

Increased DNA Yield Following Enzymatic Release of *Borrelia* from a Collagen Matrix in Culture

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Sheila Wood* and Amy Rattelle

Advanced Laboratory Services, Rowan University, USA

***Corresponding author:** Sheila Wood, Advanced Laboratory Services, Rowan University, 501 Elmwood Ave, Sharon Hill, 19079 Pennsylvania, USA, Tel: 484 494 6125; Fax: 484 494 6800; Email: swood@advanced-lab.com

Received: March 04, 2015 | **Published:** March 27, 2015**Abstract**

Laboratory culture methods were developed to provide a means of harvesting greater amounts of DNA from *Borrelia* in culture. *Borrelia burgdorferi* B31 and *Borrelia burgdorferi* N40 were used in the initial experiments to develop optimal conditions. A collagenase enzyme preparation, Liberase TL, was used to release the organisms growing within a collagen matrix embedded in the culture. Optimal conditions of 2 U/mL of enzyme and 4 minutes of incubation at 37 °C were obtained. Following the establishment of these parameters, 24 positive patient samples were treated with Liberase and 24 positive patient samples were left untreated. The DNA harvested from both sets was quantified and evaluated for purity. Approximately 3.3-fold more DNA was harvested from the samples treated with Liberase TL than from the untreated samples. A greater degree of DNA purity, with less residual protein remaining, was also achieved.

Keywords: *Borrelia*; Enzyme**Introduction**

Investigators have shown that the ability of an organism to cause disease in the human often depends on its ability to attach to mammalian cells. *Borrelia burgdorferi*, the primary cause of Lyme Borreliosis in humans is no exception to this phenomenon. This organism is delivered to the dermis via the bite of the *Ixodes scapularis*, *Amblyomma americanum*, or *Ixodes pacificus* tick. Primarily extracellular and migratory, *Borrelia* actively moves from the dermis to the bloodstream. Further migration into extracellular matrix (ECM), connective tissue and other tissues of the body, including the heart may occur [1-3].

Cabello et al. [2] elucidated factors that influence the establishment of extracellular matrix (ECM) *in vivo*. ECM is secreted by cells to form an interstitial matrix and basement membrane, which constitute the framework to which cells are anchored. ECM appears to function in the sequestration, storage, and presentation of growth factors and also plays a role in regulation of cell growth, differentiation, and biochemical function. Attachment to and movement along collagen or collagen-containing structures plays an important role in the establishment of *Borrelia* in the ECM and its sequestration in the bodily tissues [2].

Zambrano et al. [1] have shown that the binding of *Borrelia* to collagen *in vitro* occurred within five minutes of inoculation and the binding increased over time, with maximum binding occurring at 1 hour. *Borrelia* remained bound to the collagen for 14 days in the presence of BSK-H media both with and without serum. Motile strains invaded collagen matrices on slides and formed micro colonies. Their experiments used collagen acquired from mouse tails that was extracted and solubilized using 3% acetic acid. The preparation that resulted contained Type I collagen but no decorin or proteoglycans or glycosaminoglycans which were removed using guanidine hydrochloride in Tris-HCl. Following extraction and treatment, the collagen was allowed to reassemble and form a lattice by adjusting the pH and concentration. Additionally, *Borrelia* bound to Type I (native) collagen lattices

at higher levels than to pepsinized collagen or BSA or gelatin. *B. burgdorferi* strains MC-1, N40 and B31 showed no variation in their ability to bind type I collagen matrices when collected and used at early, mid or late log phase. On proteinase K treated surfaces, *B. burgdorferi* showed less adherence, suggesting a role of protein in adherence. Ten percent of the bacterial inoculum still bound collagen after surface treatment with proteinase K.

