

In-Vitro Screening on the Interaction of *Chaetomium Cellulolyticum* Chachal & *D. Hawksworth* with *Aspergillus flavus* Link Ex Fr. and *Curvularia lunata* (Wakker) Boedijin

Abstract

This study generally aimed to determine the interaction of *Chaetomium cellulolyticum* with *Aspergillus flavus* and *Curvularia lunata* and the antifungal properties of *Chaetomium cellulolyticum* extracts against *Aspergillus flavus* and *Curvularia lunata*. Description of the fungal microscopic association was done in slide bi-culture while the zone of colonization was studied in the co-culture method and zone of inhibition was determined in the antimicrobial bioassay. Results revealed that in slide bi-culture test, the hyphae of *A. flavus* were damaged when grown together with *C. cellulolyticum*. Co-culture method revealed that *C. cellulolyticum* does not colonize the agar lawn of *A. flavus*. In bioassay, results revealed that at 24th to 48th hours of incubation, mycelial crude hexane and ethanol extracts do not produced zone of inhibition on *A. flavus*. Statistical analysis showed that the zone of inhibition of the two crude extracts were comparable to the positive and negative control. However, at 72nd to 96th hours, the extracts were significantly smaller to the positive control and comparable to the negative control ($p < 0.05$).

In slide bi-culture test revealed that the hyphae and conidia of *C. lunata* were broken when grown together with *C. cellulolyticum*. In Co-culture method results revealed that *C. cellulolyticum* colonized the agar lawn of *C. lunata* by 0.15%. In Bioassay, results revealed that mycelial crude hexane and ethanol extracts do not produced zones of inhibition on *C. lunata* at 24th to 96th hours of incubation. Analysis of variance and comparison among means showed, that inhibitions of two extracts were comparable to the positive and negative control at 24th hours of incubation. However, at 48th to 96th hours of incubation, the crude extracts were significantly smaller to the positive control but comparable to the negative control ($p < 0.05$). *Chaetomium cellulolyticum* have antibiosis mechanism to deform or damage the morphological hyphae and either conidia of *A. flavus* and *C. lunata*. However, the affectivity of crude hexane and ethanol extracts from *C. cellulolyticum* were not observed to inhibit the growth of *A. flavus* and *C. lunata*.

Keywords: Plant pathogens; Fungicides; Antagonistic activity

Research Article

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Abbreviations: LSD: Laboratory Service Division; CPC: Crop Protection Cluster; PDB: Potato Dextrose Broth; PDA: Potato Dextrose Agar; DMSO: Dimethyl Sulfoxide; CRD: Completely Randomized Design; ANOVA: Analysis of Variance; DMRT: Duncan's Multiple Range Test; SPSS: Statistical Package for Social Sciences

Introduction

Background of the study

The search for natural fungicides has been dominating the research arena in the past years. The Environmental Protection Agency [1] described fungicides as pesticides applied to kill or inhibit growth of fungi that cause economic damage. Copper compounds, especially copper sulfate mixed with lime and water and sulfur and synthetic organic compounds have long been used for this purpose. EPA [1] added that plants, fungi and microorganisms contain antifungal substances that occur

naturally that they could be tapped as possible source. The ability of many fungi to antagonize various microorganisms and other fungi has been well established. In most cases, attention has been centered upon the antagonistic actions of various fungi against organisms capable of causing plant diseases. EPA [1] listed fungi in the groups of *Ophiobolus*, *Rhizoatonia*, and *Fusarium*. *Aspergillus* and *Chaetomium* capable of antagonizing pathogenic organisms. The search for promising microbial antagonists have been of increasing interest and used to control plant diseases [2].

Seeds are regarded as highly effective means for transporting plant pathogens over long distances. Seed-borne diseases have been found to affect the growth and productivity of crop plants [3]. A seed-borne pathogen present externally or internally or associated with seed as contaminant, may cause seed abortion, seed rot, seed necrosis, reduction or elimination of germination capacity as well as seedling damage resulting in development of disease at later stages of plant growth by systemic or

local infection [4]. Moisture and seed nutrient content in the midrange of seed development provide optimum conditions for fungal development and toxin production. Mycotoxins are carcinogenic metabolites produced by fungus such as aflatoxins liberated by pathogenic fungus *Aspergillus flavus* Link ex Fr. and Phytotoxins produced by *Curvularia lunata* Boedijin [5] as contaminations in maize (*Zea mays* L.) grain. Aflatoxin contamination of maize kernels poses a serious health hazard to both humans and animals [6]. *A. flavus* infects kernels via the corn silk or in association with insect damage. They are colored yellow green and as the spores age they turn a dark green [7]. *C. lunata* often formed on the edge of the leaf and become brown, oval to irregular. It is also reported to cause head blight in foreign countries [8]. *Chaetomium* is a large ascomycetous fungus producing perithecia. It is found on a variety of substrates containing cellulose including paper and plant compost. It can be readily found on the damaged paper in sheetrock. *Chaetomium* spp. have a worldwide distribution. They have biological control in plant diseases especially vegetable and fruits. They protect plants from diseases by producing many kinds of antibiotics and ergosterols that can suppress plant pathogens, stimulate growth of plants and induce resistance of plants [9]. Biological products are useful, not only for the protection against plant diseases, but can also be used for curative effects of plant diseases, and also promote plant growth [10].

