

Mechanisms, diagnosis and management of eosinophilic asthma

Abstract

Asthma is a common chronic airway disease affecting about 334 million people worldwide, and up to 10% of asthma patients have severe asthma, which may be uncontrolled despite high doses of the standard treatment modifiers and may require the use of chronic oral corticosteroids. It is the most common chronic disease in children in the developed countries. Asthma manifests as reversible airflow obstruction, due to airway inflammation, bronchial smooth muscle contraction, increased mucus secretion, vascular engorgement, mucosal oedema, and airway hyper responsiveness, which leads to airflow obstruction and symptoms of asthma. Eosinophilic asthma is a phenotype of asthma that is usually very severe and persistent, with frequent exacerbations. It is usually observed in adult asthmatic patients, although it may occur in children. It is characterized by the presence of high levels of eosinophils, and CD+4 Th2 cells in the lungs and airways, which can be demonstrated by a raised eosinophil count in blood, and induced sputum or bronchial biopsy. It is managed in a similar stepwise treatment for childhood-onset asthma, but some of the patients with eosinophilic asthma do not respond to this standard treatment including inhaled or oral corticosteroids. The logical approach to treat corticosteroid-refractory asthma is to target the eosinophilic interleukins which cause airway inflammation using monoclonal antibodies to block their activity on the eosinophils, and Th2 cells. Currently, the following monoclonal antibodies are used in the treatment of eosinophilic asthma: IgE antibody such as omalizumab, or interleukin receptor 5, or 4, and 13 antagonists, such as mepolizumab, reslizumab, and dupilumab. These novel agents have proved to be very useful in relieving the symptoms, and in improving the forced expired volume in one second (FEV1), and in reducing exacerbations. They are also steroid-sparing agents, and improve the quality of life in this debilitating phenotype of asthma.

Keywords: eosinophilic asthma, asthma phenotypes, inflammatory mediators, monoclonal antibodies, interleukin receptor antagonists

Volume 7 Issue 2 - 2020

Nightingale Syabbalo

Department of Medicine and Physiology, Copperbelt University, Zambia

Correspondence: Nightingale Syabbalo, Professor of Medicine and Physiology, Copperbelt University, M. C. Sata School of Medicine, P. O. Box 30243, Lusaka, Zambia, Tel +260 966 486117, Email nightsya@gmail.com

Received: March 14, 2020 | **Published:** April 02, 2020

Abbreviations: AHR, airway hyper responsiveness; FEV1, forced expired volume in one second; Th2, T-helper type 2; ILs, interleukins; ILC2s, innate lymphoid cells; GATA-1, GATA-binding protein 1; c/EBP, CCAAT-enhancing binding protein; VCAM-1, vascular cell adhesion molecule-1; MBP, major basic protein; ECP, eosinophil cationic protein; EDN, eosinophil-derived neurotoxin; EDPX, eosinophil-derived peroxide

Introduction

Asthma is a common chronic airway disease affecting about 334 million people worldwide, and its prevalence has been increasing during the last 40 years, and by 2025, there will be about 400 million people suffering from asthma.^{1,2} It is the most common chronic respiratory disease in children in the developed countries,³ and its prevalence is steadily increasing in the developing world. Asthma is characterized by reversible airflow obstruction, due to airway inflammation, bronchial smooth muscle contraction, increased mucus secretion, vascular engorgement and mucosal oedema, and airway hyper responsiveness (AHR), which leads to airflow obstruction.⁴ Airway obstruction and bronchospasm leads to the symptoms of asthma which include wheezing, coughing, dyspnoea, and chest tightness.

The pathophysiology of asthma is complex, it is a heterogeneous disease with several different phenotypes, including eosinophilic asthma, neutrophilic asthma, paucigranulocytic asthma, and childhood-onset asthma. It involves imbalance between T-helper

type 2 (Th2) lymphocytes and Th1 lymphocyte-driven airway inflammation, switching the balance towards the Th2 pathway.⁵⁻⁸ Th2-driven inflammation leads to production and liberation of an array of inflammatory mediators such as histamine, prostaglandins, leukotrienes, cytokines and chemokines.⁹ Table 1 shows the inflammatory mediators responsible for the pathophysiology of asthma.