Our laboratory culture for *Borrelia* incorporates collagen coated slides as part of the culture process. The presence of a collagen surface within the culture exploits the organism's natural ability to bind to collagen and components within the ECM *in vivo* as described by Cabello et al. [2]. The matrix allows organisms to attach and form micro colonies [4]. Although organisms are present upon harvesting, optimal DNA recovery requires the severing of attachment bonds between organisms and collagen. Our objective in this study was to develop a method that would result in the harvesting of an optimal quantity of DNA from organisms growing within the collagen matrix.

The enzyme chosen to release the organisms from the collagen was Liberase TL (Roche). Proteolytic enzymes such as this are commonly used for cell harvesting and tissue dissociation. Proteases are used to disrupt the extracellular matrix of tissue to allow the release of individual cells that are viable and functionally active. (Roche product specification sheet 2010). The collagen coated slide used in the *Borrelia* culture is coated with Rat Tail Collagen Type I derived from tendon (Corning). The slides are suitable for both serum-containing and serum-free cultures and promote cell attachment, spreading, growth and/or differentiation for a variety of cells. Sapi et al. [4] used this collagen matrix in culture to harness the natural ability of *Borrelia* to adhere and form micro colonies.

In our experiments, two strains of *Borrelia burgdorferi* were used to determine the efficacy of Liberase to improve DNA yield from culture. First, the tick derived *B. burgdorferi* B31 [5] was used to assist in quantifying DNA recovery. *B. burgdorferi* B31 is a

well-documented strain of *Borrelia* commonly used in laboratory manipulations, and used as a positive control to validate diagnostic tests. It has been described as a strain that has a decreased ability to cause infection [6,7]. The second strain was *B. burgdorferi* N40 clone D10/E9 [1]. This strain, also isolated from a tick, harbors a more complete genetic repertoire of plasmids and regulated genes that are involved in the infectious process than *B. burgdorferi* B31. It is pathogenic in the mouse model and genetic/adhesion/protein studies show differences from B31 *in vitro* [7]. Both were used to assess colonization, and subsequent harvest efficiency, from within the collagen matrix nested in our culture.

Although neither strain of *Borrelia* showed differences in adherence to Type I collagen in previous experiments [1], our approach chose to use both organisms to assess possible differences in attachment to, or release from, our matrix. This study sought to determine if Liberase TL is an appropriate enzyme to release organisms from our collagen-containing cultures and achieve an increase in the yield of DNA [8].

Materials and Method

Borrelia culture

The *Borrelia* culture media used is described by Sapi et al. [4]. Both *B. burgdorferi* B31 and N40 were maintained separately in this medium and used in these experiments prior to the tenth passage. One group of cultures contained the collagen-coated slides and one group did not contain the slides. Organisms were obtained from the ATCC (*B. burgdorferi* B31 ATCC # 35210) [9] and from Dr John Leong, Tufts University School of Medicine, *B. burgdorferi* N40 clone D10/E9.

Effects of liberase on DNA yield

The possible detrimental effects of Liberase on the DNA were tested using *B. burgdorferi* B31. The culture medium with no collagen-coated slide was seeded using 1×10^6 organisms. The culture was allowed to grow for seven days at 34 °C, 5% CO₂, and high humidity. The culture was divided into 18, 2 mL micro centrifuge tubes. Cell pellets were obtained by centrifugation. Pellets containing approximately 6×10^7 organisms each, were treated with increasing concentrations (0.1, 0.25, 0.50, 0.75, 1.00, 1.25, 1.50, 1.75, and 2.0 U/mL) of Liberase TL prepared in sterile distilled water, for eight minutes at 37 °C. Reactions were stopped using a final concentration of 0.1 mM EDTA prepared in PBS. The tubes were then centrifuged at 9,600 x g for 10 minutes and the supernatant was discarded. DNA was extracted from the organisms remaining in the pellets using the QIAamp Mini Kit (Qiagen Cat # 51306). DNA concentrations were measured using a Nanodrop 2000. Each treatment condition had a negative control not treated with Liberase TL, but keeping all other conditions the same.

