

Review Article





Non-alcoholic fatty liver disease: bases for therapeutic roles of vitamin e and n-3 PUFA

Abstract

Non-alcoholic fatty liver disease (NAFLD) is the first cause of liver disease worldwide, leading to non-alcoholic steatohepatitis (NASH), cirrhosis and finally death. The available evidence suggests that oxidative stress plays a key role in the pathophysiology of NAFLD, participating in mitochondrial increased lipid peroxidation involved in mitochondrial dysfunction. Polyunsaturated fatty acids (PUFAs) are present in the cell and mitochondrial membrane, and can act through several molecular pathways such as the peroxisome proliferator-activated receptor-α (PPAR-α) activation and sterol regulatory element-binding protein 1c (SREBP-1c) regulation to finally promote an antioxidant status within cell physiology. Vitamin E has antioxidant and anti-inflammatory properties that have been proved to reduce liver steatosis and inflammation, acting through free radical trapping and preventing docosahexaenoic acid (DHA) molecules damage through a membrane stabilization action. These two molecules could thereby be a potential target for promising therapies within the multiple pathways of this disease.

Keywords: NAFLD, oxidative stress, n-3 PUFA, vitamin e, diabetes mellitus

Volume 3 Issue 7 - 2017

Pablo Lizana, Melissa Galdames, Guillermo Valenzuela, Cristóbal Bourgeois and Ramón

Faculty of Medicine, University of Chile, Chile

Correspondence: Ramón Rodrigo, Molecular and Clinical Pharmacology Program, Institute of Biomedical Sciences, Faculty of Medicine, University of Chile, Independencia 1027, CP 8380453, Santiago, Chile, Tel +56 2 2 2978 6126, Email rrodrigo@med.uchile.cl

Received: October 05, 2017 | Published: December 06, 2017

Abbreviations: ALT, alanine transaminase; AST, aspartate amino transferase; CIMT, carotid intima media thickness; CYP450, cytochrome P450; DHA, docosahexaenoic acid; DM, diabetes mellitus; EPA, eicosapentaenoic acid; FA, fatty acids; HF, high fat; HNE, 4-hydroxy-2-nonenal; MDA, malondialdehyde; mtDNA, mitochondrial dna; n-3 PUFAs, n-3 polyunsaturated fatty acids; NAS, nafld activity score; NASH, non-alcoholic steatohepatitis; NAFL, non-alcoholic fatty liver; NAFLD, non-alcoholic fatty liver disease; Nrf2, NF-F2-related factor 2; NOS, nitric oxide synthase; NOX, NADPH oxidase; PPAR-α, peroxisome proliferator-activated receptor A; PPARs, peroxisome proliferator-activated receptors; PUFAs, polyunsaturated fatty acids; RNS, reactive nitrogen species; ROS, reactive oxygen species; SREBP-1c, sterol regulatory elementbinding protein 1c; TG, triglycerides; US, ultrasound; XO, xanthine oxidase.

Introduction

In the last years the amount of people affected with non-alcoholic fatty liver disease (NAFLD) has increased: reaching today the first cause of liver disease worldwide. The prevalence of NAFLD in the United States is between 10% and 30% with similar rates reported from Europe and Asia.1 It should be considered that the development of cirrhosis in non alcoholic patients has been recognized as an important and frequent cause of aminotransferases elevation. 1-3

NAFLD etiopathogenesis has not been completely elucidated and should be understood as a multifactorial process that includes changes in metabolic homeostasis: inflammation: insulin resistance: fibrosis and oxidative stress.4 Moreover it is known that patients who have metabolic syndrome, obesity, insulin resistance, diabetes mellitus (DM) or hyperlipidemia have an increased risk of developing non-alcoholic fatty liver (NAFL): probably because those diseases have similar multifactorial causes. In particular oxidative stress has an important role in the progression from NAFL to non-alcoholic steatohepatitis (NASH) and finally to cirrhosis and hepatocellular carcinoma (HCC).5

Oxidative stress is the result of an imbalance that favours the increase of oxidative species: which are normally managed by antioxidant mechanisms produced inside the hepatocyte. Specifically: the loss of this balance allows to the formation and increase of reactive oxygen species (ROS) and reactive nitrogen species (RNS). Both species target essential biomolecules within the hepatocytes: producing injury in cellular structure and compromising many biological functions: being this a critical event that could finally lead to hepatotoxicity. 1,6,7

Changes in lifestyle and daily diet are the main treatment in NAFLD.8 Latest evidence has shown possible beneficial effects of vitamin E and n-3 polyunsaturated fatty acids (n-3 PUFAs) in NALFD patients. Vitamin E and n-3 PUFAs are essential nutrients that have essential functions in human physiology and both are distinguished for their antioxidant properties.

The purpose of this review is to present a summary about the pathogenesis of NAFLD and to analyse the vitamin E and n-3 PUFAs role in the current treatment of this disease.

NAFLD and cardiovascular risk

One of the most important causes of death in patients with NAFLD: in particular: patients with non-alcoholic steatohepatitis: are cardiovascular diseases.9 Cohort's studies have shown ischemic heart diseases represent a 25% of the total of death in patients with NAFLD: being this the second cause of mortality. 10

Recent studies show a relationship between NAFLD and atherosclerosis. Cardiovascular mortality in NAFLD is associated with the severity of liver disease: representing the most part of NAFLD mortality.11 NAFLD may contribute to cardiovascular diseases development: by an increase of insulin resistance and atherogenic dyslipidemia, which both are proven cardiovascular risk factors. 12,13 An increased release of chemical messengers from the visceral adipose tissue: such as inflammatory cytokines: leads to inflammation of adipose tissue. This process intensifies insulin resistance: and



consequently participates in the development of atherosclerosis and cardiovascular diseases. ^{13,14} Despite, NAFLD may be an independent factor for adipose tissue inflammation and posterior cardiovascular risk increasing as adipose tissue inflammation has been observed in individuals with NAFLD independent of the obesity grade. ¹⁵

NAFLD influences on macrophage polarization

Macrophages are part of the innate immunity and play a key role in inflammation and host defense. ¹⁶ Depending on the environment stimuli these cells may undergo to different phenotypes. ¹⁷ Classical M1 activation leads to the release of pro-inflammatory cytokines: whereas M2 activation promoted the tissue remodelling and causes a regulation on immune-regulatory functions. ^{18,19} Macrophages plasticity allowed the switch from M1 to M2 and vice versa. ²⁰ For example: adipose tissues in obesity promoted a polarization shift toward M1. ²¹

