

Bioaugmentation and biostimulation: a potential strategy for environmental remediation

Abstract

As the world is heading towards rapid urbanization and technological advancements, more undesirable and unwanted activities by human is raising major environmental issues like global warming, imbalance in soil ecological processes leading to lower agricultural yield, drastic climate change, etc. Predominantly, among all, xenobiotic recalcitrant compounds are indiscriminately being disposed in the environment causing significant hazard owing to its high stability and complexity. However, there are several methods for disposing such materials but the most efficient and significant disposal strategy is said to be bioremediation. This biological approach of remediation can be executed through introduction of efficient microbial strains (bioaugmentation) or by addition of rate limiting nutrients to the soil (biostimulation) to enhance the remediation process significantly. Existing literatures provide a broad landscape of the efficiency of some of the microbial strains and a few of the rate limiting nutrients in remediation of the toxic, highly complex and resistant contaminants but lacks information regarding their degree of efficiency and eco-friendliness in comparison with those of other disposal strategies. This review extensively focuses on a comparative discussion on different strategies of bioremediation to remediate the environment from the toxic, hazardous waste with the goal of building a less-toxic, stable and a healthy environment.

Keywords: recalcitrant xenobiotic compound, bioremediation, bioaugmentation, biostimulation, sustainable environmental protection

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Introduction

With the rapid increase in population, urbanisation and industrialisation, currently, the environment is at a stake or on the verge of ecological damage.¹ In other words, the environment is endangered due to human activities that are continuously ruining it.² The ubiquitous contamination and pollution of the nature's elements such as the aquatic and terrestrial ecosystems by the industrial production of chemicals, excessive use of petroleum and its derivatives, polyethylene, pesticides and organic herbicides that are mostly used for protection against weed, insects, and fungal attacks are of serious concern.³⁻⁶ The most prominent sources of these pollutants in the soil and water resources are the oil refineries, gas stations, seeping of water from the herbicide-pesticide treated agricultural lands, petrochemical and pharmaceutical industries. Most of these pollutants are recalcitrant in nature and persist in the environment for long periods of time.⁷ Among all of the above, the most recalcitrant, non-biodegradable, severely toxic and most commonly used polymers are the polyethylene bags. This toxic and headstrong nature of polyethylene is mainly due to three factors - high molecular weight, complex structure and its hydrophobic nature.⁸ Annually 140 million tonnes of synthetic polymers are produced at a growing rate of 12% per annum.^{9,10} Each year, an estimated 500 billion to 1 trillion plastic bags are consumed worldwide.¹¹ The polyethylene bags or any other polyethylene based products are finally dumped into the landfills after their usage leading to severe environmental pollution since they are non-biodegradable under natural environmental conditions.^{12,13}

The plastic wastes generated daily or annually worldwide are one of the serious threats to environmental pollution because of their

semi-permanent stability in the environment. The semi-permanent existence of these plastics in the environment is mainly due to their high molecular weight, cross-linkages, high number of aromatic rings, halogen substitution in their structure¹⁴ and also due to the absence of the functional groups recognizable by the microorganisms^{15,16} etc. Similarly, rubber industries raise severe environmental issues due to different chemical usage at different stages of rubber processing.¹⁷ India is the third largest natural rubber producing country of the world, next to Thailand and Indonesia, producing about 9 percent of the total output.¹⁸ From about 200 hectares in 1902-03, the total area under rubber plantations increased to about 5.9 lakh hectares in 2003-04. Similarly, the production that was 80 tonnes in 1910 increased to about 76000 tonnes in 2003-04. Presently, India's rubber production has reached to 7 lakh tonnes annually.¹⁹ Despite the numerous benefits that are rendered to the World by natural as well as synthetic rubber, the consequence of rubber and its derivatives has raised a serious problem due to its highly polluted effluents.²⁰ The most common environmental issue caused by the rubber industries is the discharge of huge amounts of wastewater containing chemicals, hazardous waste and thermal emission. In addition to this, other environmental issues of rubber processing sector are high concentration of Biological Oxygen Demand (BOD), Chemical Oxygen Demand (COD), Suspended Solids (SS), high concentration of ammonia and nitrogen compounds, high level of sulphate, high level of odour and many more.¹⁷

In case of polychlorinated biphenyls (PCBs), it is accidental spills or improper disposal that leads to their release into the environment.²¹ The maximum PCB production represents a share of about 1.5 percent in world electronic hardware production.²² While talking about the toxic pollutants, the most concerned area can be considered as that

of the toxic and hazardous chemicals released by the industries every year. Each and every second 310 kg of toxic chemicals are released into the air, land and water by the industrial facilities all around the world. This amounts to approximately 10 million tonnes per year.²³ The next most hazardous substance is lubricant oil, a common essentiality that is normally used to run several engines and machines. A subsequent accumulation of these oils from the auto-mechanic workshops may result in serious environmental problems because of its hazardous nature.²⁴ Oil spills, either it could be crude oil or refined oil, is also significantly intimidating the health of the environment and living creatures including humans.

Oil spills are mainly due to the release of crude oil from tankers, offshore platforms, drilling rigs and wells, as well as spills of refined petroleum products and their by-products, heavier fuels used by large ships such as bunker fuel etc.²⁵ Spilled oil can also contaminate drinking water supplies. For example, in 2013 two different oil spills contaminated water supplies for three hundred thousand people in Miri, Malaysia, eighty thousand people in Coca, Ecuador.²⁶ In 2000, springs were contaminated by an oil spill in Clark County, Kentucky.²⁶ Though the oil spill incidents are relatively showing a downward trend from 28.1 numbers of spills in 1990 to 5.2 number of spills in 2010, but it has not been completely stopped. In reference to this, the two large recorded oil spills that occurred in the year 2015 in Singapore and Turkey can be accounted. Removal of these compounds from the environment always remains a challenging task for the people globally. In this context, there is a significant need for remediation measures to protect the environment as well as to secure the drinking water resources.²⁷ The presence of these substances in the environment not only pose health hazards to the people globally but also raises a threat to the environmental food chain due to their toxicity, mutagenic and carcinogenic properties.⁷