Table 1 Inflammatory mediators of Th2-driven asthma

Histamine
Leukotrienes (LTB4, LTC4, LTD4, LTE4)
Eicosanoids (PGD2, TXB2)
Cytokines (Interleukin-1 β , IL3, IL-4, IL-5, IL-12, IL-13, IL-23, IL-33)
Thymic stromal lymphopoietin (TSLP)
Chemokines (MCP-1, MCP-3, MCP-4, RANTES, eotaxin)
Neurokinins (NK1)
Enzymes (tryptase)
Neutrophil chemotactic factor
Neutrophil myeloperoxidase
Platelet activating factor

Abbreviations: IL, interleukin; LT, leukotriene; PG, prostaglandins; Tx, thromboxane; MCP, monocyte chemotactic protein

The cytokines, especially interleukins (ILs) produced by activated Th2 lymphocytes include: IL-3, IL-4, IL-9, IL-13 which activates mast cells, and IL-3 and IL-5 which activates eosinophils.⁹ The activated mast cells and eosinophils further generate inflammatory mediators which perpetuate and amplifies the airway inflammatory process, and causes more bronchospasm. On the other hand, Th1 cells secrete IL-2 and interferon- γ , whilst group 2 innate lymphoid cells (ILC2s) are activated to Th2 cell by the action of IL-4, and IL-33. The transformed Th2 cells further liberate inflammatory mediators, thus causing a vicious airway inflammatory process. Th2 cytokines play a pivotal role in the pathophysiology of allergic disease, including asthma.^{10–12} Apart from activating mast cells and eosinophils they also take a crucial part in the inflammatory cascade.⁹

It is well now well recognized that asthma is a complex and diverse disease with several phenotypes or endotypes, and each of these phenotypes has a characteristic pathophysiological pathways and clinical presentation. Eosinophilic asthma is one of the well-defined clinical phenotype of asthma.^{13,14} It is usually a debilitating, severe and persistent disease, with frequent exacerbations and a worse quality of life, and has a poor prognosis.^{15–19} Patient with eosinophilic asthma experience significant limitation in lung function, frequent hospitalizations and are at higher risk of death.²⁰ This subgroup of patients impart a disproportionate pharmaco-economical burden, with the mean UK annual treatment costs reaching between £2912 and £4217 per patient.²¹ The disease is usually observed in adult asthmatic patients, although it may occur in children.²² It is characterized by a high eosinophil count in blood and in induced sputum, airway eosinophilia infiltration,²³ and high IgE and periostin serum levels.²⁴ Serum IgE and periostin levels are markers of eosinophilic inflammation.

The airway inflammation in eosinophilic asthma is due to degranulation of eosinophils activated by cytokines released by Th2 cells, dendritic cells, mast cells, and basophils. This results in the release of several pro-inflammatory mediators, including cytotoxic cationic proteins, histamine, prostaglandins, leukotrienes, cytokines, chemokines, and growth factors.^{25,26} Table 2 shows the list of inflammatory mediators released by eosinophils during allergic inflammation.

Table 2 Eosinophil-derived inflammatory mediators

Major basic protein (MBP)
Eosinophil cationic protein (ECP)
Eosinophil-derived neurotoxin (EDN)
Eosinophil-derived peroxide (EDPX)
Reactive oxygen species (superoxide, peroxide, and hypobromite)
Prostaglandins (PGE2)
Leukotrienes (LTC4, LTD4, LTE4)
Cytokines (IL-2, IL-3, IL-4, IL-5, IL-9, IL-13, IL-23, IL-25, IL-33, and TNF α)
Chemokines (CXC-, CC-, CX3C, and XC)
Enzymes (elastase)
Growth factors (TGF β , VEGF, and PDGF)

Abbreviations: IL, interleukin; chemokine nomenclature depends on the spacing of conserved cysteines, where X is any amino acid