Oxidative stress and NAFLD

Oxidative stress is described as a mechanism of injury in different disease processes. The term refers to a condition in which there is an imbalance between the production of ROS and RNS and the antioxidant defense system within a particular biological system.^{22,23} Mitochondrion is the principal source of ROS production. Mitochondria consume 90% of cell's oxygen to produce ATP through oxidative phosphorylation. However about 2% of the oxygen used in cell metabolism is converted to ROS mainly through superoxide anion (O₂) production.^{24,25} ROS can also be produced outside of the mitochondria in lesser significative amounts. These processes involve non enzymatic and enzymatic reactions. In the case of NASH extra-mitochondrial ROS are generated principally through NADPH oxidase xanthine oxidase D-amino oxidase p-450 cytochromes proline and lysine hydroxylase and uncoupled nitric oxygen synthase. It has been described a direct association between NASH and cytochrome P450 2E1 (CYP2E1) isoform pro-oxidant activity.²⁶ This microsomal enzyme promotes free fatty acids B-oxidation a potential mechanism of ROS production.²⁷ In addition ROS may be generated through NADPH oxidase in Kuppfer cells.²⁸

Mitochondrial ROS production contributes to diverse liver diseases through an accumulation of mitochondrial DNA (mtDNA) mutations leading to dysfunction caused by several reactions involved in oxidative phosphorylation processes and final lethal cell injury.²⁹ There is some evidence that mtDNA mutations may affect NAFLD development as suggested by studies reporting reduced levels of mtDNA in patients with NASH.30 Reports demonstrate that mtDNA increased levels are found in patients with fatty liver but with nor-inflammation/ fibrosis. 31 Paradies has hypothesized that the accumulation of mtDNA mutations may lead to an impairment of oxidative phosphorylation reactions and mitochondrial respiratory chain dysfunction resulting in increased ROS production prior to accumulation of mtDNA mutations finally triggering a vicious cycle of oxidative damage in which ROS production promotes further mitochondrial dysfunction and oxidative damage.32 This suggests that a link may exist between mitochondrial mtDNA mutations and NAFLD etiology and pathophysiological mechanisms. Moreover damage induced to mtDNA can be transferred through mitochondria and cell division.³³

PUFAs are an important component of mitochondrial phospholipids. In the literature lipid peroxidation has been related as a consequence of oxidative stress. Mitochondria present a high concentration of

PUFAs being subtracts for oxidizing reactions and generating lipid peroxidation products such as hydroperoxides and endoperoxides. Subsequently these products may undergo fragmentation resulting in the formation of aldehyde by-products such as 4-hydroxy-2-nonenal and malondialdehyde. 34-36 The importance of these molecules formed only by peroxidation of PUFAs 15 lies in the potential to migrate to distant intracellular and extracellular targets thus amplifying oxidative stress effects (Figure 1). 35-37

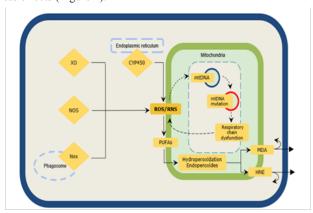


Figure I Relationship between oxidative stress and NAFLD.

The imbalance produced in the antioxidant/oxidant ratio within the hepatocyte leads to oxidative stress. ROS and RNS are principally enhanced by mitochondrial processes but, besides, a considerable amount is produced by other extra mitochondrial systems such as XO, NOS, NOX, and CYP450. Inside the mitochondria, the increased quantity of ROS and RNS produces an accumulation of mutated mtDNA, which leads to mitochondrial dysfunction and cell damage. This last event produces more ROS and RNS, which finally leads to a vicious cycle with more mutated mtDNA production, and so on. MDA and HNE are products from the peroxidation of PUFAs, and can amplify oxidative stress both intracellular and extracellular.

XO, xanthine oxidase; NOS, nitric oxide synthase; NOX, NADPH oxidase; CYP450, cytochrome P450; ROS, reactive oxygen species; RNS, reactive nitrogen species; PUFAs, polyunsaturated fatty acids; mtDNA, mitochondrial DNA; MDA, malondialdehyde; HNE, 4-hydroxy-2-nonenal.

Therapeutic approaches into NAFLD n-3 PUFAS and vitamin e

n-3 polyunsaturated fatty acids

n-3 PUFAs generalities: Polyunsaturated fatty acids (PUFAs) are safe and efficacious compounds³⁸ participating in numerous processes having a wide range of effects on biochemical and physiologic functions. PUFAs are key compounds of triglyceride and phospholipid membranes of cells and subcellular organelle membranes.³⁹ Two main groups of PUFAs can be identified n-3 PUFAs or omega 3 fatty acids and n-6 PUFAs also known as omega 6 polyunsaturated fatty acids (n-6 PUFAs). Both classes of PUFAs are essential nutrients since human body cannot synthesize them⁴⁰ n-6 PUFAs are usually obtained from corn products and soybean oil while n-3 PUFAs are part of fish oil canola walnuts among others. 41 The two fatty acids experience similar metabolic process including elongation and de-saturation processes made by enzymes localized in the endoplasmic reticulum and the mitochondria.42,43 Metabolized n-6 PUFAs lead to arachidonic acid while metabolized a-Linolenic acid the most abundant n-3 PUFAs in the western diet lead to eicosapentaenoic acid (EPA) which are further metabolized to DHA.40 Evidence suggests a pro-inflammatory role of n-6 PUFAs and an anti-inflammatory profile of n-3 PUFAs through their metabolized species.44

PUFAs and cardiovascular risk: Multiple evidence has reported a supportive effect of n-3 PUFA in cardiovascular disease principally decreasing the risk of cardiac mortality.⁴⁵ Beneficial effects of n-3 PUFA ingest to reduce cardiovascular risk are decrease in triglycerides levels reduce heart rate and blood pressure 46,47 improve in endothelial and autonomic function^{48–50} and anti-inflammatory effects. ⁴⁵ In addition cross-sectional studies have been related n-3 PUFA consumption with lower levels of atherosclerosis. 51,52 Recently pre-specified sub-study of the WELCOME study⁵³ has reported a beneficial effect in carotid intima media thickness (CIMT) progression when NAFLD severity was decreased. CIMT is a validated tool for the prediction of stroke or myocardial infarction.⁵⁴ Previously a meta-analysis described an important association between CIMT and patients with NAFLD this association might be responsible of an increase 13% of CIMT.55 However in the pre-specified sub-study of the WELCOME trial⁵³ n-3 PUFA supplementation did not improve CIMT progression. Despite of this negative result it is necessary to perform more studies using n-3 PUFA as main strategy for NAFLD treatment with the objective to deepen the analysis of cardiovascular outcomes.