Enzymatic treatment of *B. burgdorferi* B31 grown in BSK-H containing collagen coated slides

Previously established culture procedures were used [4]. *B. burgdorferi* B31 was used at a concentration of 1×10^6 cells to seed culture media. Four Coplin jars, each containing four collagen-coated slides, were seeded and allowed to grow at 34 °C in 5% CO₂ and high humidity for 28 days. Three collagen-coated slides were removed and treated with Liberase. One slide was used as

a control and not treated with Liberase. The concentrations of Liberase TL, used to treat the pellets containing *Borrelia* attached to collagen, were 0.25, 0.50, 0.75, and 1 U/mL. One mL of each concentration of Liberase was added to each slide and the slides were incubated horizontally at 37 °C for 1, 4, or 8 minutes. Each slide was washed by tipping, rinsing with 1 mL PBS collecting run-off, and then pipetting the run-off over the slide repeatedly. The run-off was then collected in a 1.5 mL micro centrifuge tube and 10 µL 0.1M EDTA was added to stop enzymatic activity.

Growth of *B. burgdorferi* N40

B. burgdorferi B31, while used as laboratory control strain, has been described as non-infectious to humans [9]. Further experiments were conducted using *B. Burgdorferi* N40 which has infectious capabilities in mice [5]. Twelve Coplin jars containing two collagen-coated slides and a total volume of 25 mL each were inoculated with 1×10^7 N40 bacterial cells and grown at 34 °C in 5 % CO₂ and high humidity.

Enzymatic treatment of *B. burgdorferi* N40 pellets harvested from cultures grown for two weeks in BSK-H containing collagen coated slides

A two-week growth of *B. burgdorferi* N40 in culture containing the collagen slides was harvested. The slides were scraped and the entire preparation was centrifuged at 9600 x g for 10 minutes. Concentrations of 1 U/mL and 2 U/mL Liberase TL at 1 minute and 4 minutes, were used to treat the pellets and the reaction was stopped with 0.1M EDTA.

Purification and analysis of recovered DNA

Removal of human DNA: Prior to extracting *Borrelia* DNA from the patient samples, human DNA was removed using the method of Zhou and Pollard [10]. This method uses Oxgall (5%) to lyse mammalian cells and Micrococcal nuclease to inactivate mammalian DNA. Controls were performed to ensure that *Borrelia* DNA would remain intact during this process. Concentrations of Oxgall at 1%, 1.5%, 2%, 5%, and 10% in PBS were mixed with *B. burgdorferi* N40 at a concentration of 2.6×10^7 cells, incubated at 37 °C for 10 minutes, and densities were then read in a counting chamber.

DNA purification and measurement: *Borrelia* DNA was then extracted using magnetic bead technology (MagJet Genomic DNA Kit, ThermoFisherScientific). Following extraction, it was measured using the Nanodrop 2000 at both 260nm (DNA) and 280nm (protein). Ratios of DNA to protein were obtained as a measure of purity. Ratios closer to 2.0 indicate high purity of DNA with little protein content.

Results

Results of the effects of liberase on DNA yield

Treatment with Liberase TL for 8 minutes showed detrimental effects on the DNA. The optimal release and thus recovery of DNA occurred at 0.50 U/mL, but other concentrations were ineffective in releasing intact organism DNA. The erratic results emphasized the need to achieve an optimal concentration and time interval when using this enzyme for release of *Borrelia* from collagen (Figure 1).

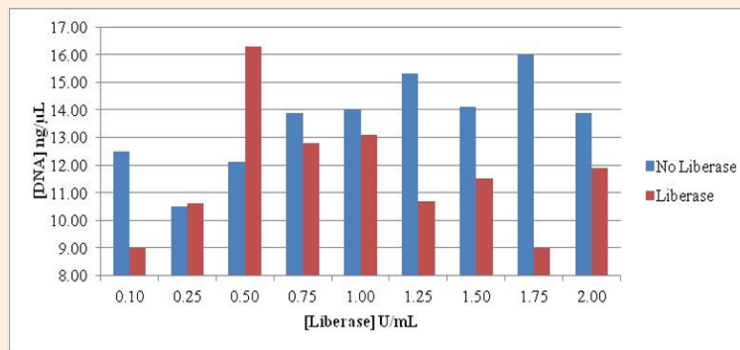


Figure 1: Intact DNA remaining after treatment of culture pellets containing *B burgdorferi* B31. The concentrations of Liberase TL ranged from 0.10 to 2.00 U/mL. Organisms were exposed to Liberase TL for 8 minutes.