Significance of the study

Many bioactive compounds from different fungi have been reported to inhibit many plant pathogenic fungi. Results of this study created an impact and significant contribution in biotechnology, particularly in the field of agriculture to limit the damage of plant pathogens in both conventional and organic farming. The information achieved was provided full support to develop new generation of bio-fungicide that will substitute to the hazardous chemical fungicides against seed-borne fungal pathogens, and promoted plant disease bio-control in practice not only in the Philippines but also in the world.

Scope and limitation of the study

Strains of seed-borne fungal pathogens (*Aspergillus flavus* and *Curvularia lunata*) and *Chaetomium cellulolyticum* that were used in this study were obtained at the UP-Natural Sciences Research Institute Culture Collection (UPCC) of University of the Philippines, Diliman, Quezon City; Laboratory Service Division (LSD) of Philmec, Science City of Munoz, Nueva Ecija and the Department of Plant Pathology, Crop Protection Cluster (CPC) of College of Agriculture, University of the Philippines, Los Banos, Laguna. This study was focus only on the interactions between *C. cellulolyticum* and the pathogens, and on antagonistic effect of *Chaetomium cellulolyticum* against seed-borne fungal pathogens.

Time and place of the study

This study was conducted from December 2010 to March 2011 at the Department of Biological Sciences, College of Arts and Sciences, Central Luzon State University, Science City of Munoz, Nueva Ecija, and at PhD Biotechnology in Plant Pathology Laboratory, Biocontrol Research Unit and Mycology Section, Department of Plant Pest Management, Faculty of Agricultural Technology, King Mongkut's Institute of Technology Ladkrabang,

Bangkok, Thailand.

Objectives of the Study

This study generally aimed to determine the interaction of *Chaetomium cellulolyticum* with *Aspergillus flavus* and *Curvularia lunata* and the antifungal properties of *Chaetomium cellulolyticum* extracts against of *Aspergillus flavus* and *Curvularia lunata*. It specifically aimed to:

- i. Describe the interaction between *C. cellulolyticum* and *A. flavus*.
- ii. Determine the zone of colonization of *C. cellulolyticum* on agar lawn of *A. flavus*.
- iii. Determine the zone of inhibition of mycelial crude extracts of *C. cellulolyticum* on *A. flavus*.
- iv. Describe interaction between *C. cellulolyticum* and *C. lunata*.
- v. Determine the zone of colonization of *C. cellulolyticum* on agar lawn of *C. lunata* and
- vi. Determine the zone of inhibition of mycelial crude extracts of *C. cellulolyticum* on *C. lunata*.

Review of Related Literature

Chaetomium spp.

Chaetomium is a large fungal genus species of which are found worldwide on variety of substrates including damp sheetrock, carpet, plant compost, soil and other substrates containing cellulose. It is also commonly found on deteriorating wood products. *Chaetomium* is an ascomycete that produces ascospores in dense, hairy fruiting bodies called perithecia. Perithecia are flask-shaped structures with a hole through which spores are extruded. The perithecial hairs can take a variety of forms depending upon the species. The ascospores are dark brown and often lemon-shaped. They collect in a dense mass just outside the perithecium. The perithecia, surrounded by the long hairs, can be as large as 1 mm, and are visible to the naked eye. Surfaces supporting *Chaetomium* growth with sporulation may superficially resemble old colonies of *Stachybotrys*. In culture, colonies of *Chaetomium* are rapidly growing, cottony and initially white in color. As the colonies mature, they become gray to olive in color and from the reverse the color is tan to red or brown to black. *Chaetomium* species grow best at between 25 and 35 °C [11]. Charoenporn et al. [10] used Potato Dextrose Broth (PDB) media for mycelial matting of *Chaetomium* sp. The optimum pH of PDB for *Chaetomium* is at 4.5 to 5.5 pH. *Chaetomium cellulolyticum* is a thermo tolerant fungus rather than thermophilic, optimal growth resulted at 37 °C but temperatures above 40 °C were inhibitory.

Materials and Methods

Test organisms

Preparation of samples: The strain of *Chaetomium cellulolyticum* and seed-borne fungal pathogens (*Aspergillus flavus* and *Curvularia lunata*) were obtained from the UP-Natural Sciences Research Institute Culture Collection (UPCC) of

University of the Philippines, Diliman, Quezon City; Laboratory Service Division (LSD) of Philmec, Science City of Munoz, Nueva Ecija and the Department of Plant Pathology, Crop Protection Cluster (CPC) of College of Agriculture, University of the Philippines, Los Banos, Laguna. Sterilized Potato Dextrose Agar (PDA) was used as medium for the mycelial propagation of the samples. The pure cultures of the organisms were used as source and sub culturing was done by touching the spores using sterile inoculating needle. Triple point inoculation, where in the distances of the three (3) point growth are equal, was used in inoculating the medium. These sub-cultures were incubated at room temperature.