Effects on macrophage polarization: PPAR-y modulate the immune inflammatory response due to its anti-inflammatory properties⁵⁶ generating a negative interference of multiples genes including nuclear factor kappa B (NF-κ B).57 PPAR-γ is a regulator of macrophage M2 polarization.²⁰ Moreover it participated in acquisition and maintenance of M2 phenotype in adipose tissue⁵⁸ and its disruption in mice impaired M2 macrophage activation leading to a susceptible to obesity and insulin resistance.⁵⁹ Kupffer cells are the resident macrophages of the liver. Evidence has showed that Kupffer cell contributes to the pathogenesis of NAFLD. Studies have investigated the effect of DHA on Kupffer cells/macrophages polarization in vitro in a NAFLD model. High fat (HF) diet-induced hepatic steatosis and local proinflammatory response was closely associated with M1-predominant polarization of Kupffer cells.²⁰ On the other hand PPAR-y results to have the potential to balance lipid-induced M1/M2 macrophages/ Kupffer cells polarization preventing the development of NAFLD in these HF diet mice. n-3 PUFA administration favours a switch of Kupffer cells/macrophages to an M2 phenotype.20

n-3 PUFAs antioxidant properties: Through its metabolism to structurally related prostaglandin and leukotrienes n-3 PUFAs derived species act as ligands that stimulate transcriptions genes. For example it joins nuclear receptor proteins such as peroxisome proliferatoractivated receptors (PPARs) to defend against ROS production and PPARα excess.⁶⁰⁻⁶² Thus n-3 PUFAs produce an up regulation in both fat oxidation genes and antioxidant genes generating and imbalance that benefits antioxidants molecules.⁶⁰ Within the properties of n-3 PUFAs evidence suggests a modulation of redox signaling pathways.⁶³ For example oxidized omega 3 react with keap 1 thereby inducing NF-F2-related factor 2(Nrf2) participating in the expression of genes that encode proteins responsible to regulate detoxification of ROS such as heme-oxygenase-1.⁶¹

Relationship between n-3 PUFAs and NAFLD: Some patients with NAFLD have concomitantly diminished levels of n-3 PUFA. It has been observed that NAFLD condition associated with a decline in hepatic n-3 PUFAs is more severe than with decline in n-6 PUFAs. 64,65 High blood and hepatic $\omega 6/\omega 3$ PUFAs ratio is associated with inflammation and NAFLD progression. 38 DHA would be more effective than EPA at attenuating western diet-induced hepatic fat inflammation and fibrosis and controlling multiple liver lipid metabolism signaling pathways which take control over liver lipid metabolism inflammation and fibrosis. 38,65

It was reported that dietary supplementation with fatty acids can improve NAFLD associated with hyperlipidemia by modifying the function of platelets and leukocytes. ^{66,67} Also n-3 PUFAs especially DHA have demonstrated to reduce triglycerides (TG) accumulation and improve hepatic steatosis being important regulators of hepatic gene transcription. ⁶⁸ In fact PUFAs might prevent NAFLD by the activation of PPARs and the inhibition of sterol regulatory element-binding protein-1c gene (SREBP-1c) by down regulating over expressed glycolytic and lipogenic genes. ⁶⁹

As mentioned above n-3 PUFA ingestion leads to PPAR α regulation and activation. Mice lacking PPAR α have elevated free fatty acid levels and fatty livers consequences of their inability to combust fatty acids. ⁶² Humans studies support a role of SREBP-1c in the pathogenesis of steatosis. ^{70,71} Increased SREBP-1c leads to mixed insulin resistance and sensitivity in livers of lipodystrophic and ob/ob mice. ⁷² Shimomura et al. ⁷³ described that combination of insulin resistance and insulin sensitivity establishes a vicious cycle that aggravates hyperinsulinemia and insulin resistance in lipodystrophic and ob/ob mice. ⁷³

Clinical trials in NAFLD with n-3 PUFAs supplementation: Evidence has shown different results in the use of n-3 PUFAs in patients with NAFLD.

- **A.** What has not worked: A double blind randomized placebo controlled multicenter clinical trial was performed by Dasarathy et al. ⁷⁴ in NASH patients with DM. Participants were randomized into two study groups and received either an oral dose of 2160mg of EPA and 1440mg of DHA in divided two pills or placebo the study has a duration of 48weeks. Primary endpoint was histology changes. Research showed that on adults there were no differences between n-3 PUFAs group and placebo in the mean serum transaminases and histological evaluations. ^{74,75}
- B. What has worked: Different reviews have summarized clinical trials performed in NAFLD patients such as one performed by Jump et al.³⁸
 - I. Analyses assess the clinical trials impacts of n-3 PUFAs supplementation on children with NAFLD. Disparate doses of DHA or DHA+EPA were used with treatment duration ranging from 3 to 24 months. Nobili et al. 76 performed studies that showed a decrease in plasma alanine transaminase (ALT) on ultrasound (US) hepatosteatosis76-78 NAFLD activity score (NAS)79 and aspartate aminotransferase (AST) HOMA-IR fasting insulin and plasma TG. 78 Janczyk et al. conducted a randomized control trial on 64 subjects treatment group received DHA+EPA in a 32 proportion weight adjusted dose treatment with n-3 PUFAs improved AST and gammaglutamyl transpeptidase levels in children with NAFLD compared with placebo no difference was observed at US.80 Some studies such as one performed by Pacifico et al.81 also informed a diminished-on body weight and waist circumference.81
 - II. On adults results are similar of those seen in children. Moreover it has been demonstrated a decrease in hepatic steatosis joined to a diminished on hepatic fibrosis in studies performed in the past five years. 82-84 Hepatic enzymes such as ALT and AST decrease in the treated group compared to placebo one. 84-86 A diminish on plasma TG was also observed. 82,84-86

C. What is new: Based on the hypothesis that supplementation with DHA and vitamin D would benefit the whole spectrum of NAFLD. Della Corte et al. 87 evaluated in a randomized double-blind placebo-controlled trial the effect of daily DHA (500mg) plus vitamin D (800IU) in 41 obese children with biopsy-proven NAFLD and vitamin D deficiency after 12months of use. Results demonstrated that DHA plus vitamin D treatment improved insulin-resistance lipid profile ALT and NAFLD Activity Score.