Cytokines such as IL3, IL-5, IL-4, IL-12, IL-13, IL-23, IL-33, and thymic stromal lymphopoietin (TSLP) play a critical role in orchestrating, perpetuating, and amplifying the respiratory response in asthma. Cytokines cause bronchial smooth muscle contraction, mucus secretion, microvascular leakage and airway oedema. All of these pathophysiological processes lead to airway narrowing and bronchoconstriction.^{26–28} Long-term effects of the cytokines, particularly due to IL-13, include goblet gland hyperplasia, smooth muscle hypertrophy, airway remodeling, and bronchial hyperreactivity.²⁷

Th2 lymphocyte and eosinophil cytokines possess overlapping biological activities; they can synergize or antagonize the effects of other cytokines. For example, IL-5, IL-4, IL-13, and IL-33 are the key drivers of the inflammatory process in asthma; and IL-4 and IL-13 are central Th2 cytokines with distinct overlapping roles, particularly in airway remodeling and bronchial hyperresponsiveness.²⁹ Similarly, interferon- γ , a Th1 cytokine acts in conjunction with Th2 cytokines (IL-3, IL-4, and IL-5) in maintaining chronic airway inflammation in patients with asthma.

Eosinophils

The eosinophil was first described by Paul Ehrlich in 1879, after he developed eosin which colored basic protein bright red. It was identified as eosinophil in an autopsy of a 48-year-old male patient who died of status asthmaticus by Dr. Fraenkel in 1900.³⁰ Eosinophils are polymorphonuclear cells and they normally constitute about 2.3% of all blood leukocytes. They are slightly larger than neutrophils, about 10–16 μ m in diameter, and are characterized by a nucleus with usually two lobes, and large cytoplasmic granules that stain deeply red after staining with eosin, using the Romanowsky method. Eosinophils and other leukocytes are formed from bone marrow CFU-GM progenitor cells during myelopoiesis. The pluripotent myeloid progenitor cells give rise to CD34+ IL-5R α eosinophil progenitor cells. The differentiation of eosinophils is regulated by transcription factors GATA-binding protein 1 (GATA-1), PU.1, and the CCAAT-enhancing binding protein (c/EBP) family. IL-5, IL-3, and GM-CSF synergistically contribute to the development of mature eosinophils. After being released from the bone marrow, they mature in the circulation and migrate to tissues where they live for 2–5 days. Their life span may be extended depending on tissue cytokines, particularly IL-4, IL-5, and IL-13 which prevent them from apoptosis. Eosinophils are weak phagocytes, and exhibit chemotaxis, and diapedesis. They appear to be used selectively for fighting helminth parasitic infections, particularly intestinal nematodes (*Ancylostoma duodenale*); tissue-dwelling nematodes (filarial worms); trematodes (*Schistosoma haematobium* and *S. mansoni*); and cestodes (*Taenia saginata*, and *Diphyllobothrium latum*). The cationic proteins secreted by the eosinophils, particularly major basic protein, and eosinophilic cationic protein produce ballooning and detachment of the tegmental membrane, complete fragmentation and complete disruption of the large multicellular helminth parasites.

Eosinophils have a special propensity to collect in tissues in which allergic reactions occur, such as the peribronchial tissues of the lungs in people with asthma, and in the skin after an allergic reaction. Mast cells and basophils release an eosinophil chemotactic factor that causes eosinophils to migrate toward the inflamed allergic tissue such as the bronchial mucosa.

Eosinophils play a pivotal role in the pathogenesis of asthma.

They possess a wide repertoire of surface adhesion molecules and receptors such as cytokine and growth receptors, lipid mediator receptors, chemoattractant receptors, adhesion receptors, and Fc receptors.²⁵ Eosinophilic inflammatory activity is regulated by several Th2 cytokines, including IL-5, IL-3, IL-4, IL-13, IL-33, IL-17, IL-25, and TSLP.