Interaction studies

The slide bi-culture test and co-culture method were done to determine interactions of the *C. cellulolyticum* with plant pathogens.

Slide bi-culture: Ten milliliter (10 ml) of sterilized PDA was poured in the Petri dish. After it had cooled, this was sliced into blocks like the size of the cover slip using sterile scalpel and this was aseptically inoculated using the L- shape inoculating needle in the sterile glass slide. The spores at the edge of radial mycelial growth of the *C. cellulolyticum* was touched by sterile inoculating needle and used to inoculate the agar block and the sterile cover slip was placed. Same procedure was done using the pathogen. The distance between the antagonist and the pathogen was approximately one centimeter (1 cm). The slide was placed in a sterile Petri dish and was lined with sterile paper as moist chamber. This was replicated three (3) times and incubated at room temperature. This was periodically observed everyday under compound microscope. The interaction of the antagonist and the pathogen was observed under photomicroscope at PhD Biotechnology in Plant Pathology Laboratory, Biocontrol Research Unit and Mycology Section, Department of Plant Pest Management, Faculty of Agricultural Technology, King Mongkut's Institute of Technology Ladkrabang, Bangkok, Thailand.

Co-culture method: Co-culture method was conducted to determine if *C. cellulolyticum* could colonize the agar lawn of the plant pathogens using the method of Soyong [12]. Potato Dextrose Agar (PDA) at pH 7 was poured in the Petri plate and marked into two sections. Aseptically, approximately six millimeters (6 mm) of the seven (7) day old culture of *C. cellulolyticum* was bored using a sterile cork borer and this was inoculated at the half section of the agar along the middle line of the agar. The disc was tapped lightly to make the disc attach to the surface. The same method was applied using the pathogen. The Petri plates were marked properly and these were incubated in inverted position at room temperature. During incubation, the fungus that first crossed the midline was recorded. The zone of colonization of *Chaetomium cellulolyticum* was measured in millimeters (mm) using vernier caliper and this was done by measuring along the middle mark. To determine the area of colonization, the width and the length of the mycelial growth was measured in every twenty four hours (24 hrs).

Extract bio assay

Experimental treatments: The study also looked into the inhibitory activities of crude extracts of *C. cellulolyticum* against

the plant pathogens, *A. flavus* and *C. lunata*. There were four (4) treatments used in this study and each treatment was replicated five (5) times. These treatments were:

- i. T1- Hexane extract of *C. cellulolyticum*
- ii. T2- Ethanol extract of *C. cellulolyticum*
- iii. T3- Positive Control (Dithane M-45)
- iv. T4- Negative Control (Dimethyl sulfoxide)

Preparation of crude extract

Preparation of mycelial mat: One hundred milliliter (100 ml) Potato Dextrose Broth (PDB) at pH 5.5 was used as medium for propagation of *Chaetomium cellulolyticum*. The broth was sterilized in autoclave at one hundred twenty one degree Celsius (121 °C) for thirty minutes (30 mins). The seven (7) day old culture *C. cellulolyticum* was sliced into blocks with sterilized scalpel and this was aseptically inoculated in the prepared broth using sterile inoculating needle and incubated at room temperature for 30 days.

Ethanol and hexane extraction: After thirty (30) days, the fungal mycelia were removed from broth using sterile forceps and were placed in Petri dish to dry over night at room temperature. The extraction of *C. cellulolyticum* was performed by the method describe by Kanokmedhakul et al. [13]. Ten grams (10 g) dried mycelial mat was submerged and extracted in one hundred milliliter (100 ml) 95% hexane and 95% ethanol for twenty four hours (24 hr) and was incubated at room temperature. And after extracted this was filtered using filter paper (Whatman no. 1). The solvents were separately evaporated in rotary evaporator under reduced pressure at fifty degree Celsius (50°C) to yield crude hexane and ethanol respectively and this was done at Rice Chem Laboratory, Phil Rice, Maligaya, Science City of Munoz, Nueva Ecija. Each extract was dissolved in 2% Dimethyl sulfoxide (DMSO).

Paper disc bioassay

Lawn preparation: Suspension of *A. flavus* and *C. lunata* were prepared by touching the spores using sterile inoculating needle and mixing in the ten milliliter (10 ml) distilled water. Dilution was done and spore concentration of 1×10^6 spores/ml of each fungus was determined using sterile heamacytometer and it was mixed with a prepared sterile PDA media. The paper discs were submerged in Ethanol extract, Hexane extract, Dithane M-45 and DMSO for twenty four (24) hours. Each lawn of the pathogens, two (2) discs was placed according to treatments. Each treatment was replicated five (5) times. The treatments were marked properly and this was incubated at room temperature for four (4) days.