Hodson et al.88 conducted a randomized control trial double-blind and placebo-controlled study. This was a pre-specified sub-study of the WELCOME trial (Wessex Evaluation of fatty Liver and Cardiovascular markers in NAFLD with OMacor therapy). They measured whether the treatment with 3 PUFA in patients was associated with changes in hepatic fatty acids (FA) synthesis post prandial FA partitioning and hepatic and peripheral insulin sensitivity. Twenty-four subjects were allocated to two groups group A 12 received EPA+DHA 4g/day for 15-18months and group B received placebo during the same time. Results showed that patients who have an increase in erythrocyte DHA enrichment of >2% (a surrogate marker of changes in liver enrichment) have significantly lower concentrations of plasma TG and very-low density lipoproteins pre- and post-prandial (P<0.001) a reduction on de-novo lipogenesis and favourable changes in both hepatic insulin sensitivity and hepatic FA metabolism. No changes were observed in whole-body insulin sensitivity or peripheral glucose disposal.

Vitamin E

Vitamin E sources: Vitamin E is an essential micronutrient for humans and achieving an optimal status is assumed to produce beneficial health outcomes. SP There are eight lipophilic forms of vitamin E naturally occurring which include four tocopherols ($\alpha T \beta T \gamma T \delta T$) and four tocotrienols ($\alpha T \beta T \epsilon \gamma T \epsilon \delta T \epsilon \epsilon$

Effects on cardiovascular risk: Vitamin E cardiovascular benefits have been suggested as it inhibits the oxidation of low-density lipoprotein cholesterol in plasma. So Clinical trials have not provided convincing evidence of a vitamin E protective effect for cardiovascular risk in general population. However a meta-analysis shown vitamin E treatment significantly reduced the risk of myocardial infarction and death by 40% and 50% respectively in a diabetic homozygous for haptoglobin allele population. Diabetes mellitus has been associated with a low concentration of antioxidants in particular vitamin E concentration. Recent study suggested a relationship between both vitamin E deficiency and oxidative status with prediabetes in a sample of apparently healthy individuals. These outcomes suggest vitamin E may be effective to reduce cardiovascular risk in patients exposed to oxidative stress related damage such as the case of diabetes and NAFLD.

Vitamin E antioxidant properties: All tocopherols and tocotrienols are potent antioxidants assuming its role by scavenging lipid peroxyl which can produce severe damage on the cell membranes via lipid peroxidation. So Some anti-inflammatory properties have been seen with the vitamin E γ T natural form such as inhibiting pro-inflammatory eicosanoids and being able to trap electrophiles including reactive nitrogen species which could diminish the pro-oxidant environment enhanced during inflammation. So,100,101

Relationship between vitamin E and NAFLD: As seen before oxidative stress plays a key role in NAFLD pathogenesis and in fact patients with this disease have an enhanced oxidative stress and an antioxidant deficiency which may lead to increased lipid peroxidation and cell death due to mitochondrial impairment. Even though there are no standard protocols for the treatment of NAFLD a higher intake of vitamin E is thought to counteract the increased oxidative stress found in this kind of patients and it is commonly prescribed as a supplement in the clinical practise line pharmacological treatment in the management of NASH especially when diet and other lifestyle changes are insufficient.

Clinical trials in NAFLD with vitamin E supplementation: Many studies have been run to prove the impact of the use of vitamin E on NAFLD patients. For example there is a cohort study on patients with NAFLD and metabolic syndrome treated with vitamin E for six months showing a reduction on ALT levels compared with the control group. 104 Magosso et al. 105 examined the effects of a one-year treatment of mixed palm tocotrienols on the echogenic response of hyper cholesterolemic adult patients with NAFLD showing hepatoprotective effects. Another clinical trial showed that obese children with NAFLD treated with a six-month lifestyle change plus vitamin E therapy is associated with a significant reduction of oxidative stress represented with decreased levels of prostaglandin F2α and ALT and increased levels of endogenous secretory receptor for advanced glycation end products compared with the lifestyle only group leading to conclude that vitamin E supplementation have potential and positive results on the oxidative profile of this kind of patients. 106

PIVENS trial compared non-diabetic biopsy-proven NASH patients who received vitamin E to patients receiving pioglitazone and placebo showing that vitamin E was better than placebo in reducing ALT levels liver steatosis and inflammation. ¹⁰⁷ Another trial called TONIC performed in children and adolescents recorded similar findings. ¹⁰⁸ Finally a meta-analysis was published by Sawangjit et al. ¹⁰⁹ in which they analysed 44 randomized controlled trials up to November 2015 comparing different interventions for NAFLD that involved a total of 3802 patients. Vitamin E therapies were supported by high quality evidence in resolution of NASH and improvement in NAS characterized by a decrease in steatosis ballooning and lobular inflammation.

Combined n-3 PUFAs and vitamin E therapy

A mixed vitamin E and n-3 PUFAs could be an interesting therapeutic strategy in NAFLD. This hypothesis is based on antioxidant effect of these molecules that was described above. Both nutrients are related vitamin E has an antioxidant effect that consists in avoiding PUFAs oxidation through free radical scavenging by hydrogen donor mechanisms.¹¹⁰ Moreover a recent theory has been postulated to explain that vitamin E is preferably accumulated in membrane places where a higher concentration of DHA coexists and in this place vitamin E would act as a membrane stabilizator protecting DHA molecules from oxidative stress. 111,112 Human scale studies have demonstrated that vitamin E would play an important role in n-3 PUFAs plasma concentration existing a positive association between high n-3 PUFAs levels and vitamin E intake probably because vitamin E has a role in the support of plasma n-3 PUFAs. 113 This has to be taken into account when a combined antioxidant therapy with Vitamin E and n-3 PUFAs is planned. However there are no studies performed on NAFLD animal models likewise these nutrients have not been mixed at the same time in humans. Considering the beneficial results that were obtained in clinical trials with vitamin E or n-3 PUFAs

further studies could investigate a potential synergistic effect from the association of vitamin E and n-3 PUFAs in NAFLD models (Figure 2).

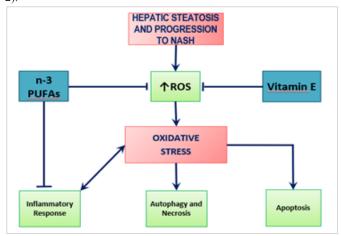


Figure 2 General overview of n-3 PUFAs and vitamin E effects on oxidative stress.