Limitations of present disposal techniques

There are several methods that have been designed for the removal or elimination of these hazardous toxic pollutants from the soils. Some of the methods used for the disposal of the hazardous pollutants from the environment include incineration, recycling and landfills.²⁸ Disposal of the toxic waste by the process of incineration is not a good approach as incineration of the waste materials results in a large amount of toxic emissions such as carbon monoxide and methane. The release of these gases to the atmosphere ultimately brings severe changes in climate.²⁹ At the same time, disposal of the used materials like polyethylene bags, rubber tyres, etc into the landfills will pose serious threats not only to the soil microflora but also proves hazardous to the mankind. Existing literature tried focussing on the biodegradation of polythene because of the demerits of the existing disposal strategies for Polythenes like incineration, dumping into landfills, recycling that is not only a hazard towards environmental health but also highly expensive, time-consuming and labour intensive.³⁰

Some other techniques that are under development includes extraction of pollutants with organic solvents, oxidation of organic pollutants under subcritical or supercritical conditions, vitrification, electro-reclamation, dehalogenation of chlorinated organic compounds, chemical reduction or oxidation of contaminants, steam stripping, plasma torch techniques etc.^{31,32} Out of these techniques mentioned above, the biological method for the removal of soil contaminants was considered as the most potent and environmentally friendly approach.³³ This approach is covered under an umbrella term called bioremediation. By definition, bioremediation is described as

the use of microorganisms to destroy or detoxify waste materials.^{34,35} Biological approach for elimination of soil contaminants involves a wide array of microorganisms. The reason behind this is their biodiversity and vast catabolic potential.^{36,37} For effective removal of the recalcitrant compounds from the soil, the best biological approach is considered to be the “*in-situ* bioremediation”. This process of detoxification targets the harmful chemicals by mineralisation, transformation, or alteration.^{38,39} Bioremediation at its natural pace is incapable of eliminating the toxic and hazardous waste from the environment considerably. In order to enhance the degradation rate of the waste through bioremediation, bioremediation is been coupled with two other approaches named as bioaugmentation and biostimulation.

Bioaugmentation

Bioaugmentation is the method of application of autochthonous or allochthonous wild type or genetically modified microorganisms to polluted hazardous waste sites in order to accelerate the removal of undesired compounds.³ Figure 1 outlines the process of bioaugmentation. Bioaugmentation is mainly undertaken in oil contaminated environments as an alternate strategy for bioremediation.

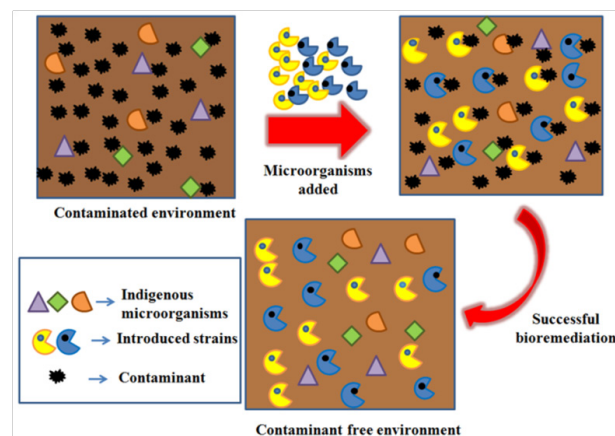


Figure 1 The pictorial diagram of bioaugmentation.

Principle of bioaugmentation

The rationale of this approach is to enhance the degree or rate of degradation of the complex pollutants by the addition of pollutant-degrading microorganisms.^{40,41} Enhancing the microbiota of a contaminated site will not only enhance the elimination of the pollutants from the particular site but also at the same time increases the genetic capacity of the desired site. Therefore, bioaugmentation corresponds to an increase in the gene pool and, thus, the genetic diversity of the site. In principle, this genetic diversity could be increased by augmenting the microbial diversity.^{42,43}

Factors influencing bioaugmentation

The success of bioaugmentation process depends mainly on the adaptation of the microbial consortia to the site that needs to be decontaminated. The success of the process also relies on the ability of the newly introduced microbial consortia to compete with the indigenous microorganisms, predators and various abiotic factors.²⁵ Previous studies have shown that it improves the biodegradation capabilities of the contaminated soil by improving the efficiency of the remediation process.^{3,44} Bioaugmentation has been mostly undertaken in soils with a lower number of pollutant-degrading microorganisms

and with compounds that require multi-process remediation.^{3,45} Apart from these, there are several other parameters that control the rate of bioaugmentation occurring in soil. Some of the noted parameters or factors that influence the process of bioaugmentation are pH, temperature, moisture, organic matter content, aeration, nutrient content and soil type.^{3,46} Lack of any of these soil parameters under natural condition makes the remediation process inefficient.⁴⁶⁻⁴⁸ Existing literature stated that *Burkholderia* sp. FDS-1 can optimally degrade nitrophenolic pesticides at a temperature of about 30°C and at a slightly alkaline pH. As mentioned, the diverse catabolic activities of the microbial cells are mostly due to the presence of catabolic genes and enzymes.⁴⁹⁻⁵²

Moreover, the existence of a variety of adaptation strategies in microorganisms such as the ability to modify the cellular membrane to maintain the necessary biological functions,^{53,54} the production of surface-active compounds such as biosurfactants⁵⁵ and of the use of efflux pumps to decrease the concentration of toxic compounds inside the cells^{53,56} makes them a significant weapon for the remediation of contaminated soils. One of the most important soil parameters influencing the effectiveness of bioaugmentation is organic matter content. It plays a crucial role in bioavailability of pollutants and impairs the survival of inoculated strains and their ability to degrade contaminants. In this context, the studies carried out by Mrozik and Piotrowska-Seget 2010 and Greer and Shelton 1992 was quite convincing.^{3,57} They reported that degradation of 2, 4-dichlorophenoxyacetic acid was found to be optimum at high organic matter soil in comparison with soil containing lower organic matter. In addition to this, the moisture availability in the soil also influences the rate of bioaugmentation. In this context, the studies carried out by Ronen et al.,⁵⁸ can be taken into account.

The article studied the effect of water content on the survival of *Achromobacter piechaundii* TBPZ and the degradation rate for tribromophenol. They observed that in the presence of 25% and 50% water content in the soil, the degradation rate was found to be rapid whereas with the decrease in the water content the degradation rate decreases. Existing literature revealed that low soil water content decreased the activity of bacteria mainly due to diffusion limitation of substrate supply and adverse physiological effects associated with cell dehydration.^{3,59} Similarly, the studies carried out by Kim et al.,⁶⁰ revealed that *Pseudomonas spadix* BD-a59 grow very fast in slurry systems amended with sterile soil in comparison with that of the soil that was combusted earlier for the elimination of organic matter. This particular study made by Kim et al.,⁶⁰ gave an idea of the essentiality of the insoluble organic compounds for the BTEX degrading microorganisms.