Measuring of the zone of inhibition: During incubation, the inhibition zones from the center of the discs to the inner margin of the surroundings of fungal growth was measured in millimeters using vernier caliper in every twenty four hours for four (4) days and this was recorded.

Data gathered

The data gathered were:

- i. The interaction between *C. cellulolyticum* and *A. flavus*.
- ii. The zone of colonization of *C. cellulolyticum* on agar lawn of *A. flavus*.
- iii. The zone of inhibition of mycelial crude extracts of *C. cellulolyticum* on *A. flavus*.
- iv. Interaction between *C. cellulolyticum* and *C. lunata*.
- v. The zone of colonization of *C. cellulolyticum* on agar lawn of *C. lunata* and
- vi. The zone of inhibition of mycelial crude extracts of *C. cellulolyticum* on *C. lunata*.

Experimental layout

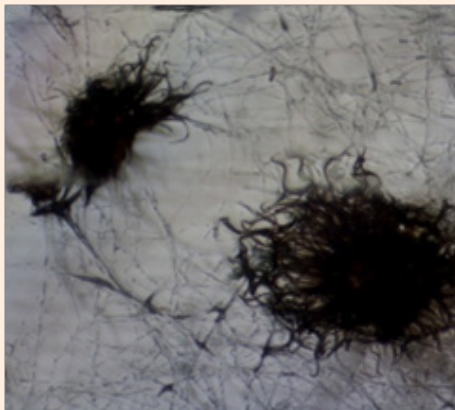
The experiment was laid out following Completely Randomized Design (CRD). The fungal interactions were qualitatively described and documented. Zones of inhibition were analyzed using Analysis of Variance (ANOVA) and compared using Duncan's Multiple Range Test (DMRT) at 5% level of significance. Statistical Package for Social Sciences (SPSS) was used to analyze the data.

Results and Discussion

Interactions of *Chaetomium cellulolyticum* and *Aspergillus flavus*

The interactions of *C. cellulolyticum* and *A. flavus* were studied using the slide bi-culture, co-culture techniques and extract bioassay. Description of the fungal microscopic association was done in slide bi-culture while the zone of colonization was studied in the co-culture method and zone of inhibition was determined in the antimicrobial bioassay.

Slide bi-culture: Slide bi-culture was done to determine the interactions between *C. cellulolyticum* and *A. flavus* under photomicroscope. Figure 1 shows some features of *Chaetomium cellulolyticum* and *Aspergillus flavus* photo microscopy. It shows the spores of *C. cellulolyticum*, normal hypha of *A. flavus*, damaged hyphae of *A. flavus*. It was observed that the hyphae of *A. flavus* were damaged when grown together with *C. cellulolyticum*. Based on the results, *C. cellulolyticum* may produce substances that could damage the hyphae of *A. flavus*. Similar observation was reported by Pornsuriya [14] about the reaction of some strains of *Chaetomium* like *C. cochloides* against some pathogens. It was reported that the pathogen showed abnormal features of hyphae and demonstrated uncommon shapes.

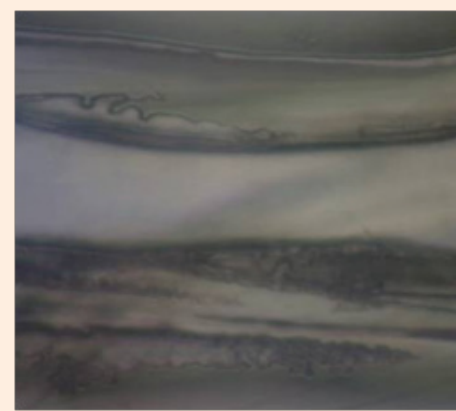
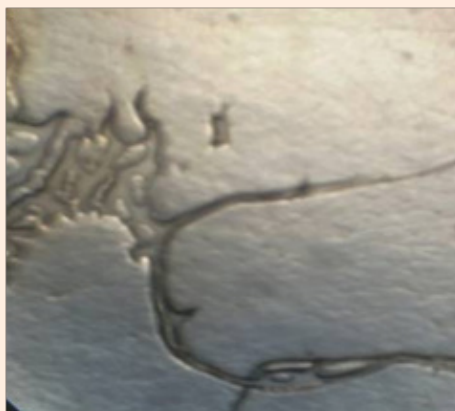


C. cellulolyticum
Spores



normal hyphae
of *A. flavus*

Figure 1a: Features of *C. cellulolyticum* and *A. flavus* under photomicroscope (HPO).



Damaged hyphae *A. Flavus*

Figure 1b: Damaged hyphae of *A. flavus* (A and B) under photomicroscope (HPO).