Vitamin E plays its role via lipid peroxide scavenging avoiding the potential damage to the cellular and mitochondrial membranes. N-3 PUFAs act inducing both antioxidant and oxidant genes producing an imbalance that favours the antioxidant mechanisms. Besides: n-3 PUFAs can reduce inflammation: diminishing its signalling pathways.

ROS, reactive oxygen species; n-3 PUFAs,n-3 polyunsaturated fatty acids.

Conclusion

NAFLD remains as an unsolved problem in the clinical practice. Despite the pertinent pathophysiologic mechanisms of the disease have not been fully elucidated oxidative stress is a key factor within the development of the metabolic damage. Nevertheless evidence suggests that both vitamin E and PUFAs could be beneficial individually to reduce the pejorative molecular effects of the disease and improve outcomes in clinical practice through diverse oxidative stress associated pathways. The combination of both therapies seems to be an interesting novel therapeutic strategy to reinforce the antioxidant system and finally ameliorate the lethal cellular consequences of oxidative stress in NAFLD patients. However now a days no studies have been carried with this proposal. We suggest that clinical trial realization is more than relevant to prove the effectiveness of this treatment and finally contribute to reduce and prevent the actual increase in prevalence and global impact of this disease.

Acknowledgements

This study was supported by FONDEF grant ID15I10285.

Conflict of interest

The authors declare that there is no conflict of interest.

References

- Day CP. From Fat to Inflammation. Gastroenterology. 2006;130(1):207–210.
- Angulo P. Nonalcoholic Fatty Liver Disease. N Engl J Med. 2002;346(16):1221–1231.
- Ludwig J, Viggiano TR, McGill DB, et al. Nonalcoholic steatohepatitis: Mayo Clinic experiences with a hitherto unnamed disease. *Mayo Clin Proc.* 1980;55(7):434–438.

- Perazzo H, Dufour JF. The therapeutic landscape of non-alcoholic steatohepatitis. Liver Int. 2017;37(5):634–647.
- Feldstein AE, Werneburg NW, Canbay A, et al. Free fatty acids promote hepatic lipotoxicity by stimulating TNF-alpha expression via a lysosomal pathway. *Hepatology*. 2004;40(1):185–194.
- Videla LA, Rodrigo R, Orellana M, et al. Oxidative stress-related parameters in the liver of non-alcoholic fatty liver disease patients. *Clin* Sci. 2004;106(3):261–268.
- Casoinic F, Sampelean D, Buzoianu AD, et al. Serum Levels of Oxidative Stress Markers in Patients with Type 2 Diabetes Mellitus and Non-alcoholic Steatohepatitis. Rom J Intern Med. 2016;54(4):228–236.
- 8. EASL EASD EASO. Clinical Practice Guidelines for the management of non-alcoholic fatty liver disease. *J Hepatol*. 2016;64(6):1388–1402.
- Rinella ME. Nonalcoholic fatty liver disease: a systematic review. *JAMA*. 2015;13(22):2263–2273.
- Adams LA, Lymp JF, St Sauver J, et al. The natural history of nonalcoholic fatty liver disease: a population-based cohort study. *Gastroenterology*. 2005;129(1):113–121.
- Bhatia LS, Curzen NP, Calder PC, et al. Non-alcoholic fatty liver disease: a new and important cardiovascular risk factor. Eur Heart J. 2012;33(10):1190–1200.
- Fargion S, Porzio M, Fracanzani AL. Nonalcoholic fatty liver disease and vascular disease: State-of-the-art. World J Gastroenterology. 2014;20(7):13306–13324.
- Povero D, Feldstein AE. Novel Molecular Mechanisms in the Development of Non-Alcoholic steatohepatitis. *Diabetes Metab J.* 2016;40(1):1–11.
- Hajer GR, Van Haeften TW, Visseren FL. Adipose tissue dysfunction in obesity, diabetes, and vascular diseases. *Eur Heart J.* 2008;29(24):2959–2971.
- 15. Kolak M, Westerbacka J, Velagapudi VR, et al. Adipose tissue inflammation and increased ceramide content characterize subjects with high liver fat content independent of obesity. *Diabetes*. 2007;56(8):1960–1968.
- Wynn TA, Chawla A, Pollard JW. Macrophage biology in development, homeostasis and disease. *Nature*. 2013;496(7446):445–455.
- Ginhoux F, Schultze JL, Murray PJ, et al. New insights into the multidimensional concept of macrophage ontogeny, activation and function. *Nat Immunol.* 2016;17(1):34–40.
- Biswas SK, Mantovani A. Macrophage plasticity and interaction with lymphocyte subsets: cancer as a paradigm. *Nat Immunol*. 2010;11(10):889–896.
- Sica A, Mantovani A. Macrophage plasticity and polarization: in vivo veritas. J Clin Invest. 2012;122(3):787–795.
- Luo W, Xu Q, Wanf Q, et al. Effect of modulation of PPAR-γ activity on Kupffer cells M1/M2 polarization in the development of non-alcoholic fatty liver disease. Sci Rep. 2017;7:44612.
- 21. Lumeng CN, Bodzin JL, Saltiel AR. Obesity induces a phenotypic switch in adipose tissue macrophage polarization. *J Clin Invest.* 2007;117(1):175–184.
- 22. Mittler R, Vanderauwera S, Suzuki N, et al. ROS signaling: the new wave. *Trends Plant Sci.* 2011;16(6):300–309.
- Dautreaux B, Toledano MB. ROS as signalling molecules: mechanisms that generate specificity in ROS homeostasis. *Nat Rev Mol Cell Biol*. 2007;8(10):813–824.
- Boveris A, Chance B. The mitochondrial generation of hydrogen peroxide. General properties and effect of hyperbaric oxygen. *Biochem J.* 1973;134(3):707–716.