Interleukin-5 is a homodimeric cytokine (115 amino acids per chain) that belong to the haematopoietic growth factor cytokine family, which also include IL-3 and GM-CSF. Interleukin-5 stimulates production, proliferation, and differentiation of eosinophils from myeloid progenitor cells in the bone marrow. Peripherally, it participates in the terminal maturation of the eosinophil in the circulation, recruitment and activation in the lungs, and is important for eosinophil survival. It plays an important role in diapedesis of eosinophils by facilitating endothelial adhesion, and promotes chemotaxis.³¹ Adhesion receptor such as integrins allow cell such as the eosinophil to adhere to the extracellular matrix and other cells plays a major role in eosinophilic migration. They also allow the eosinophil to sense their surrounding and respond accordingly.^{32, 33}

IL-4 plays a key role in eosinophilic inflammatory response, which include induction of the IgE isotype switch, expression of vascular cell adhesion molecule-1 (VCAM-1), differentiation of Th2 lymphocyte leading to cytokine release, mucus secretion, and promoting eosinophil transmigration across the endothelium.³⁴ In addition IL-4 and IL-13 drive the trafficking of eosinophils to sites of allergic inflammation. It is very important in activating eosinophils, and leads to eosinophil degranulation and release of inflammatory mediators.

Upon activation, eosinophils degranulate and release an array of cytotoxic cationic proteins, such as major basic protein (MBP), eosinophil cationic protein (ECP), eosinophil-derived neurotoxin (EDN), eosinophil-derived peroxidase (EDPX). In addition, they release a plethora of mediators including prostaglandins, leukotrienes, cytokines, chemokine, enzymes and reactive oxygen species. Eosinophils synthesize and release several interleukins, such as IL-3, IL-4, IL-5, IL-9, IL-13, IL-15, IL-23, IL-25, IL-33, and TSLP. These inflammatory mediators orchestrate, and amplify bronchial smooth muscle contraction, microvascular leakage and airway oedema, mucus secretion, and goblet gland hyperplasia.^{35, 36} The cationic proteins (MBP, ECP, EDN, and EDPX) are very cytotoxic to the airway epithelium and myelinated neurones, and they cause epithelial and neuronal injury, and damage. EDPX form reactive oxygen species and reactive nitrogen metabolites that promote oxidative stress, causing cell death by apoptosis and necrosis of epithelial cells. In addition the eosinophilic inflammatory mediators lead to airway epithelial injury, smooth muscle hypertrophy, airway remodeling, and airway hyperresponsiveness.^{35, 36}

IL-13 is a pleiotropic Th2 cytokine that has been shown to be central in the pathogenesis of asthma. It is a key inflammatory cytokine, it causes goblet cell differentiation, mucus secretion, elevation of bronchial hyperresponsiveness, and switching of B cell antibody production from IgG to IgE. Interleukin-13 also causes activation of fibroblasts and sub endothelial fibrosis and eotaxin production.³⁵ The eotaxins 1 (CCL-11), 2 (CCL-24), and 3 (CCL-26) subfamily of chemokines and their CCR3 chemokine receptor have coordinated interaction with IL-13 in the pathogenesis of asthma.^{37, 31} They are involved in the recruitment, and in inducing eosinophilic degranulation. Eotaxin-2, and eotaxin-3 are associated with persistent eosinophilic bronchial inflammation in patients with asthma after allergen challenge.

The bronchial epithelial cytokines ('alarmins'), IL-33, IL-25, and TSLP play an important role act in activating innate lymphoid cells (ILC2s) in an antigen independent manner through their receptors (IL-17 receptor B (IL-17RB), ST2, and TSLP receptor (TSLPR)). These cytokines, particularly IL-25 and IL-33 have been implicated in the airway remodeling process. Airway structural changes can result in a loss of elastic recoil with increased lung compliance, most pronounced at the peri-bronchiolar level, resulting in irreversible obstruction. IL-33 a member of the IL-1 family, in particular, triggers eosinophils to release their cytotoxic pro-inflammatory mediators, and superoxide generation. This result in a vicious cycle of inflammation promoting further airway epithelial cell injury, and inflammation, which can progress to chronic eosinophilic inflammation.