Co-culture method: The *C. cellulolyticum* was tested to determine if this fungus could colonize the agar lawn of *A. flavus* using co-culture method as shown in Figure 2. The area and the percentage colonization of *C. cellulolyticum* and the pathogen were recorded at 30 days as shown in Figure 3. The results indicated that *A. flavus* ramified faster than *C. cellulolyticum* as shown in Figure 3. *A. flavus* colonized by 4,234.33 mm² or 52.28% the agar lawn of *C. cellulolyticum* at 30 days and first

crossed the midline. It is clearly observed that there was no colonization of *C. cellulolyticum* on agar lawn of *A. flavus*. On the other hand, the *C. cellulolyticum* colonized by 2,566 mm² or 31.68%. Results showed that, *C. cellulolyticum* was grown slowly, similarly as what Soyong [15] said that *Chaetomium* spp. is a slow growing fungus compared to pathogens, and the growth and development is at 25 to 30 days.



Figure 2: The co-culture of *C. cellulolyticum* (A) and *A. flavus* (B) per replicates at 30 days.

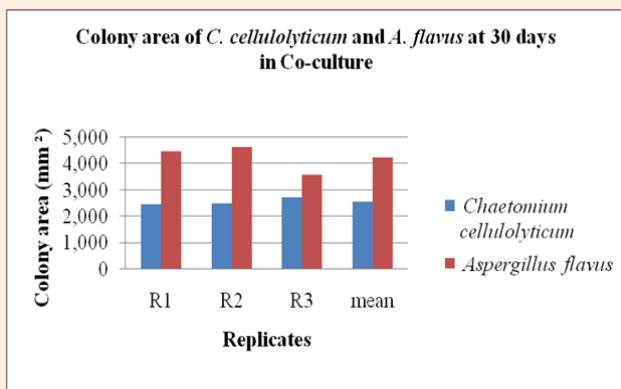


Figure 3: The colony area of *C. cellulolyticum* and *A. flavus* per replicates at 30 days in co-culture.

Antimicrobial bioassay: The mycelia of *C. cellulolyticum* were extracted using Hexane and Ethanol extractions. This method was done to determine the presence of inhibitory effect of *C. cellulolyticum* extracts on the mycelial growth of *A. flavus*. Table 1 shows the mean diameter of zone of inhibition of all the treatments. Results revealed that at 24th to 48th hours of incubation see Figures 4 & 5, mycelial crude hexane and ethanol extracts do not produced zone of inhibition on *A. flavus*. Statistical analysis showed that the zone of inhibition of the two crude extracts were comparable to the positive and negative control. However, at 72nd to 96th hours, the extracts were significantly smaller to the positive control and comparable to the negative control see Appendix I & J. The affectivity of the mycelial crude extracts was not observed at 24th to 96th hours may because of the little amount of bioactive compounds found in the crude extracts. The active components capability may also have short time effect to control the pathogen or the crude extracts were diffused across time in contrast to the report of Soyong et al. [15] that substances extracted from *Chaetomium* spp. could exhibit activity towards some pathogens.

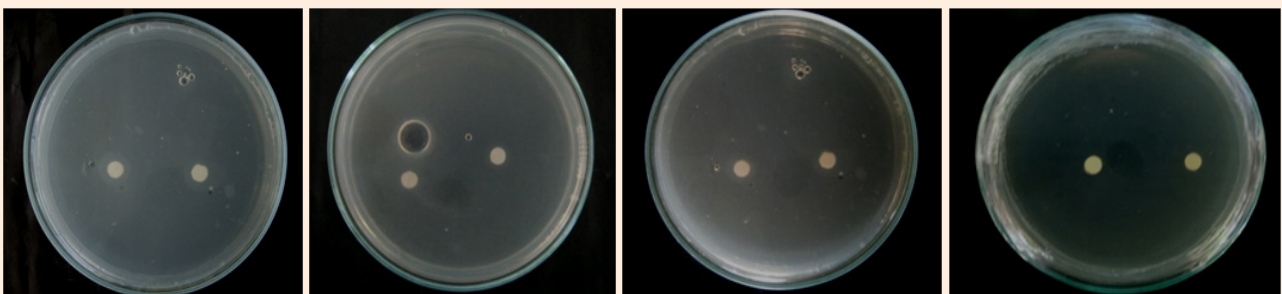


Figure 4: Zone of Inhibition exhibited by all the treatments against *A. flavus* at 24th hours of incubation. (T1- crude hexane extract, T2- crude ethanol extract, T3- positive control (Dithane), T4- negative control (DMSO)).