- 25. Kohen R, Nyska A. Oxidation of biological systems: oxidative stress phenomena, antioxidants, redox reactions, and methods for their quantification. Toxicol Pathol. 2002;30(6):620-650.
- 26. Weltman MD, Farrell GC, Hall P, et al. Hepatic cytochrome P450 2E1 is increased in patients with nonalcoholic steatohepatitis. Hepatology. 1998;27(1):128-133.
- 27. Schattenberg JM, Wang Y, Singh R, et al. Hepatocyte CYP2E1 overexpression and steatohepatitis lead to impaired hepatic insulin signaling. J Biol Chem. 2005;280(11):9887–9894.
- 28. Kono H, Rusyn I, Yin M, et al. NADPH oxidase-derived free radicals are key oxidants in alcohol-induced liver disease. J Clin Invest. 2000;106(7):867-872.
- 29. Ricci C, Pastukh V, Leonard J, et al. Mitochondrial DNA damage triggers mitochondrial-superoxide generation and apoptosis. Am J Physiol Cell Physiol. 2008;294(2):C413-C422.
- 30. Sookoian S, Rosselli MS, Gemma C, et al. Epigenetic regulation of insulin resistance in nonalcoholic fatty liver disease: impact of liver methylation of the peroxisome proliferator-activated receptor y coactivator 1a promoter. Hepatology. 2010;52(6):1992-2000.
- 31. Chiappini F, Barrier A, Saffroy R, et al. Exploration of global gene expression in human liver steatosis by high-density oligonucleotide microarray. Lab Invest. 2006;86(2):154-165.
- 32. Paradies G, Paradies V, Ruggiero FM, et al. Oxidative stress, cardiolipin and mitochondrial dysfunction in nonalcoholic fatty liver disease. World J Gastroenterol. 2014;20(39):14205-14218.
- 33. Van Remmen H, Hamilton ML, Richardson A. Oxidative damage to DNA and aging. Exerc Sport Sci Re. 2003;31(3):149-153.
- 34. Musatov A, Carroll CA, Liu YC, et al. Identification of bovine heart cytochrome c oxidase subunits modified by the lipid peroxidation product 4-hydroxy-2-nonenal. Biochemistry. 2002;41(25):8212-8220.
- 35. Esterbauer H, Schaur RJ, Zollner H. Chemistry and biochemistry of 4-hydroxynonenal, malonaldehyde and related aldehydes. Free Radic Biol Med. 1991;11(1):81-128.
- 36. Gardner HW. Oxygen radical chemistry of polyunsaturated fatty acids. Free Radic Biol Med. 1989;7(1):65-86.
- 37. Wagner BA, Buettner GR, Burns CP. Free radical-mediated lipid peroxidation in cells: oxidizability is a function of cell lipid bis-allylic hydrogen content. Biochemistry. 1994;33(15):4449-4453.
- 38. Jump DB, Lytle KA, Depner CM, et al. Omega-3 Polyunsaturated Fatty Acids as a Treatment Strategy for Nonalcoholic Fatty Liver Disease. Pharmacol Ther. 2017.
- 39. Georgiadi, Kersten S. Mechanisms of Gene Regulation by Fatty Acids. Adv Nutr. 2012;3(2):127-134.
- 40. Burdge G. Calder P. a-Linolenic acid metabolism in adult humans: the effects of gender and age on conversion to longer-chain polyunsaturated fatty acids. Eur J Lipid Sci Technol. 2005;107(6):426439.
- 41. Williams C, Burdge G. Long-chain n-3 PUFA: plant v. marine sources. Proc Nutr Soc. 2006;65(1):42-50.
- 42. Sassa T, Kihara A. Metabolism of very long-chain Fatty acids: genes and pathophysiology. Biomol Ther (Seoul). 2014;22(2):83-92.
- 43. Caldwell S. NASH Therapy: omega 3 supplementation, vitamin E, insulin sensitizers and statin drugs. Clin Mol Hepatol. 2017;23(2):103-108.
- 44. Serhan CN, Chiang, N, Van Dyke TE. Resolving inflammation: dual anti-inflammatory and proresolution lipid mediators. Nat Rev Immunol. 2008;8(5):349-361.
- 45. Mozaffarian D, Wu J. Omega-3 fatty acids and cardiovascular disease: effects on risk factors, molecular pathways, and clinical events. J Am Coll Cardiol. 2011;58(20):2047-2067.

- 46. Geleijnse JM, Giltay EJ, Grobbee DE, et al. Blood pressure response to fish oil supplementation: metaregression analysis of randomized trials. J Hypertens. 2002;20(8):1493-1499.
- 47. Mozaffarian D, Geelen A, Brouwer IA, et al. Effect of fish oil on heart rate in humans: a meta-analysis of randomized controlled trials. Circulation. 2005;112(13):1945-1952.
- Stirban A, Nandrean S, Gotting C, et al. Effects of n-3 fatty acids on macro- and microvascular function in subjects with type 2 diabetes mellitus. Am J Clin Nutr. 2010;91(3):808-813.
- 49. Leeson CP, Mann A, Kattenhorn M, et al. Relationship between circulating n-3 fatty acid concentrations and endothelial function in early adulthood. Eur Heart J. 2002;23(3):216-222.
- 50. Dangardt F, Osika W, Chen Y, et al. Omega-3 fatty acid supplementationimproves vascular function and reduces inflammation in obese adolescents. Atherosclerosis. 2010;212(12):580-585.
- 51. Sekikawa A, Curb JD, Ueshima H, et al. Marine-derived n-3 fatty acids and atherosclerosis in Japanese, Japanese-American, and white men: a crosssectional study. J Am Coll Cardiol. 2008;52(6):417-424.
- 52. Dai XW, Zhang B, Wang P, et al. Erythrocyte membrane n-3 fatty acid levels and carotid atherosclerosis in Chinese men and women. Atherosclerosis. 2014;232(1):79-85.
- 53. Bhatia L, Scorletti E, Curzen N, et al. Improvement in non-alcoholic fatty liver disease severity is associated with a reduction in carotid intima-media thickness progression. Atherosclerosis. 2016;246:13-20.
- 54. Lorenz MW, Markus HS, Bots ML, et al. Prediction of clinical cardiovascular events with carotid intima-media thickness: a systematic review and meta-analysis. Circulation. 2007;115(4):459-467.
- 55. Sookoian S, Pirola CJ. Non-alcoholic fatty liver disease is strongly associated with carotid atherosclerosis: a systematic review. J Hepatol. 2008:49(4):600-607.
- 56. Chinetti G, Fruchart JC, Staels B. Peroxisome proliferator-activated receptors: new targets for the pharmacological modulation of macrophage gene expression and function. Curr Opin Lipidol. 2003;14(5):459-468.
- 57. Pascual G, Fong AM, Ogawa S, et al. A SUMOylation-dependent pathway mediates transrepression of inflammatory response genes by PPAR-gamma. Nature. 2005;437(7059):759-763.
- 58. Mandard S, Patsouris D. Nuclear control of inflammatory response in mammals by peroxisome proliferator-activated receptors. PPAR Res. 2013:2013:613864.
- 59. Odegaard JI, Ricardo Gonzalez RR, Goforth MH, et al. Macrophagespecific PPARgamma controls alternative activation and improves insulin resistance. Nature. 2007;447(7148):1116-1120.
- 60. Takahashi M, Tsuboyama Kasaoka N, Nakatani T, et al. Fish oil feeding alters liver gene expressions to defend against PPARalpha activation and ROS production. Am J Physiol Gastrointest Liver Physiol. 2002;282(2):G338-G348.
- 61. Gao L, Wang J, Sekhar K, et al. Novel n-3 Fatty Acid Oxidation Products Activate Nrf2 by Destabilizing the Association between Keap1 and Cullin3. J Biol Chem. 2007;282(4):2529-2537.
- 62. Brown JD, Plutzky J. Peroxisome Proliferator Activated Receptors as Transcriptional Nodal Points and Therapeutic Targets. Circulation. 2007;115(4):518-533.
- 63. Farias J, Molina V, Carrasco R, et al. Antioxidant Therapeutic Strategies for Cardiovascular Conditions Associated with Oxidative Stress. Nutrients. 2017;9(9):E966.
- 64. Depner CM, Philbrick KA, Jump DB. Docosahexaenoic acid attenuates hepatic inflammation, oxidative stress, and fibrosis without decreasing hepatosteatosis in a Ldlr (-/-) mouse model of western diet-induced nonalcoholic steatohepatitis. J Nutr. 2013;143(3):315-323.