Table 1 Microorganisms used in bioaugmentation studies

Microorganism used	Pollutants degraded
<i>Pseudomonas putida</i> PaV340/pDH5	4 chlorobenzoic acid ⁹³ Chlorobenzoates
<i>Cupriavidus necator</i> RW112	Arochlor 1221 and 1232 ⁹⁴ Arochlor 1242 ⁹⁵
<i>Burkholderia xenovorans</i> LB400 (ohb)	2,4-dinitrotoluene ⁹⁶
<i>Pseudomonas fluorescens</i> RE	2,4-dinitrotoluene ⁹⁶
<i>Pseudomonas fluorescens</i> MP	2,4-dinitrotoluene ⁹⁶
<i>Pseudomonas putida</i> KT2442	Naphthalene ⁸⁷

Spectrum of microbes used in bioaugmentation

Existing pieces of the literature showed a varied role of bioaugmenting microorganisms in the augmentation of the contaminated sites. The studies carried out by Wang et al.,⁶¹ and Xu et al.,⁶² reported the biodegradation efficiency of *Burkholderia pickettii* against Quinoline. It was observed that though the indigenous organisms of the contaminated site were incapable of degrading Quinoline but these organisms in cooperation with the newly introduced Quinoline degrader *Burkholderia pickettii*, can degrade Quinoline (concentration of 1mg/g of soil), within 6 and 8 hours. The experiments carried out by Tchelet et al.,⁶³ showed the efficiency and capability of β -Proteobacterium *Pseudomonas* sp. strain P51 to degrade chlorinated benzenes. Their results demonstrated the possibility of successful bioaugmentation by applying preselected strains to degrade poorly degradable substance like TCB in the contaminated site. A wide variety of microbial strains or microbial consortia can be employed for the process of soil augmentation. Contaminants in the soil not only affect the soil quality but also the soil microbial communities are known for rendering several important functions in the soil.

Selection of appropriate microorganism: Choosing the best tool

Successful soil augmentation requires appropriate selection of microbial strains or microbial consortia. The selection of a suitable strain or a suitable microbial consortium requires the consideration of few of the features of microorganisms like fast growth, easily culturable, ability to withstand high concentrations of contaminants and to survive in a wide range of environmental conditions. There are several approaches that can be considered for selection of a suitable bacterial strain. One of the approaches involves isolation of bacterial strain from a contaminated soil followed by its culturing under laboratory conditions for its pre adaptation and finally augmented back into the same contaminated soil. This approach is called as re inoculation of soil with indigenous microorganisms. Table 1 shows the success of bioaugmentation in polluted environments. Tribedi et al.,⁶⁴ had isolated an organism *Pseudomonas* sp. AKS2 from landfill soil and characterised the polyethylene succinate (PES) degradation activity in laboratory condition. They also showed that the isolated organism AKS2 exhibited PES degradation in soil contaminated with PES without damaging soil ecological balance, proving an efficient bioaugmentation tool against PES remediation.⁴⁴ Similarly, another approach for augmentation of contaminated soil can be through selection of appropriate microorganisms from different polluted sites with similar contaminants and finally augmenting them into the desired contaminated site.^{41,65,66}

Table continued...

Microorganism used	Pollutants degraded
<i>Pseudomonas fluorescens</i> F113rifpcbrmBP1::gfpmut3	Biphenyl, polychlorinated biphenyl ⁹⁷
<i>Rhodococcus</i> sp. StrainRHA1	4-chlorobenzoate ⁹⁸
<i>Escherichia coli</i> AtzA	Atrazine ⁹⁹
<i>Pseudomonas</i> sp. <i>Pseudomonasputida</i> B13ST1 (pPOB)	3 phenoxybenzoic acid ¹⁰⁰
<i>Pseudomonas fluorescens</i> F113rifPCB	Biphenyl, polychlorinated ¹⁰¹
<i>Pseudomonas fluorescens</i> CS2	Biphenyl Ethylbenzene ¹⁰²
<i>Pseudomonas putida</i> BCRc14349	Phenol, trichloroethane ¹⁰³
<i>Rhodococcus</i> sp.F92	Various petroleum products ¹⁰⁴
<i>Arthrobacter</i> , <i>Burkholderia</i> , <i>Pseudomonas</i> , <i>Rhodococcus</i> etc.	Petroleum hydrocarbons ¹⁰⁵

Biostimulation: an efficient strategy of bioremediation

Biostimulation is a remediation technique that is highly efficient, cost effective and eco-friendly in nature. Biostimulation refers to the addition of rate limiting nutrients like phosphorus, nitrogen, oxygen, electron donors to severely polluted sites to stimulate the existing bacteria to degrade the hazardous and toxic contaminants.^{4,67-69} Figure 2 outlines the process of biostimulation.

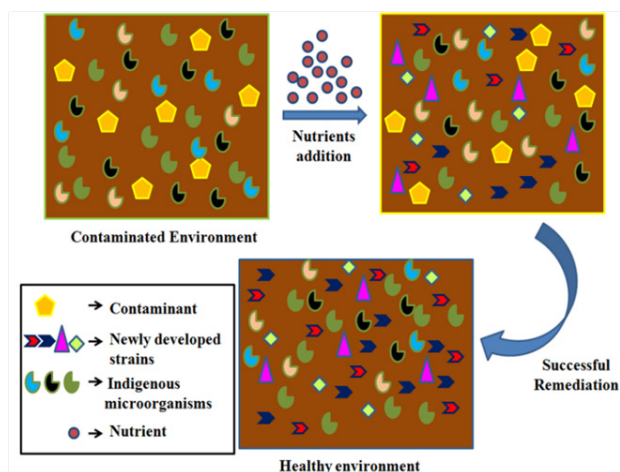


Figure 2 The pictorial diagram of biostimulation.