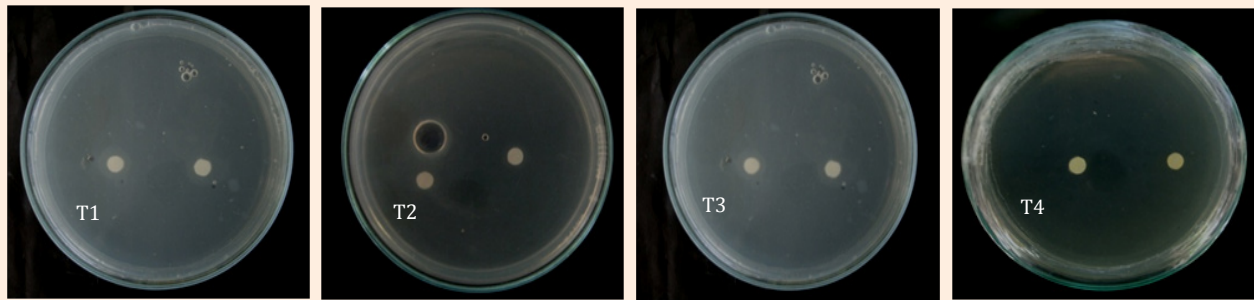


Figure 5: Zone of Inhibition exhibited by all the treatments against *A. flavus* at 48th hours of incubation. (T1- crude hexane extract, T2- crude ethanol extract, T3- positive control (Dithane), T4- negative control (DMSO)).

Interactions of *Chaetomium cellulolyticum* and *Curvularia lunata*

The interactions of *C. cellulolyticum* and *C. lunata* were also studied using the slide bi-culture, co-culture techniques and antimicrobial bioassay. Description of the fungal microscopic association was done in slide bi-culture while the zone of colonization was studied in the co-culture method and zone of inhibition was determined in the antifungal bioassay.

Slide bi-culture: This slide bi-culture was done to determine the interactions between *C. cellulolyticum* and *C. lunata* under photomicroscope. Figure 6 shows some features of *C. lunata* under photomicroscope. It shows the normal hyphae and conidia of *C. lunata* and damaged hyphae and conidia of *C. lunata*. Results revealed that the hyphae and conidia of *C. lunata* were broken when grown together with *C. cellulolyticum*. Based on the results, *C. cellulolyticum* may produce inhibitory substances that could break the hyphae and conidia of *C. lunata*. It was reported by Pornsuriya [14] that the pathogen showed abnormal features of hyphae and demonstrated uncommon shapes when grown together with some strains of *Chaetomium*.



Figure 6: Some features of *C. lunata* under photomicroscope (HPO). Normal hypha (A), conidia (B), broken hyphae (C), broken conidia (D).

Co-culture method: The *C. cellulolyticum* was tested to determine if this fungus could colonize the agar lawn of *C. lunata* using co-culture method as shown in Figure 7. The colonization of *C. cellulolyticum* was observed on the agar lawn of *C. lunata* by 0.15% at 30 days as shown in Figure 8, but the pathogen ramified faster at 10 days of incubation. Results revealed that the colonization of *C. cellulolyticum* and *C. lunata* in co-culture

method were 2,867.33 mm² or 35.40% and 3,115.33 mm² or 38.46% respectively as shown in Figure 9. Based on the results, *C. cellulolyticum* was grown slowly than the pathogen. In a personal communication with Soyong [16] at KMITL, Ladkrabang, Thailand, proved that *Chaetomium* spp. is a slow growing fungus compared to pathogens, and the growth and development is at 25 to 30 days.

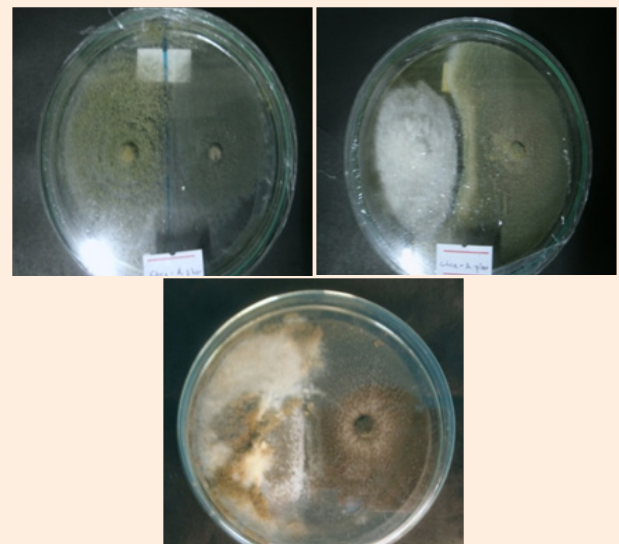


Figure 7: The co-culture of *C. cellulolyticum* (A) and *C. lunata* (B) per replicates at 30 days.

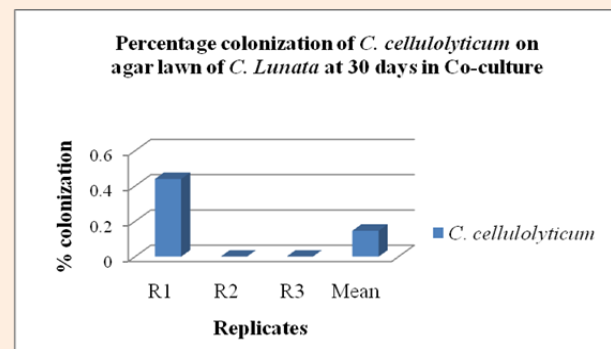


Figure 8: Percentage colonization of *C. cellulolyticum* on agar lawn of *C. Lunata* at 30 days in Co-culture.

