrina* in CM on the other hand were isolated in anus and penis/vagina only. Similarly Kruskal-Wallis test revealed no significant difference between number of fungal species in different anatomical sites, $\chi^2(2)=1.832$, p = 0.4. However, anus and penis/vagina site revealed higher fungi distribution compared to throat in *M. fascicularis*, whereas in *M. nemestrina* fungal species was higher in throat compared to anus and penis/vagina.
In JR, 4 fungal species were isolated from anus and penis/vagina while 3 species were isolated from throat in *M. fascicularis*, whereby in *M. nemestrina*, 3 species were isolated from anus and penis/vagina while 4 species were isolated from throat (Table 2). Compared to JR, throat in both macaques in CM did not show positive for any fungi species, as can be found in both anus and penis/vagina sites. Most of the isolated fungi were from the anus region of both *M. fascicularis* and *M. nemestrina*. Only *Aspergillus fumigatus* fungi species was shared between both macaques species, in all anatomical sites in JR, but it was absent in samples from CM. However, *Apergillus ruber* and *Aspergillus flavus* were common isolates in both geographic locations.

**Discussion**

The study support our hypothesis as there were higher fungal distribution in macaques residing forest with oil palm plantation, compared to Acacia plantation matrix, for both species. The difference between the numbers of fungal species isolated from the study areas could be explained by the difference in topological cover between the two study areas. Even though both forest matrix were deforested for monoculture plantation, most of the foregone forest in CM was substituted by Acacia plantation. *Acacia mangium*, has been widely used for forest restoration and rehabilitation in Malaysia due to its rapid growth, and high productivity apart from its robust and high adaptability. These trees are believed to serve as additional food sources for many wildlife, as well as providing shelter for them, hence the macaques would not have to encroach into human’s habitat in order to find food. Silva et al. reveal high abundance of phytophagous insect in the plantation, which could attract more wildlife species, including macaques. On the other hand, in JR, macaques are forced to forage in nearby human settlements, where such act of finding food in human settlements will eventually expose them to rubbish and food leftovers, thus increasing the chances of fungus transmission. As opposed to the Acacia trees, oil palm plantation did not serve as an adequate food source for the macaque because it does not provide enough nutrients that the forest usually offers.

In addition to that, with a more disturbed habitat, the macaques have higher susceptibility towards fungal transmission and infections. The behavior of raiding through food trash in human settlement increase their chances of encountering growing pathogenic fungi. According to USDA, many fungi can be found growing in food materials such as poultry and meat including those in genus *Alternaria, Aspergillus, Geotrichum* and many others. These fungi can later infect or transmit to the macaques through inhalation of the spores, or simply by skin contact and the spores can get stuck between their furs. Even though most of the times the fungi will not cause infections in the macaques, however they can act as vectors that can transmit the spores to humans. Being extremely tolerant, the spores can survive up to a very long time being on the macaques’ body before being transmitted to a more suitable medium for their growth. Fungal species were higher in *M. nemestrina* especially in CM. This can be explained by their difference in food choices as previous studies have shown that *M. nemestrina* has been observed to include fungi as part of their dietary intakes.

Overall, *Aspergillus* sp. shows the highest prevalence and being the most ubiquitous among other genus in both macaque species. *Aspergillus* sp. grows very well in decaying organic matter such as food trash as they feed off the carbon. *A. clavatus* was isolated from all three anatomical sites of *M. fascicularis* although the condition offered in each site was different. Rectal region usually have anaerobic condition, in contrast to penis/vagina and throat which have open access to aerobic condition. The pH of each site is also different. Temperature of throat and rectal region may lies in the same range as both are located inside the body cavity (37°C) as opposed to penis/vagina. This is because most of *Aspergillus* sp. such as *A. clavatus* are adaptive and very tolerant, allowing them to grow in different physical environment. *A. fumigatus* was isolated from all three anatomical sites of both *M. fascicularis* and *M. nemestrina*. According to Kwon-Chung and Sugui, there are four features that make *A. fumigatus* an ubiquitous pathogen: 1) ability to survive and grow in a wide range of environmental conditions, 2) effective spore dispersal in air, 3)
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Authors declare that there is no conflict of interest.

**Conflicts of interest**

**References**


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