Results of enzymatic treatment of *B. burgdorferi* B31 grown in BSK-H containing collagen coated slides

Further attempts to optimize the concentration of Liberase TL

and the time of exposure (Table 1), showed very little recovery of DNA when using less than or equal to 1 U/mL. Time periods of 1, 4 and 8 minutes and concentrations of 0, 0.25, 0.50, 0.75 and 1.0 U/mL were used (Table 2).

Table 1: Concentrations of Liberase TL and time intervals used.

| Treatment | Liberase Treatment (U/mL) | | | | |
|------------|---------------------------|------|-----|------|---|
| Time (min) | 0 | 0.25 | 0.5 | 0.75 | 1 |
| 1 min | 0 | 0.25 | 0.5 | 0.75 | 1 |
| 4 min | 0 | 0.25 | 0.5 | 0.75 | 1 |
| 8 min | 0 | 0.25 | 0.5 | 0.75 | 1 |

Table 2: DNA recovered from *B. burgdorferi* B31 grown in BSK-H containing collagen coated slides after 28 days. Liberase concentrations ranged from 0.25 U/mL to 1U/mL. Times intervals were 1,4 and 8 minutes.

| Liberase conc U/mL | Treatment time minutes | DNA ng/uL from each slide |
|--------------------|------------------------|---------------------------|
| 0 | 1 | 3.4 |
| 0.25 | 1 | 2.2 |
| 0.5 | 1 | 0.6 |
| 0.75 | 1 | 2.8 |
| 1 | 1 | 4.4 |
| 0 | 4 | 1.4 |
| 0.25 | 4 | 4 |
| 0.5 | 4 | 0.9 |
| 0.75 | 4 | 2.8 |
| 1 | 4 | 0.1 |
| 0 | 8 | 0.6 |
| 0.25 | 8 | 2.3 |
| 0.5 | 8 | 2.5 |
| 0.75 | 8 | 3 |
| 1 | 8 | 3.7 |

Results of enzymatic treatment of *B. burgdorferi* N40 pellets harvested from cultures grown for two weeks in BSK-H containing collagen coated slides

An optimal time of 4 minutes incubation at 37 °C in combination with an optimal concentration of 2 U/mL Liberase TL gave recovery of 51% more DNA than the control. As shown in Figure 2, no treatment with Liberase gave DNA levels from 1.4 to 4.8 ng/

uL. DNA concentrations obtained after treatment with either 1 or 2 U/mL Liberase for 1 minute (with scraping) ranged from 7.5 to 10.5 ng/uL. The greatest DNA yield was 20.6 ng/uL and was obtained after treating culture pellets with 2 U/mL Liberase for 4 minutes. Liberase treatment of culture pellets plus material and organisms scraped from the collagen coated slides resulted in 51% more DNA retrieval.