The mechanism of stimulation: Role of rate limiting nutrients

The addition of rate limiting nutrients improves the degradation potential of the inhabitant microorganisms efficiently as it significantly accelerates the decontamination rate.^{41,70} Among all of the bioremediation techniques, biostimulation is considered to be the most efficient method for remediation of hydrocarbons.⁴¹ Existing works of literature have established biostimulation as an important remediation tool for the degradation of hydrocarbons especially petroleum products and its derivatives.⁷¹ To be specific, petroleum contaminated sites having less efficient and metabolically poor microbial population can be remediated significantly by the addition of some of the rate limiting nutrients or through the process of biostimulation.^{4,72} This is mostly due to the easy availability of carbon

(C) source which is one of the rate-limiting nutrients required by the indigenous microorganisms for their metabolic activities from the petroleum contaminants. So it can be said that just meagre addition of few of the other rate limiting nutrients except carbon onto the soil boosts up the petroleum degradation rate significantly. In addition to the aforementioned rate limiting nutrients, implementation of certain other nutrient rich organic matter can also trigger the remediation process extensively. In this context, the study carried out by Sarkar et al.,⁷³ is worth mentioning. They have observed that addition of bio solids (nutrient rich organic matter) obtained from the treatment of domestic sewage and inorganic fertilisers, rich in nitrogen and phosphorus gears up the degradation rate of petroleum hydrocarbons up to 96%.

Factors influencing the biostimulation

Bioremediation of a polluted site through biostimulation is influenced by few of the environmental parameters like pH, moisture content, temperature, etc.²⁴ In addition to these factors, the prevailing environmental physiology also influences biostimulation rate. In this context, the example of bioremediation process in the marine environment can be taken into account. In the marine ecosystem, the bioremediation rate is significantly low as it becomes fairly difficult for the microorganisms to target the polymer for degradation as there are high chances that the polymer will get diluted or washed out by the wave action. It is not always necessary that addition of nutritious matter onto the soil will enhance remediation process but there are examples that reflect the adverse effects of excessive addition of nutrients onto the soil. The experimental work carried out by Nikolopoulou & Kalogerakis⁷⁰ showed that higher concentrations of N and P sources can cause eutrophication, which enhance the algal growth and ultimately reduce the dissolved oxygen concentration in the water resulting in the death of several aquatic lives.⁷⁰ Thus, we can say that dependency of biostimulation on environmental factors can limit the progress or efficiency of the method being adopted.⁷⁴ To be specific, biostimulation works efficiently towards the elimination of complex contaminants from the ecosystem by maintaining a proper balance between the desirable and undesirable addition of rate limiting nutrients onto the soil.

Application of biostimulation

The major contaminants that can be successfully remediated through biostimulation are petroleum hydrocarbons, sulphate and

polyester polyurethanes. Sulphate contamination of groundwater is a threat not only to the environmental ecology but also towards human health. This can be remediated biologically through the process of biostimulation. It requires amendment of electron-donor that will enhance sulphate reduction and thus remediate the same.⁷⁵ In the case of polyester polyurethanes, biostimulation plays a significant role in enhancing its degradation rate in soil. The polyester polyurethanes (PU) are a diverse group of synthetic polymers with many industrial and commercial applications, including insulating and packaging foams, fibers, fabrics, and synthetic leather goods.^{76,77} These polymers contain intra-molecular bonds analogous to those found in biological macromolecules (such as ester and urethane linkages). It is due to

the presence of these intramolecular bonds that enhances microbial degradation as these bonds act as a site for microbial attack.⁷⁷⁻⁷⁹ Pre-existing literature regarding petroleum contaminated sites indicated that biostimulation stands out to be the best approach in those cases where microbial population gets acclimatised due to exposure to hydrocarbons at contaminated sites.^{40,71,80} Thus, in the due course of remediation process, the adapted populations showed higher remediation rates in comparison to those with no contamination exposure history. This is an indicative study reflecting the efficiency of biostimulation of polluted sites. Table 2 has been introduced in the manuscript to show the success of biostimulation in polluted environments.

Table 2 Reports on biostimulation based bioremediation

Nutrients used	Target pollutants
Animal manure and sewage sludge	Atrazine and alachlor ¹⁰⁶
Activated sludge	Atrazine and simazine ¹⁰⁷
Plant residues, ground seed, or commercial meal	Alchlor, metolachlor, atrazine and trifluralin ¹⁰⁸
Cellulose, straw and compost	Atrazine ¹⁰⁹
Cornmeal, ryegrass and poultry litter	Cyanazine and fluometuron ¹¹⁰
Dairy manure	Atrazine ¹¹¹
Maize straw	Methabenzthiazuron ¹¹²
Ammonium nitrate, potassium nitrate and ammonium phosphate	Atrazine ¹¹³
Phosphorus	2,6 Di chloro benzonitrile and atrazine ¹¹⁴
Nitrate	(R)-mecoprop ¹¹⁵
Nitrate and phosphorus	Isoproturon ¹¹⁶
Tryptic soy broth	Dichlofop ¹¹⁷
Mannitol	Atrazine ¹¹⁸
Minimal nutrient medium, casamino acid, glucose and phosphate	2,4-Di chloro phenoxy acetic acid, methylchlorophenoxypropionic acid ¹¹⁵

Advantages and pitfall of biostimulation

The primary advantage of biostimulation is that it is done by native microorganisms that are well suited to the environment and are already well distributed spatially.⁴¹ Secondly, biostimulation is an eco-friendly and cost effective technique which can be performed anywhere⁸¹ and lastly, biostimulation helps in the degradation of contaminants internally which prevents any kind of disturbances to the environment. In spite of its high efficiency in the remediation process, biostimulation has certain well defined disadvantages. The major disadvantage of biostimulation is its dependency on environmental factors that controls its potentiality. Secondly, when the contaminants are firmly engrossed to the soil particles or the contaminant is non biodegradable, then biostimulation cannot be executed. Thirdly, biostimulation is extremely site specific and requires immense scientific observation.⁸¹

Roles of environmental genomics on bioremediation

Environmental functional genomics is highly significant for understanding the gene arrangement and metabolic properties of microorganisms residing in a particular environment. For an in-depth study of few of the non-cultured, significantly potential microorganisms playing pivotal role in ecological balance, environmental genomics is of great importance.⁸² Environmental genomics is essential for those habitats that harbours a wide array of microorganisms involved in the transformation of organic nitrogen, carbon and phosphorus.⁸³⁻⁸⁵ This study enables in understanding and revealing the gene pool of the microbiota associated with the particular habitat.^{86,87} In the present times, people have adopted bioremediation and biostimulation as efficient disposal and remediation strategies without having the knowledge about the degradation pathways going on in the process of remediating and eliminating polymers from the environment. This is mostly due to our inability in isolating majority of earth's

microorganisms and cultivating them in appropriate media though there is huge advancement in the microbial techniques.⁸⁸ In spite of the molecular techniques there is very less known about the protein coding genes and thereby the microbial diversity existing in the environment.⁸⁹ Metagenomics or environmental genomics takes up a significant place in analysing the existing microbial communities.⁹⁰ Few of the advantages of environmental genomics or Metagenomics are-helps in the search for new catabolic genes for the degradation of a wide range of xenobiotic, aromatic compounds, etc. It also enables in screening clones capable of expressing certain desired traits on certain appropriate media.^{91,92}