Activated innate lymphoid cells (ILCs) or Th0 cells produce large amounts of IL-5 and IL-13, which can lead to mucus hyper secretion, airway hyper responsiveness, and both cytokines are capable of inducing eosinophilic airway inflammation independent of T-cells.³⁸ ILC2s have also been shown to be essential for the persistence of eosinophilic asthma.³⁵ IL-33 is a central regulator of immunologic reaction. Up regulation of IL-33 results in activation of IL-13, which in turn activates Th2 cells, dendritic cell, eosinophils, and basophils, leading to the release of a cascade of mediators.

One of the best approaches to treat eosinophilic asthma is to block the actions of interleukins which are important in eosinophils production, proliferation, differentiation, activation, and survival. This will reduce eosinophil numbers and function, and the release of the injurious inflammatory mediators.

Clinical features

Eosinophilic asthma is a severe refractory disease which occurs in about 4% of adult patients with asthma.³⁶⁻³⁹ The disease has no gender preponderance in the distribution.³⁸ Eosinophilic asthma usually manifests in early adulthood with a peak incidence between 20 to 35 years. The classical symptoms include recurrent episodes of wheezing, breathlessness, chest tightness, and cough. The symptoms are usually worse at night (nocturnal asthma). Cough is a frequent symptom particularly in children and it may be mistaken for an upper respiratory tract infection or bronchitis. Precipitating factors include viral upper respiratory tract infection, exercise particularly in cold weather, exposure to pollutants such as cigarette smoke, SO₂, NO₂, and ozone, and medications (Table 3). The drugs which are likely to precipitate asthma include beta-blockers, even when administered topically as eye drops, e.g., timolol for glaucoma, aspirin and other non-steroidal anti-inflammatory drugs (NSAIDs). Other medications include oral contraceptives, cholinergic agents, and prostaglandin F_{2a}.

There is a tremendous variation in the frequency and duration of the attacks. Some patients have one or two attacks per year, but the majority of the patients have severe frequent exacerbations which may last for weeks. Some patients have chronic symptoms. Patients with mild asthma are usually asymptomatic between exacerbations, whereas patients with persistent asthma have symptoms of breathlessness and wheeze most of the time, particularly in early mornings.

Apart from displaying the above clinical features, patients with eosinophilic asthma have unique clinical presentation, which differ greatly from those of the classical symptoms of childhood-onset, allergic asthma. This phenotype of asthma is rarely precipitated by allergens such as grass pollen, pet dander allergens, and

Dermatophagoides pteronyssinus compared to childhood-onset asthma.¹⁹ Eosinophilic asthma is very easily provoked by exertion or exercise, and patients with eosinophilic or late-onset asthma are very sensitive to aspirin, and may have co morbid aspirin exacerbated respiratory disease.⁴⁰ Clinically the disease is very severe with frequent exacerbations to near fatal asthma which requires frequent hospitalization.^{17–21,41} Patients have a poor quality of life and the prognosis of the disease is poor.^{17–21}

Table 3 Causes and precipitating factors for eosinophilic asthma

Viral upper and lower respiratory infections

Rhinovirus

Parainfluenza virus

Respiratory syncytial virus

Occupational sensitizers

Isocyanate

Colophony fumes

Atmospheric pollution

Sulphur dioxide, nitrogen dioxide

Ozone

Irritant dusts, vapour and fumes

Cigarette smoke

Perfumes

Exercise

Food anaphylaxis

Shrimps, peanuts, wheat allergy

Medication

Aspirin, non-steroidal anti-inflammatory drugs

β₂-blockers

Angiotensin-converting enzyme (ACE) inhibitors

Aeroallergens (rare)

Dermatophagoides

Grass pollen

Domestic pet dander

Cockroaches

Eosinophilic asthma is often associated with chronic rhino sinusitis and nasal polyposis, which should be treated.⁴² Treatment of these co morbidities, is associated with improvement in asthma symptoms. Additionally, patients with eosinophilic asthma have persistent airflow obstruction characterized by constantly very low forced expired volume in 1 second (FEV₁).⁴³ As a result of fixed low FEV₁, and FEV₁/FVC ratio, eosinophilic asthma may be misdiagnosed as chronic obstructive pulmonary disease (COPD).