- Lytle KA, Depner CM, Wong CP, et al. Docosahexaenoic acid attenuates western diet induced hepatic fibrosis in Ldlr-/- mice by targeting the TGF-beta-Smad pathway. *J Lipid Res.* 2015;56(10):1936–1946.
- 66. Vognild E, Elvevoll EO, Brox J, et al. Effects of dietary marine oils and olive oil on fatty acid composition, platelet membrane fluidity, platelet responses, and serum lipids in healthy humans. *Lipids*. 1998;33(4):427–436.
- Zhu FS, Liu S, Chen X, et al. Effects of n-3 polyunsaturated fatty acids from seal oils on nonalcoholic fatty liver disease associated with hyperlipidemia. World J Gastroenterol. 2008;14(41):6395–6400.
- Masterton GS, Plevris JN, Hayes PC. Review article: omega-3 fatty acids-a promising novel therapy for non-alcoholic fatty liver disease. *Aliment Pharmacol Ther*. 2010;31(7):679–692.
- Dongiovanni P, Valenti L. A Nutrigenomic Approach to Non-Alcoholic Fatty Liver Disease. *Int J Mol Sci.* 2017;18(7):E1534.
- Ferre P, Foufelle F. Hepatic steatosis: a role for de novo lipogenesis and the transcription factor SREBP-1c. *Diabetes Obes Metab.* 2010;12:83–92.
- Elam M, Yellaturu C, Howell G, et al. Dysregulation of sterol regulatory element binding protein–1c in livers of morbidly obese women is associated with altered suppressor of cytokine signaling–3 and signal transducer and activator of transcription–1 signaling. *Metabolism*. 2010;59(4):587–598.
- Shimomura I, Bashmakov Y, Horton J. Increased Levels of Nuclear SREBP-1c Associated with Fatty Livers in Two Mouse Models of Diabetes Mellitus. *J Biol Chem.* 1999;274(42):30028–30032.
- Shimomura I, Matsuda M, Hammer R, et al. Decreased IRS-2 and Increased SREBP-1c Lead to Mixed Insulin Resistance and Sensitivity in Livers of Lipodystrophic and ob/ob Mice. Mol Cell. 2000;6(1):77–86.
- Dasarathy S, Dasarathy J, Khiyami A, et al. Double blind randomized placebo controlled clinical trial of omega 3 fatty acids for the treatment of diabetic patients with nonalcoholic steatohepatitis. *J Clin Gastroenterol*. 2015;49(2):137–144.
- Sanyal AJ, Abdelmalek MF, Suzuki A, et al. No significant effects of ethyl-eicosapentaenoic acid on histologic features of nonalcoholic steatohepatitis in a Phase 2 trial. *Gastroentero*. 2014;147(2):377–384.
- Nobili V, Bedogni G, Alisi A, et al. Docosahexaenoic acid supplementation decreases liver fat content in children with non-alcoholic fatty liver disease: double blind randomized controlled clinical trial. *Arch Dis Child*. 2011;96(4):350–353.
- Nobili V, Alisi A, Della Corte C, et al. Docosahexaenoic acid for the treatment of fatty liver: Randomized controlled trial in children. *Nutr Metab Cardiovasc Dis.* 2013;23(11):1066–1070.
- Nobili V, Carpino G, Alisi A, et al. Role of docosahexaenoic acid treatment in improving liver histology in pediatric nonalcoholic fatty liver disease. *Plos One.* 2014;9(2):e88005.
- Boyraz M, Pirgon O, Dundar B, et al. Long-term treatment with n-3 polyunsaturated fatty acids as a monotherapy in chicken with nonalcoholic fatty liver disease. J Clin Res Pediatr Endocrinol. 2015;7(2):121–127.
- Janczyk W, Labensztejn D, Wierzbicka Rucinska A, et al. Omega-3 fatty acids therapy in children with nonalcoholic fatty liver disease: a randomized control trial. J Pediatr. 2015;166(6):1358–1363.
- Pacifico L, Bonci E, Di Martino M, et al. A double-blind, placebocontrolled randomized trial to evaluate the efficacy of docosahexaenoic acid supplementaion on hepatic fat and associated cardiovascular factors in overweight children with nonalcoholic fatty liver disease. *Nutr Meta Cardiovasc Dis.* 2015;25(8):734–741.
- Scorletti E, Bhatia L, McCormic KG, et al. Effects of purified eicosapentaenoic acid and docosahexaenoic acids in nonalcoholic fatty lifer disease: Results from the Welcome Study. *Hepatology*. 2014;60(4):1211–1221.