Conclusion

The earth is at a severe risk due to rapid industrialisation and urbanisation resulting in serious environmental issues that include global warming, acid rain, eutrophication, loss of microbial functional diversity of soil, depletion in soil quality and much more. Moreover, dumping of toxic waste into the soil like rubber, polyethylene, PCB, etc raised a severe threat against the soil microbial community responsible for several important ecological processes. The most potential disposal strategy among all was believed to be bioremediation. This is because of its eco-friendly and cost effective nature. The current study extensively focuses on the benefits of applying bioremediation using bioaugmentation and biostimulation as a proposed disposal strategy on the environment to compensate the loss incurred as a consequence of the implementation of other disposal strategies.

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None.

Conflicts of interest

The authors declare that there is no conflicts of interest.

References

- Cherniwchan J. Economic growth, industrialization and the environment. *Resource and Energy economics*. 2012;34(4):442–467.
- Goudie AS. *The human impact on the natural environment: past, present and future*. John Wiley and Sons; 2013. 424 p.
- Mrozik A, Piotrowska-Seget Z. Bioaugmentation as a strategy for cleaning up of soils contaminated with aromatic compounds. *Microbiol Res*. 2010;165(5):363–375.
- Tyagi M, da Fonseca MM, de Carvalho CC. Bioaugmentation and biostimulation strategies to improve the effectiveness of bioremediation processes. *Biodegradation*. 2010;22(2):231–241.
- Federici E, Giubilei M, Santi G, et al. Bioaugmentation of a historically contaminated soil by polychlorinated biphenyls with *Lentinus tigrinus*. *Microb Cell Fact*. 2012;11:35.
- Schultz-Jansen N, Aamand J, Sorenson SR. Bioaugmentation potential of free and formulated 2,6-dichlorobenzamide (BAM) degrading *Aminobacter* sp. MSH1 in soil, sand and water. *AMB Express*. 2016;6(1):33.
- Srirangan K, Akawi L, Moo-Young M, et al. Towards sustainable production of clean energy carriers from biomass resources. *Applied Energy*. 2012;100:172–186.
- Harshavardhan K, Jha B. Biodegradation of low-density polyethylene by marine bacteria from pelagic waters, Arabian Sea, India. *Mar Pollut Bull*. 2013;77(1-2):100–106.
- Shimao M. Biodegradation of plastics. *Current Opinion in Biotechnology*. 2001;12(3):242–247.
- Leja K, Lewandowicz G. Polymer biodegradation and biodegradable polymers—a review. *Polish J Environ Stud*. 2010;19(2):255–266.
- Roy PK, Surekha P, Tulsi E, et al. Degradation of abiotically aged LDPE films containing pro-oxidant by bacterial consortium. *Polym Degrad Stab*. 2008;93(10):1917–1922.
- Burd D. Plastic not fantastic. *Reports of the Canada Wide Science Fair*. 2008;1:1–5.
- Jalil MA, Mian MN, Rahman MK. Using plastic bags and its damaging impact on environment and agriculture: An alternative proposal. *International Journal of Learning and Development*. 2013;3(4):1–4.
- Botre S, Jadhav P, Saraf L, et al. Screening and Isolation of Polyethylene degrading Bacteria from various sources. *Int Res J Environmental Sci*. 2015;4(11):58–61.
- Soni R, Kapri A, Zaidi MG, et al. Comparative biodegradation studies of non-poritized and poritized LDPE using indigenous microbial consortium. *Journal of Polymers and the Environment*. 2009;17(4):233–239.
- Zahra S, Abbas SS, Mahsa MT, et al. Biodegradation of low-density polyethylene (LDPE) by isolated fungi in solid waste medium. *Waste Manag*. 2010;30(3):396–401.
- Jagadale SC, Rajkumar K, Chavan RP, et al. Environmental concern of pollution in rubber industry. *International Journal of Research in Engineering and Technology*. 2015;4(11):187–191.
- Chennakrishnan P. Natural rubber production in India. *International Journal of research in commerce, IT and management*. 2012;2(11).
- Joseph KJ. Exploring exclusion in innovation systems: case of plantation agriculture in India. *Innovation and Development*. 2014;4(1):73–90.
- Jamatia A, Chakraborty S, Das D, et al. Evaluation of Physicochemical characteristics of disposed rubber industry effluent: A case study of Bodhjungnagar industrial growth centre. *IOSR Journal of Engineering*. 2014;4(7):44–50.
- Beyer A, Biziuk M. Environmental fate and global distribution of polychlorinated biphenyls. *Rev Environ Contam Toxicol*. 2009;201:137–158.
- Breivik K, Sweetman A, Pacyna J, et al. Towards a global historical emission inventory for selected PCB congeners—a mass balance approach. Global production and consumption. *Science of the Total Environment*. 2012;290(1-3):181–198.
- Tiefenbacher JP, Hagelman III RR. Environmental equity in Urban Texas: race, income and patterns of acute and chronic toxic air releases in metropolitan countries. *Urban Geography*. 2013;20(6):516–533.
- Abdulsalam S, Omale AB. Comparison of Biostimulation and Bioaugmentation techniques for the Remediation of Used Motor Oil Contaminated Soil. *Brazilian Archives of Biology and Technology*. 2009;52(3):747–754.
- Godleads OA, Prekeyi TF, Samson EO, et al. Bioremediation, Biostimulation and Bioaugmentation: A Review. *International Journal of Environmental Bioremediation & Biodegradation*. 2015;3(1):28–39.
- Campbell R, Clifford K. *Gulf Spill Is the Largest of Its Kind, Scientists Say*. The New York Times; 2010.
- Atlas R, Bragg J. Bioremediation of marine oil spills: when and when not—the Exxon Valdez experience. *Microb Biotechnol*. 2009;2(2):213–221.
- Pramila S, Fulekar MH, Pathak B. E-Waste—A challenge for tomorrow. *Research Journal of Recent Sciences*. 2012;1(3):86–93.
- Einas IAM, Hago EFH. Effect of the low density polyethylene bags waste on the asphalt mixture. *International Journal of Engineering Research and Science and Technology*. 2014;3(2):1–10.