The disease is difficult to treat with the standards-of-care asthma control medicines and can require the use of high dose inhaled corticosteroids or oral corticosteroids, thus, it is also referred to as corticosteroid-refractory disease or “difficult asthma”.⁴⁴ The patients may require large doses of oral corticosteroids which may cause the numerous serious side-effects from the glucocorticoids.

Investigations

The diagnosis of asthma is predominantly clinical and based on a characteristic history. It is supported by measurement of pulmonary function. Spirometry should be performed in all patients suspected of having asthma. It shows an obstructive pattern characterized by a low FEV₁ and the FEV₁/FVC ratio, and a reduction in flow rates at 25% to 75% of the vital capacity (FEF_{25–75}). If possible, a methacholine or histamine bronchoprovocation test should be performed in order to demonstrate airway hyperreactivity.^{45,46} Both the American Thoracic Society (ATS) and the European Respiratory Society (ERS) recommend a reduction of 10% or more in FEV₁ or PEF in the laboratory as criterion for the diagnosis of asthma.^{47,48} A post-bronchodilator response to β₂-adrenoceptor agonists such as salbutamol may be required in patients with bronchospasms to assess reversibility. The criterion for a bronchodilator response recommended by the ERS is a 12% increase in FEV₁, expressed as a per cent predicted after inhaled bronchodilator or a 200ml increase in FEV₁ (Table 4).⁴⁷

Table 4 Drugs for the treatment of asthma

Inhaled β₂-agonist

Short acting (salbutamol, levalbuterol, terbutaline, pirbuterol)

Long-acting (salmeterol, formoterol)

Combination of LABA and inhaled corticosteroids

Salmeterol and fluticasone (AdvairDiskus)

Formoterol and budesonide (Symbicort)

Cromones

Cromolyn sodium, nedocromil sodium

Inhaled anti-cholinergics

Short-acting (ipratropium bromide)

Long-acting (oxitropium bromide, tiotropium bromide)

New long-acting (aclidium bromide, glycopyrronium)

Corticosteroids

Betamethasone dipropionate

Budesonide, fluticasone, flunisolone

Ciclesonide, mometasone

Oral methylxanthines

Rapid release theophyllines

Sustained release theophyllines (Theo-24, Theocron, Uniphyll)

Leukotriene receptor antagonists

Montelukast, pranlukast

Cinalukast, zafirlukast

5-lipoxygenase inhibitors

Zileuton

Patients with eosinophilic asthma are very likely to develop exercise-induced asthma because of the association of the disease with chronic rhino sinusitis and nasal polyposis,³⁹ which impair air conditioning by the nasal apparatus. Exercise spirometry may be an adjunct test in this phenotype of asthma to document exercise-induced bronchoconstriction (EIB). Eosinophilic asthmatic patients have very

variable and frequent attacks of asthma, ideally they should have peak flow charts, recording the peak expiratory flow rates (PEF) on waking, in the middle of the day, and before bed.

Specific tests for eosinophilic asthma include eosinophil count in blood (≥ 300 cells/ μ L), and induced sputum ($\geq 2\%$),²³ serum IgE (≥ 250 kU/L), and exhaled fraction of nitric oxide (FeNO).⁴⁹ FeNO can also be used to monitor the response to treatment with ICSs. Serum periostin levels can be measured which is a signature of IL-13 and eosinophilic inflammation. Periostin is an extracellular matrix protein belonging to the fasciclin family, which contributes to airway remodeling in patients with eosinophilic asthma. Periostin serum levels have been described as the best predictor of sputum eosinophilia,²⁴ and can be used to monitor response to treatment.

There are no diagnostic features of asthma on chest roentgenogram in stable patients. Due to the severity of eosinophilic asthma, a chest X-ray is necessary to exclude hyperinflation and barotraumas including pneumothorax and pneumomediastinum. Hospitalized patients with severe asthma require oximetry monitoring, and blood gases analysis.