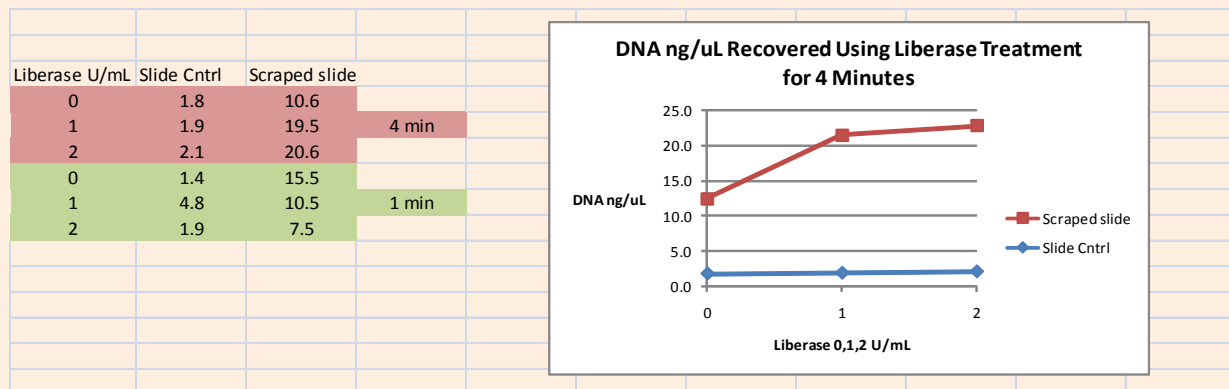


Figure 2: *B. burgdorferi* N40 DNA harvested from BSK-H cultures containing collagen-coated slides. Concentrations of 0, 1, and 2 U/mL; times of 1 and 4 minutes.

Results of purification and analysis of recovered DNA

Release of *Borrelia* from collagen: Using the optimal treatment conditions for release of *Borrelia* from collagen, 24 patient samples each of treated and untreated pellets showed that the samples treated with Liberase TL had 3.3-fold more intact DNA than the untreated pellets.

Removal of human DNA: Controls for the Oxgall treatment for removal of human DNA showed that at all concentrations of Oxgall, 1.0%, 1.5%, 2.0%, 5.0%, and 10%, approximately 1.5×10^7 long spirochete forms were recovered (data not shown) showing that Oxgall could be used to remove human DNA with optimal *Borrelia* cells remaining.

Purification of the DNA: Purification using the Magnetic Bead

technology MagJet kit yielded optimal concentrations of DNA on both Liberase treated and Liberase untreated samples.

Amounts of DNA recovered from liberase treated and untreated samples

The amounts of DNA obtained from untreated samples ranged from 2.9 to 5.6 ng/uL whereas the amounts of DNA recovered from Liberase treated samples ranged from 7.2 to 30.7 ng/uL. The purity of the DNA from these samples was also higher as they showed less contamination with protein (Table 3 and Figure 3). Ratios of 260nm/280nm, (DNA/impurities), were obtained. Ratios closer to 2.0 indicate high purity of DNA with little protein content. Ratios in the untreated samples ranged from 1.03 to 1.89 whereas ratios in the treated samples ranged from 1.74 to 2.03.

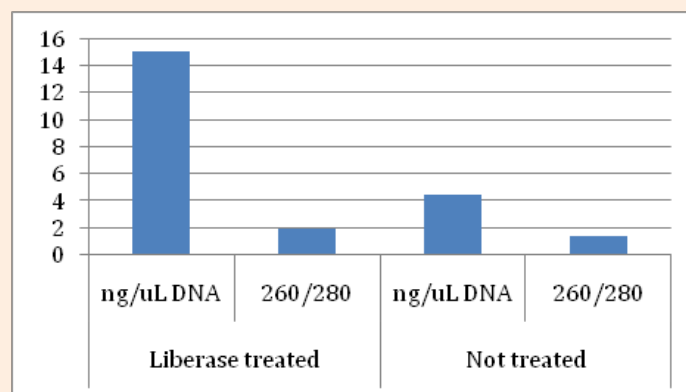


Figure 3: Average DNA yields and purity of pellets treated with Liberase versus pellets not treated with Liberase. DNA was analyzed using the Nanodrop 2000 and ratios of 260/280 were obtained; 260 nm for DNA, 280 nm for impurities such as protein.