30. Sen SK, Raut S. Microbial degradation of low density polyethylene (LDPE): a review. *Journal of Environmental Chemical Engineering*. 2015;3(1):462–473.
31. Rulkens WH, Tichy R, Grotenhuis JTC. Remediation of polluted soil and sediment: perspectives and failures. *Water Sci Tech*. 1998;37(8):27–35.
32. Garbisu C, Alkorta I. Basic concepts on heavy metal soil bioremediation. *European Journal of Mineral Processing and Environmental Protection*. 2003;3(1):58–66.
33. Hamdi H, Benzarti S, Manusadzianas L, et al. Bioaugmentation and biostimulation effects on PAH dissipation and soil ecotoxicity under controlled conditions. *Soil Biol Biochem*. 2007;39(8):1926–1935.
34. Shanahan P. *Waste Containment and Remediation Technology*. USA: Massachusetts Institute of Technology; 2004.
35. Sinha RK, Valani D, Sinha S, et al. *Bioremediation of contaminated sites: a low-cost nature's biotechnology for environmental clean up by versatile microbes, plants & earthworms*. Nova Publishers; 2010. 73 p.
36. Dua M, Singh A, Sethunathan N, et al. Biotechnology and bioremediation: successes and limitations. *Appl Microbiol Biotechnol*. 2002;59(2-3):143–152.
37. Kumar BL, Gopal DS. Effective role of indigenous microorganisms for sustainable environment. *Biotech*. 2015;5(6):867–876.
38. Shannon MJ, Unterman R. Evaluating bioremediation: distinguishing fact from fiction. *Annu Rev Microbiol*. 1993;47:715–738.
39. Marmulla R, Harder J. Microbial monoterpene transformations-A review. *Front Microbiol*. 2014;5:346.
40. Leahy JG, Colwell RR. Microbial Degradation of hydrocarbons in the environment. *Microbiol Rev*. 1990;54(3):305–315.
41. Adams GO, Fufeyin PT, Okoro SE, et al. Bioremediation, biostimulation and bioaugmentation: a review. *International Journal of Environmental Bioremediation & Biodegradation*. 2015;3(1):28–39.
42. Dejonghe W, Boon N, Seghers D, et al. Bioaugmentation of soils by increasing microbial richness: missing links. *Environ Microbiol*. 2001;3(10):649–657.
43. Shukla KP, Singh NK, Sharma S. Bioremediation: developments, current practices and perspectives. *Genetic Engineering and Biotechnology Journal*. 2010;3:1–20.
44. Tribedi P, Sil AK. Bioaugmentation of polyethylene succinate-contaminated soil with *Pseudomonas* sp. AKS2 results in increased microbial activity and better polymer degradation. *Environ Sci Pollut Res*. 2013;20(3):1318–1326.
45. Forsyth JV, Tsao YM, Bleam RD. Bioremediation: when is augmentation needed? In: Hinchee RE, Fredrickso J, Alleman BC, editors. *Bioaugmentation for site remediation*. Columbus, OH: Battelle Press; 1995. 14 p.
46. Hong Q, Zhang Z, Hong Y, et al. A microcosm study on bioremediation of fenitrothion-contaminated soil using *Burkholderia* sp. FDS-1. *Int Biodeterior Biodegrad*. 2007;59(1):55–61.
47. Atlas RM. Bioremediation of petroleum pollutants. *Int Biodeterior Biodegrad*. 1995;35(1-3):317–327.
48. Jimenez N, Vinas M, Sabate J, et al. The Prestige oil spill. 2. Enhanced biodegradation of a heavy fuel oil under field conditions by the use of an oleophilic fertilizer. *Environ Sci Technol*. 2006;40(8):2578–2585.
49. der Geize R, Dijkhuizen L. Harnessing the catabolic diversity of *Rhodococci* for environmental and biotechnological applications. *Curr Opin Microbiol*. 2004;7(3):255–261.
50. de Carvalho CCCR, da Fonseca MMR. The remarkable *Rhodococcus erythropolis*. *Appl Microbiol Biotechnol*. 2005;67(6):715–726.
51. Khomenkov VG, Shevelev AB, Zhukov VG, et al. Organization of metabolic pathways and molecular-genetic mechanisms of xenobiotic degradation in microorganisms: a review. *Appl Biochem Microbiol*. 2008;44(2):117–135.
52. Rivelli V, Franzetti A, Gandolfi I, et al. Persistence and degrading activity of free and immobilised allochthonous bacteria during bioremediation of hydrocarbon-contaminated soils. *Biodegradation*. 2013;24(1):1–11.
53. Isken S, de Bont JA. Bacteria tolerant to organic solvents. *Extremophiles*. 1998;2(3):229–238.
54. de Carvalho CCCR, Wick LY, Heipieper HJ. Cell wall adaptations of planktonic and biofilm *Rhodococcus erythropolis* cells to growth on C5 to C16 n-alkane hydrocarbons. *Appl Microbiol Biotechnol*. 2009;82(2):311–320.
55. Ron EZ, Rosenberg E. Biosurfactants and oil bioremediation. *Curr Opin Biotechnol*. 2002;13(3):249–252.
56. Van Hamme JD, Singh A, Ward OP. Recent advances in petroleum microbiology. *Microbiol Mol Biol Rev*. 2003;67(4):503–549.
57. Greer LE, Shelton DR. Effect of inoculants strain and organic matter content on kinetics of 2,4-dichloro-phenoxyacetic acid degradation in soil. *Appl Environ Microbiol*. 1992;58(5):1459–1465.
58. Ronen Z, Vasiluk L, Abeliovich A, et al. Activity and survival of tribromophenol-degrading bacteria in a contaminated desert soil. *Soil Biol Biochem*. 2000;32(11):1643–1650.
59. Mashreghi M, Prosser JL. Survival and activity of lux-marked phenanthrene-degrading *Pseudomonas stutzeri* P16 under different conditions. *Iran J Sci Technol*. 2006;30(1):71–80.
60. Kim JM, Le NT, Chung BS, et al. Influence of soil components on the biodegradation of benzene, toluene, ethylbenzene and o-, m-, and p-xylenes by the newly isolated bacterium *Pseudoxanthomonas spadix* BD-a59. *Appl Environ Microbiol*. 2008;74(23):7313–7320.
61. Wang JL, Mao ZY, Han LP, et al. Bioremediation of quinoline-contaminated soil using bioaugmentation in slurry-phase reactor. *Biomed Environ Sci*. 2004;17(2):187–195.
62. Xu P, Ma W, Han H, et al. Biodegradation and interaction of quinoline and glucose in dual substrates system. *Bull Environ Contam Toxicol*. 2015;94(3):365–369.
63. Tchelet R, Meckenstock R, Steinle P, et al. Population dynamics of an introduced bacterium degrading chlorinated benzenes in a soil column and in sewage sludge. *Biodegradation*. 1999;10(2):113–125.
64. Tribedi P, Sarkar S, Mukherjee K, et al. Isolation of a novel *Pseudomonas* sp from soil that can efficiently degrade polyethylene succinate. *Environ Sci Pollut Res*. 2012;19(6):2115–2124.
65. Goux S, Shapir N, El Fantroussi S, et al. Long term maintenance of rapid atrazine degradation in soils inoculated with atrazine degraders. *Water Air Soil Pollut Focus*. 2003;3(3):131–142.
66. Ghazali FM, Rahman RNZA, Salleh AB, et al. Biodegradation of hydrocarbons in soil by microbial consortium. *Int Biodeterior Biodegrad*. 2004;54(1):61–67.
67. Elektorowicz M. Bioremediation of petroleum-contaminated clayey soil with pretreatment. *Environ Technol*. 1994;15(4):373–380.
68. Piehler MF, Swistak JG, Pinckney JL, et al. Stimulation of Diesel Fuel Biodegradation by Indigenous Nitrogen Fixing Bacterial Consortia. *Microb Ecol*. 1999;38(1):69–78.
69. Rhykerd RL, Crews B, McInnes KJ, et al. Impact of bulking agents, forced aeration and tillage on remediation of oil-contaminated soil. *Bioresource Technol*. 1999;67(3):279–285.
70. Nikolopoulou M, Kalogerakis N. Biostimulation strategies for fresh and chronically polluted marine environments with petroleum hydrocarbons. *J Chem Technol Biotechnol*. 2009;84(6):802–807.