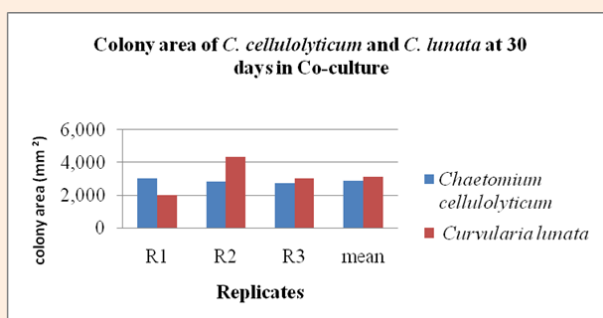


Figure 9: Percentage colonization of *C. cellulolyticum* and *C. lunata* at 30 days in Co-culture.

Antimicrobial bioassay: This method was done to determine the inhibitory effect of *C. cellulolyticum* on the mycelial growth of *C. lunata*. Table 2 shows the mean diameter of zone of inhibition of all the treatments. Results revealed that mycelial crude hexane and ethanol extracts do not produced zones of inhibition on *C. lunata* at 24th to 96th hours of incubation. Analysis of variance and comparison among means showed, that inhibitions of two extracts were comparable to the positive and negative control at 24th hours of incubation. However, at 48th to 96th hours of incubation see (Appendix K-M) the crude extracts were significantly smaller to the positive control but comparable to the negative control. The inhibitory effect of the mycelial crude extracts was not observed at 24th to 96th hours of incubation may because of the small quantity of bioactive compounds in the crude extracts, so the capability to eradicate the pathogen is not possible across time or the pathogen may have already became tolerant or diffused as time of incubation increases. In contrast in study reported by Soyong et al. [17] that extracts from *Chaetomium cupreum* and *Chaetomium globosum* are able

to suppress plant pathogens such as *Curvularia lunata* *in-vitro*.

Summary

This study was conducted to test the antagonistic activity of *Chaetomium cellulolyticum* against *Aspergillus flavus* and *Curvularia lunata* using different methods *in-vitro*. Using Slide bi-culture test, results revealed that the hyphae of *A. flavus* were deformed and damaged when grows with *C. cellulolyticum*. Using co-culture method, the results indicated that *A. flavus* ramified faster than *C. cellulolyticum* by 4,234.33 mm² or 52.28% at 30 days. In bioassay, results revealed that at 24th to 48th hours of incubation, mycelial crude hexane and ethanol extracts do not produced zone of inhibition on *A. flavus*. Statistical analysis showed that the zone of inhibition of the two crude extracts were comparable to the positive and negative control. However, at 72nd to 96th hours, the extracts were significantly smaller to the positive control and comparable to the negative control.

On the other hand, the hyphae and conidia of *C. lunata* were broken as revealed in the Slide bi-culture test. Based on the results of Co-culture method, it was observed and measured that the colonization of *C. cellulolyticum* on the agar lawn of *C. lunata* is 0.15%. The colonization of *C. cellulolyticum* and *C. lunata* in co-culture method are 2,867.33 mm² or 35.40% and 3,115.33 mm² or 38.46% respectively. In Bioassay, Results revealed that mycelial crude hexane and ethanol extracts do not produced zones of inhibition on *C. lunata* at 24th to 96th hours of incubation. Analysis of variance and comparison among means showed, that inhibitions of two extracts were comparable to the positive and negative control at 24th hours of incubation. However, at 48th to 96th hours of incubation, the crude extracts were significantly smaller to the positive control but comparable to the negative control (Figure 10).

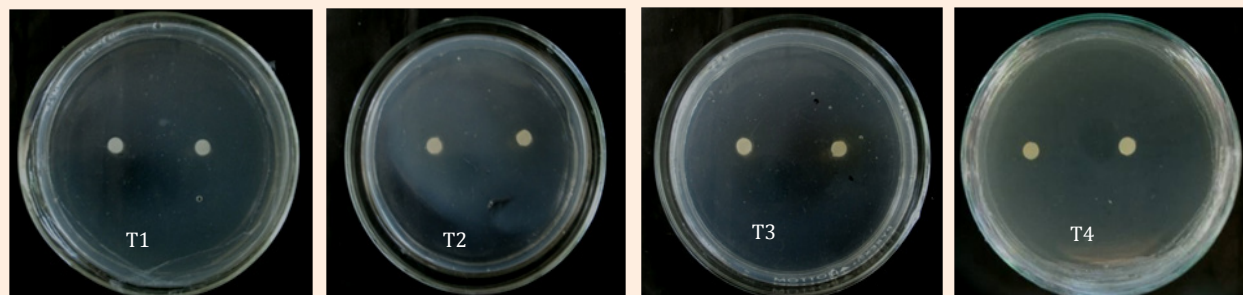


Figure 10: Zone of Inhibition exhibited by all the treatments against *C. lunata* at 24th hours of incubation. (T1- crude hexane extract, T2- crude ethanol extract, T3- positive control (Dithane), T4- negative control (DMSO).