Liberase treated (ng/uL): DNA - 15; 260/280 - 1.95
 Not treated (ng/uL): DNA - 4.5; 260/280 - 1.4

Table 3: Comparison of 24 patient samples from culture treated with Liberase during harvesting and 24 patient samples from culture not treated with Liberase during the harvesting process. Following Liberase treatment of the pellets, samples were treated for the removal of human DNA (Zhou and Pollard, 2012). The remaining *Borrelia* DNA was then extracted and purified using the MagJet Genomic DNA Kit. (Thermo Fisher Scientific). The resultant DNA was analyzed using the Nanodrop 2000 and ratios of 260/280 were obtained; 260 nm for DNA, 280 nm for impurities such as protein.

| Not treated with Liberase | | | | Treated with Liberase | | | |
|---------------------------|---------|-------|---------|-----------------------|---------|-------|---------|
| Lab # | Study # | ng/uL | 260/280 | Lab # | Study # | ng/uL | 260/280 |
| 5260 | 8MS1 | 4.8 | 1.89 | 5338 | 8MS49 | 15.6 | 1.98 |
| 5261 | 8MS2 | 4.8 | 1.66 | 5339 | 8MS50 | 15.9 | 1.9 |
| 5263 | 8MS3 | 4.6 | 1.64 | 5279 | 8MS51 | 30.7 | 1.74 |
| 5264 | 8MS4 | 5.4 | 1.32 | 5299 | 8MS52 | 16.3 | 2.03 |
| 5265 | 8MS5 | 6.3 | 1.63 | 5397 | 8MS53 | 16 | 1.93 |
| 5266 | 8MS6 | 7.7 | 1.37 | 5398 | 8MS54 | 14.1 | 1.88 |
| 5269 | 8MS7 | 5.6 | 1.48 | 5399 | 8MS55 | 17.8 | 1.95 |
| 5270 | 8MS8 | 5 | 1.83 | 5400 | 8MS56 | 16.6 | 2.02 |
| 5273 | 8MS9 | 5.6 | 1.45 | 5401 | 8MS57 | 20.7 | 1.82 |
| 5277 | 8MS10 | 5 | 1.52 | 5404 | 8MS58 | 24.4 | 1.81 |
| 5278 | 8MS11 | 5.1 | 1.5 | 5408 | 8MS59 | 18 | 1.88 |
| 5280 | 8MS12 | 4.7 | 1.81 | 5409 | 8MS60 | 15.6 | 2.2 |
| 5282 | 8MS13 | 3.1 | 1.18 | 5340 | 8MS61 | 14.2 | 2.3 |
| 5283 | 9MS14 | 3.8 | 1.45 | 5341 | 8MS62 | 11.9 | 1.92 |
| 5284 | 8MS15 | 4.4 | 1.33 | 5342 | 8MS63 | 10.9 | 2.1 |
| 5285 | 8MS16 | 3.3 | 1.28 | 5343 | 8MS64 | 13.7 | 1.96 |
| 5287 | 8MS17 | 4.2 | 1.41 | 5345 | 8MS65 | 14.6 | 1.96 |
| 5289 | 8MS18 | 3.1 | 1.63 | 5349 | 8MS66 | 13.2 | 1.87 |
| 5290 | 8MS19 | 3.4 | 1.11 | 5350 | 8MS67 | 9.4 | 2.03 |
| 5295 | 8MS20 | 2.9 | 1.27 | 5351 | 8MS68 | 9.4 | 1.97 |
| 5296 | 8MS21 | 4.1 | 1.18 | 5358 | 8MS69 | 15.7 | 1.88 |
| 5299 | 8MS22 | 3.6 | 1.17 | 5359 | 8MS70 | 11.5 | 1.76 |
| 5300 | 8MS23 | 3.6 | 1.45 | 5360 | 8MS71 | 11.2 | 1.87 |
| 5302 | 8MS24 | 3 | 1.03 | 5366 | 8MS73 | 7.2 | 2.04 |
| Average | | 4.46 | 1.44 | | | 15.19 | 1.95 |

Discussion

Often it is assumed that primary virulence factors of microbes are genetically controlled in response to their immediate environment. In experiments using *Vibrio cholerae*, Anjali et al. [11] showed that the regulation of virulence factors occurs in the body and is very different from genetic regulation in laboratory media. Pathogens adapt to the host environment by altering their gene expression. Using RNA sequence based transcriptome analysis of *V. cholerae* during infection in rabbits and mice, data was obtained for the recognition of regulatory response during infection. Genes and small RNAs were discovered that previously had not been linked to virulence. These studies clarified the need to examine genetic regulation in response to an internal mammalian system rather than just from studies performed in the laboratory.