71. Abid A, Zaafour K, Aydi A, et al. Feasibility of a bioremediation process using biostimulation with inorganic nutrient NPK for hydrocarbon contaminated soil in Tunisia. *Journal of Bioremediation and Biodegradation*. 2014;5(4):224.
72. Delille D, Coulon F, Pelletier E. Effects of temperature warming during a bioremediation study of natural and nutrient-amended hydrocarbon-contaminated sub-Antarctic soils. *Cold Reg Sci Technol*. 2004;40(1-2):61–70.
73. Sarkar D, Ferguson M, Datta R, et al. Bioremediation of petroleum hydrocarbons in contaminated soils: comparison of biosolids addition, carbon supplementation, and monitored natural attenuation. *Environ Pollut*. 2005;136(1):187–195.
74. Zawierucha I, Malina G. *Bioremediation of contaminated soils: Effects of bioaugmentation and biostimulation on enhancing biodegradation of oil hydrocarbons*. Bioaugmentation, Biostimulation and Biocontrol; 2011. 187–201 p.
75. Miao Z, Brusseau ML, Johnson B, et al. Sulphate reduction in Groundwater: Characterization and applications for remediation. *Environ Geochem Health*. 2012;34(4):539–550.
76. Matsumura S, Soeda Y, Toshima K. Perspectives for synthesis and production of polyurethanes and related polymers by enzymes directed toward green and sustainable chemistry. *Appl Microbiol Biotechnol*. 2006;70(1):12–20.
77. Cosgrove L, McGeechan PL, Handley PS, et al. Effect of biostimulation and bioaugmentation on degradation of polyurethane buried in soil. *Applied and Environmental Biology*. 2010;76(3):810–819.
78. Kawai F. Breakdown of plastics and polymers by microorganisms. *Adv Biochem Eng Biotechnol*. 1995;52:151–194.
79. Zheng Y, Yanful EK, Bassi AS. A review of plastic waste biodegradation. *Crit Rev Biotechnol*. 2005;25(4):243–250.
80. Simon MA, Bonner JS, Mc Donald TJ, et al. Bioaugmentation for the enhanced bioremediation of petroleum in a wetland. *Polycycl Aromat Compd*. 1999;14(1-4):231–239.
81. Mohee R, Mudhoo A. *Bioremediation and sustainability: Research and Applications*. John Wiley & Sons; 2012.
82. Quaiser A, Ochsenreiter T, Lanz C, et al. Acidobacteria form a coherent but highly diverse group within the bacterial domain: evidence from environmental genomics. *Mol Microbiol*. 2003;50(2):563–575.
83. Whitman WB, Coleman DC, Wiebe, WJ. Prokaryotes: the unseen majority. *Proc Natl Acad Sci USA*. 1998;95(12):6578–6583.
84. Dunbar J, Takala S, Barns SM, et al. Levels of bacterial community diversity in four arid soils compared by cultivation and 16S rRNA gene cloning. *Appl Environ Microbiol*. 1999;65(4):1662–1669.
85. Torsvik V, Ovreas L, Thingstad TF. Prokaryotic diversity–magnitude, dynamics, and controlling factors. *Science*. 2002;296(5570):1064–1066.
86. Nesbo CL, Boucher Y, Doolittle WF. Defining the core of non transferable prokaryotic genes: the euryarchaeal core. *J Mol Evol*. 2001;53(4-5):340–350.
87. Filonov AE, Akhmetov LI, Puntus IF, et al. The construction and monitoring of genetically tagged, plasmid-containing, naphthalene-degrading strains in soil. *Microbiology*. 2005;74(4):453–458.
88. Leadbetter JR. Cultivation of recalcitrant microbes: cells are alive, well and revealing their secrets in the 21st century laboratory. *Curr Opin Microbiol*. 2003;6(3):274–281.
89. Amann RI, Ludwig W, Schleifer KH. Phylogenetic identification and in situ detection of individual cells without cultivation. *Microbiol Rev*. 1995;59(1):143–169.
90. Schloss PD, Handelsman J. Biotechnological prospects from metagenomics. *Curr Opin Biotechnol*. 2003;14(3):303–310.
91. El Fantroussi S, Naveau H, Agathos SN. Anaerobic dechlorinating bacteria. *Biotechnol Prog*. 1998;14(2):167–188.
92. Pas van de BA, Jansen S, Dijkema C, et al. Energy yield of respiration on chloroaromatic compounds in *Desulfotobacterium dehalogenans*. *Appl Environ Microbiol*. 2001;67(9):3958–3963.
93. Massa V, Infantino A, Radice F, et al. Efficiency of natural and engineered bacterial strains in the degradation of 4-chlorobenzoic acid in soil slurry. *International Biodeterioration & Biodegradation*. 2009;63(1):112–115.
94. Wittich RM, Wolff P. Growth of the genetically engineered strain *Cupriavidus necator* RW112 with chlorobenzoates and technical chlorobiphenyls. *Microbiology*. 2007;153(Pt 1):186–195.
95. Rodrigues JL, Kachel CA, Aiello MR, et al. Degradation of Aroclor 1242 dechlorination products in sediments by *Burkholderia xenovorans* LB400 (ohb) and *Rhodococcus* sp. strain RHA1 (fcb). *Appl Environ Microbiol*. 2006;72(4):2476–2482.
96. Monti MR, Smania AM, Fabro G, et al. Engineering *Pseudomonas fluorescens* for biodegradation of 2, 4-dinitrotoluene. *Appl Environ Microbiol*. 2005;71(12):8864–8872.
97. Boldt TS, Sorensen J, Karlson U, et al. Combined use of different Gfp reporters for monitoring single-cell activities of a genetically modified PCB degrader in the rhizosphere of alfalfa. *FEMS Microbiology Ecol*. 2004;48(2):139–148.
98. Rodrigues JL, Maltseva OV, Tsoi TV, et al. Development of a *Rhodococcus* recombinant strain for degradation of products from anaerobic dechlorination of PCBs. *Environ Sci Technol*. 2001;35(4):663–668.
99. Strong LC, McTavish H, Sadowsky MJ, et al. Field-scale remediation of atrazine-contaminated soil using recombinant *Escherichia coli* expressing atrazine chlorohydrolase. *Environ Microbiol*. 2000;2(1):91–98.
100. Halden RU, Tepp SM, Halden BG, et al. Degradation of 3-Phenoxybenzoic acid in soil by *Pseudomonas pseudoalcaligenes* POB310 (pPOB) and two modified *Pseudomonas* Strains. *Appl Environ Microbiol*. 1999;65(8):3354–3359.
101. Brazil GM, Kenefick L, Callanan M, et al. Construction of a rhizosphere *pseudomonad* with potential to degrade polychlorinated biphenyls and detection of bph gene expression in the rhizosphere. *Appl Environ Microbiol*. 1995;61(5):1946–1952.
102. Parameswarappa S, Karigar C, Nagenahalli M. Degradation of ethylbenzene by free and immobilized *Pseudomonas fluorescens*-CS2. *Biodegradation*. 2008;19(1):137–144.
103. Chen YM, Lin TF, Huang C, et al. Degradation of phenol and TCE using suspended and chitosan-bead immobilized *Pseudomonas putida*. *J Hazard Mater*. 2007;148(3):660–670.
104. Quek E, Ting YP, Tan HM. *Rhodococcus* sp. F92 immobilized on polyurethane foam shows ability to degrade various petroleum products. *Bioresour Technol*. 2006;97(1):32–38.
105. Adebusey SA, Illori MA, Amund OO, et al. Microbial degradation of petroleum hydrocarbons in a polluted tropical strain. *Journal of Microbiology and Biotechnology*. 2007;23(8):1149–1159.
106. Guo L, Bicki TJ, Felsot AS, et al. Phytotoxicity of atrazine and alachlor in soil amended with sludge, manure and activated carbon. *Journal of Environmental Science & Health Part B*. 1991;26(5-6):513–527.
107. Leoni V, Cremisini C, Giovanazzo R, et al. Activated sludge biodegradation test as a screening method to evaluate persistence of pesticides in soil. *Science of the Total Environment*. 1992;123:279–289.