- 83. Argo CK, Patrie JT, Lackner C, et al. Effects of n-3 fish oil on metabolic and histological parameters in NASH: A double-blind, randomized, placebo-controlled trial. *J Hepatol*. 2015;62(1):190–197.
- Li YH, Yand LH, Sha KH, et al. Efficacy of poly-unsaturated fatty acid therapy on patients with nonalsoholic steatohepatitis. World J Gastroentero. 2015;21(12):7008–7013.
- 85. Qin Y, Zhou Y, Chen SH, et al. Fish oil supplements lower serum lipids and gllucose in correlation with a reduction in plasma fibroblast growth factor 21 and prostaglandin E2 in nonalcoholic fatty liver disease associated with hyperlipidemia: A randomized clinical trial. *Plos One.* 2015;10(7):e0133496.
- Nogueira MA, Oliveira CP, Alves VAF, et al. Omega-3 polyunsaturated fatty acids in treating non-alcoholic steatohepatitis: A randomized, couble-bline, placebo-controlled trial. Clin Nutr. 2016;35(3):578–586.
- 87. Della Corte C, Carpino G, De Vito R, et al. Docosahexanoic Acid Plus Vitamin D Treatment Improves Features of NAFLD in Children with Serum Vitamin D Deficiency: Results from a Single Centre Trial. *Plos One.* 2016;11(12):e0168216.
- 88. Hodson L, Bhatia L, Scorletti E, et al. Docosahexaenoic acid enrichment in NAFLD is associated with improvements in hepatic metabolism and hepatic insulin sensitivity: a pilot study. *Eur J Clin Nutr.* 2017;71(8):973–979.
- Institute of Medicine (US) Panel on Dietary Antioxidants and Related Compounds. Dietary Reference Intakes for Vitamin C, Vitamin E, Selenium, and Carotenoids. Washington (DC): National Academies Press (US). 2000.
- Jiang Q, Christen S, Shigenaga MK, et al. Gamma-tocopherol, the major form of vitamin E in the US diet, deserves more attention. *Am J Clin Nutr.* 2001;74(6):714–722.
- Li J, Cordero P, Nguyen V, et al. The Role of Vitamins in the Pathogenesis of Non-alcoholic Fatty Liver Disease. *Integr Med Insights*. 2016;11:19–25.
- 92. Chun J, Lee J, Ye H, et al. Tocopherol and tocotrienol contents of raw and processed fruits and vegetables in the United States diet. *J Food Compos Anal*. 2006;19(2–3):196–204.
- 93. Mclaughlin PJ, Weihrauch JL. Vitamin E content of foods. *J Am Diet Assoc*. 1979;75(6):647–665.
- 94. Dreher ML. Pistachio nuts: composition and potential health benefits. *Nutr Rev.* 2012;70(4):234–240.
- Levy AP, Blum S. Pharmacogenomics in prevention of diabetic cardiovascular disease: utilization of the haptoglobin genotype in determining benefit from vitamin E. Expert Rev Cardiovasc Ther. 2007;5(6):1105–1111.
- Vardi M, Levy NS, Levy AP. Vitamin E in the prevention of cardiovascular disease: the importance of proper patient selection. J Lipid Res. 2013;54(9):2307–2314.
- 97. Levy AP, Gerstein HC, Miller Lotan R, et al. The effect of vitamin E supplementation on cardiovascular risk in diabetic individuals with different haptoglobin phenotypes. *Diabetes Care*. 2004;27(11):2767.
- Ceriello PA. Oxidative stress and diabetes associated complications. *Endocr Pract.* 2006;12:60–62.
- Rodriguez Ramirez G, Simental Mendia LE, Carrera Gracia MA, et al. Vitamin E Deficiency and Oxidative Status are Associated with Prediabetes in Apparently Healthy Subjects. Arch Med Res. 2017;48(3):257–262.
- Jiang Q, Lykkesfeldt J, Shigenaga MK, et al. Gamma-Tocopherol supplementation inhibits protein nitration and ascorbate oxidation in rats with inflammation. Free Radic Biol Med. 2002;33(11):1534–1542.

170

- 101. Jiang Q, Ames BN. gamma-Tocopherol, but not alpha-tocopherol, decreases proinflammatory eicosanoids and inflammation damage in rats. FASEB J. 2003;17(8):816-822.
- 102. Chalasani N, Younossi Z, Lavine JE, et al. The diagnosis and management of non-alcoholic fatty liver disease: practice guideline by the American Association for the Study of Liver Diseases, American College of Gastroenterology, and the American Gastroenterological Association. Am J Gastroenterol. 2012;107(6):811-826.
- 103. Oseini AM, Sanyal AJ. Therapies in non-alcoholic steatohepatitis (NASH). Liver Int. 2017;37:97-103.
- 104. Kim GH, Chung JW, Lee JH, et al. Effect of vitamin E in nonalcoholic fatty liver disease with metabolic syndrome: A propensity score-matched cohort study. Clin Mol Hepatol. 2015;21(4):379-386.
- 105. Magosso E, Ansari MA, Gopalan Y, et al. Tocotrienols for normalisation of hepatic echogenic response in nonalcoholic fatty liver: a randomised placebo-controlled clinical trial. Nutr J. 2013;12(1):166.
- 106. Dadamo E, Marcovecchio ML, Giannini C, et al. Improved oxidative stress and cardio-metabolic status in obese prepubertal children with liver steatosis treated with lifestyle combined with Vitamin E. Free Radic Res. 2013;47(3):146-153.
- 107. Sanyal AJ, Chalasani N, Kowdley KV, et al. Pioglitazone, vitamin E, or placebo for nonalcoholic steatohepatitis. N Engl J Med. 2011;362(18):1675-1685.

- 108. Lavine JE, Schwimmer JB, Van Natta ML, et al. Effect of vitamin E or metformin for treatment of nonalcoholic fatty liver disease in children and adolescents: the TONIC randomized controlled trial. JAMA. 2011;305(16):1659-1668.
- 109. Sawangjit R, Chongmelaxme B, Phisalprapa P, et al. Comparative efficacy of interventions on nonalcoholic fatty liver disease (NAFLD), A PRISMA-compliant systematic review and network meta-analysis. Medicine. 2016;95(32):e4529.
- 110. Peter S, Moser U, Pilz S, et al. The challenge of setting appropriate intake recommendations for vitamin E: considerations on status and functionality to define nutrient requirements. Int J Vitam Nutr Res. 2013;83(2):129-136.
- 111. Raederstorff D, Wyss A, Calder PC, et al. Vitamin E function and requirements in relation to PUFA. Br J Nutr. 2015;114(8):1113-1122.
- 112. Atkinson J, Harroun T, Wassall SR, et al. The location and behavior of alpha-tocopherol in membranes. Mol Nutr Food Res. 2010;54(5):641-651.
- 113. Zhao Y, Monahan FJ, McNulty BA, et al. Plasma n-3 polyunsaturated fatty status and its relationship with vitamin E intake and plasma level. Eur J Nutr. 2017;56(3):1281-1291.