DeRita et al. [12] contrasted the complex regulatory pathway in *V. cholerae* that evolved to control virulence by induction of gene expression *in vivo* with that of *Streptococcus pyogenes*

that exhibits at least one mode of pathogenesis that manifests in the absence of regulation. Both organisms respond to their environment in differing adaptive ways. Additionally, Moreira et al. [13] elucidated the effects of a global regulator, QseC, in *Salmonella enterica serovar typhimurium*, which regulates genes within pathogenesis islands I and II both *in vitro* and *in vivo*. Bent et al. [14] showed that the chromosomally encoded Ysa T3SS in *Yersinia enterocolitica* is expressed in a contact dependent manner *in vitro* and is also expressed *in vivo* during infection in mice. The differing strategies used by organisms to survive and thrive in their environments reflect evolutionary adaptation and contribute to pathogenesis.

The production of adhesins by *Borrelia* occurs in certain types of media but not others. Studies have shown the presence or absence of BBK32, an adhesion that allows the organism to attach to fibronectin, depending on the growth media and conditions that the organism is experiencing in its immediate environment [15,16]. Two adhesins that *Borrelia* produces for attachment to collagen have recently been elucidated, CspA and CspZ [17].

These outer surface glycoproteins may influence the migration and sequestration of the organism in the human body. Although investigators defend differing positions as to the presence or absence of such factors and whether or not they constitute virulence factors, it is clear that migration through tissue in the human body can be aided by attachment to and then penetration of tissue. If the organism did not have a way to anchor itself to the ECM, vascular epithelium, and/or other components within the blood that wash the tissue, then its ability to sequester and survive in the niches of the human body would not lead to chronic infection and evasion of the immune system. Taking these assumptions into consideration, the demonstration of and the exploitation of the organism's survival mechanisms has greatly facilitated our ability to grow and harvest *Borrelia* that has attached to collagen *in vitro*.

Borrelia and its interaction with interstitial components of the mammalian body have led investigators to incorporate some of these components into *in vitro* cultivation methodology [1,4]. One of the most studied means of attachment is the interaction of *Borrelia* with collagen, both *in vivo* and *in vitro* [2]. Two of the known adhesins that *Borrelia* uses to bind collagen include CspA and CspZ [13]. Although they are recognized as adhesins, the molecular regulation of their production has not been studied in cultures of human blood, so it is not known if their production is down regulated or up regulated in blood culture. Nevertheless, the ability of the organism to thrive in culture and interact with collagen becomes evident in our study of the present methodology.

Our improved method for harvesting *Borrelia* results in the recovery of higher concentrations of DNA. The organisms attachment to, and growth in, the collagen matrix in culture is used to our advantage in the new harvesting procedure. Their release from the collagen, after incubation with Liberase, results in our ability to harvest more biomass, and thus more DNA. In addition, the DNA obtained is less contaminated with residual protein.

Conclusion

Our laboratory was able to increase the concentration and purity of *Borrelia* DNA harvested from culture pellets by using an enzyme preparation capable of breaking bonds within the collagen matrix and also between *Borrelia* and collagen. The preparation, Liberase TL, is composed of collagenases and neutral proteases. The release of *Borrelia* from the collagen matrix by the enzyme preparation was concentration and time dependent. Optimal enzymatic treatment conditions yielded 3.3-fold more DNA. The DNA isolated had higher purity with less residual protein than that harvested from untreated pellets.

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