108. Felsot AS, Dzantor EK. Enhancing biodegradation for detoxification of herbicide waste in soil. *ACS Symposium series*. 1990;426:249–268.
109. Abdelhafid R, Houot S, Barriuso E. How increasing availabilities of carbon and nitrogen affect atrazine behaviour in soils. *Biology and Fertility of Soils*. 2000;30(4):333–340.
110. Wagner SC, Zablotowicz RM. Effect of organic amendments on the bioremediation of cyanazine and fluometuron in soil. *Journal of Environmental Science & Health Part B*. 1997;32(1):37–54.
111. Topp E, Tessier L, Gregorich EG. Dairy manure incorporation stimulates rapid atrazine mineralization in an agricultural soil. *Canadian Journal of Soil Science*. 1996;76(3):403–409.
112. Printz H, Burauel P, Führ F. Effect of organic amendment on degradation and formation of bound residues of methabenzthiazuron in soil under constant climatic conditions. *Journal of Environmental Science & Health Part B*. 1995;30(4):435–456.
113. Hance RJ. The effect of nutrients on the decomposition of the herbicides atrazine and linuron incubated with soil. *Pesticide Science*. 1973;4(6):817–822.
114. Qiu Y, Pang H, Zhou Z, et al. Competitive biodegradation of dichlobenil and atrazine coexisting in soil amended with a char and citrate. *Environ Pollut*. 2009;157(11):2964–2969.
115. de Liphay JR, Sorensen SR, Aamand J. Effect of herbicide concentration and organic and inorganic nutrient amendment on the mineralization of mecoprop, 2, 4-D and 2, 4, 5-T in soil and aquifer samples. *Environ Pollut*. 2007;148(1):83–93.
116. Perrin-Ganier C, Schiavon F, Morel JL, et al. Effect of sludge-amendment or nutrient addition on the biodegradation of the herbicide isoproturon in soil. *Chemosphere*. 2001;44(4):887–892.
117. Wolfaardt GM, Lawrence JR, Robarts RD, et al. The role of interactions, sessile growth, and nutrient amendments on the degradative efficiency of a microbial consortium. *Can J Microbiol*. 1994;40(5):331–340.
118. Assaf NA, Turco RF. Influence of carbon and nitrogen application on the mineralization of atrazine and its metabolites in soil. *Pesticide Science*. 1994;41(1):41–47.