Conclusion

Chaetomium cellulolyticum have antibiosis mechanism to deform or damage the morphological hyphae and either conidia of *A. flavus* and *C. lunata*. However, the affectivity of crude hexane and ethanol extracts from *C. cellulolyticum* were not observed to inhibit the growth of *A. flavus* and *C. lunata*.

Recommendation

Chaetomium cellulolyticum have antibiosis mechanism against the pathogens. The product of this study can contribute to conventional and organic farming. However, the following are

suggested for further study:

More mycelia of *C. cellulolyticum* should be extracted and other solvents should be used to test and verify the inhibitory activity. Use other Bioactivity Assay of crude extracts of *C. cellulolyticum* like the method used by Sibounnavong et al. [2]. Measurement of zone of inhibition and determination of spore inhibition should be done in co-culture method. Test the different age interval of the *C. cellulolyticum* and the pathogens in the co-culture method. Use volatile and non-volatile antibiosis method. Determination of bioactive constituents present in *C. cellulolyticum*. DNA analysis of *C. cellulolyticum* should be done. *In-vivo* biological activities of *C. cellulolyticum* should be done

using different tests like, disease suppression, pathogenicity and toxicology tests.

References

1. Environmental Protection Agency (2004) Pesticides: Regulating Pesticides. Environmental Protection Agency, USA.
2. Sibounnavong P, Cynthia DC, Kanokmedhakul S, Soyong K (2008) The new antagonist fungus, *Emericella nidulans* strain EN against Fusarium Wilt of Tomato. *Journal of Agricultural Technology* 4(1): 89-99.
3. Weber R, Kita W, Runowska HB (2001) Effects of Tillage Methods on The Occurrence of Culm Base Disease In Several Winter Wheat Cultivars. *EJPAU* 4(2): 06.
4. Ali MK, Abdul ML, Saleem S (2002) Pathogenicity of *Lasiodiplodia theobromae* and *Fusarium solani* On Mango. *Pak J Bot* 36(1): 181-189.
5. Shu-Qin X, Xue C, Ying S (2006) Nature of Physical and Chemical and Pathogenic Function of Toxin of *Curvularia lunata*(Wakker) Boed. *Journal of Maize Sciences*.
6. Alwala S (2007) Identification of Molecular Markers Associated with Resistance to *Aspergillus flavus* in Maize.
7. Smith CA, Robertson D, Yates B, Nielsen DM, Brown D, Dean RA (2008) The effect of temperature on Natural Antisense Transcript (NAT) expression in *Aspergillus flavus*. *Curr Genet* 54(5): 241-269.
8. Tsukiboshi A (2001) Diseases of Forage Crops.
9. Zhang H, Yang Q (2007) Expressed sequence tags-based identification of genes in the biocontrol agent *Chaetomium cupreum*. *Appl Microbiol Biotechnol* 74(3): 650-658.
10. Charoenporn C, Kanokmedhakul S, Lin F, Poeaim S, Soyong K (2010) Evaluation of bio-agent formulations to control Fusarium wilt of tomato. *African Journal of Biotechnology* 9(36): 5836-5844.
11. Aggarwal R, Sharma V, Kharbikar L, Renu (2008) Molecular characterization of *Chaetomium* species using URP-PCR. *Genetics and Molecular Biology* 31(4): 943-946.
12. Soyong K (1992) Antagonism of *Chaetomium cupreum* to *Pyricularia oryzae*. *Journal of Plant Protection in the Tropics* 9(1): 17-23.
13. Kanokmedhakul S, Kanokmedhakul K, Nasomjai P, Louangsysouphanh S, Soyong K (2006) Antifungal Azaphilones from the fungus *Chaetomium cupreum* CC3003. *J Nat Prod* 69(6): 891-895.
14. Pornsuriya C, Lin C, Kanokmedhakul S, Soyong K (2010) Efficacy of antifungal metabolites from some antagonistic fungi against *Pythium aphanidermatum*. *Journal of Agricultural Technology* 6(2): 299-308.
15. Soyong K, Srinon W, Rattanacherdchai K, Kanokmedhakul S, Kanokmedhakul K (2005) Application of antagonistic fungi to control anthracnose disease of grape. *Journal of Agricultural Biotechnology* 1: 33-41.
16. Soyong K (2011) Personal communication.
17. Soyong K, Kanokmedhakul S, Kukongviriyapa V, Isobe M (2001) Application of *Chaetomium* species (Ketomium®) as a new broad spectrum biological fungicide for plant disease control: A review article. *Fungal Diversity* 7: 1-15.