Management

The goals of asthma treatment are to achieve disease control. Poor control is linked with recurrent asthma attacks which is associated with poor future control.⁵⁰ Like any other individuals with asthma, patients with eosinophilic asthma should be treated according to the BTS/SIGN guidelines,⁵¹ or the ERS/ATS guidelines.⁵² Patients with uncontrolled asthma should have their treatment intensified by escalating up the treatment steps until control is achieved for at least 3 months.⁵¹ Corticosteroids are the mainstay therapy for patient with severe, recurrent disease. However, about 10-20% of the patients do not achieve symptoms control despite high doses of inhaled corticosteroids up to 2000 μ g/day, and require chronic use of oral corticosteroids.⁵³ It is important to try to achieve control without resorting to oral steroids which are linked with osteoporosis, adrenal suppression, hypertension, hypercholesterolemia, cataract, weight gain and diabetes.^{54, 55}

This subgroup of patients with eosinophilic asthma or corticosteroid-refractory asthma requires an additional or alternative therapeutic agent in order to achieve disease control. The underlying pathophysiology of asthma is airway inflammation and hyperresponsiveness due to inflammatory mediators release by activated mast cells, dendritic cells, Th2 lymphocytes and eosinophils. The best strategy for the treatment of eosinophilic asthma is to suppress the production, proliferation, differentiation and activation of eosinophils, which play a pivotal role in the airway inflammatory process. Interleukins, particularly, IL-5, IL-4, IL-13, IL-33, and TSLP play an important role in fostering eosinophilic function, chemotaxis and survival. Monoclonal antibodies and interleukin receptor antagonists (ILRAs) have been developed which target interleukins, with the aim of providing precision tailored treatment for patients with eosinophilic asthma. Table 5 shows the list of some of the mAbs and ILRAs currently available, and in clinical trials for the treatment of eosinophilic asthma, and corticosteroid-refractory asthma.

Immunoglobulin E released by activated mast cells, basophils and eosinophils plays an important role in the pathophysiology of the allergic inflammation in patients with asthma. Monoclonal antibodies targeted against IgE, such as omalizumab have been shown to attenuate both the early- and late-phase responses to inhaled allergens in patients with asthma.⁵⁶

Table 5 Monoclonal antibodies and interleukin receptor antagonists, and their target

Agent	Target	Stage of Development
Omalizumab	IgE	Marketed 2003
Mepolizumab	IL-5	Marketed 2015
Reslizumab	IL-5	Marketed 2016
Benralizumab	IL-5R	Marketed 2017
Dupilumab	IL-4 α (IL-4/IL-13)	Marketed 2018
Tezepelumab	TSLP	Marketed 2018
Pitrakinra	IL-4 α (IL-4/IL-13)	II
Lebrikizumab	IL-13	III
Tralokinumab	IL-13	III
Secukinumab	IL-17	II
Brodalumab	IL-17RA	II

Omalizumab

Omalizumab (Xolair®) was the first monoclonal antibody to be approved by the U. S. Food and Drug Administration (FDA) for the treatment of severe asthma in 2003. Xolair is a recombinant humanized monoclonal antibody to IgE, and is directed against the binding of IgE for its high affinity Fc ϵ R1 receptor.⁵⁷ It binds with the Fc portion of IgE and forms omalizumab: IgE complex. This reduces free IgE and prevents serum IgE from attaching to the Fc ϵ R1 receptors on mast cell, basophils and eosinophils. This prevents release of inflammatory mediators by these cells. In addition, omalizumab treatment indirectly reduces Fc ϵ R1 receptor on cells involved in the allergic responses.⁵⁷

Clinical trials using omalizumab treatment in patients with severe eosinophilic asthma has been shown to reduce airway and blood eosinophils counts, and reduce exacerbations in most patients with asthma.^{58, 59} Treatment with subcutaneous omalizumab has also been shown to improve asthma control, and improve health-related quality of life (HRQoL). It has also been shown to reduce the need for rescue medication, allow patients to reduce or discontinue their ICS and/OCSs.^{60, 61}

Xolair is given subcutaneously every two or four weeks depending on the patient's allergy status and body weight. It has an excellent safety profile. Patients need to be monitored for severe allergic reactions after the injection at a medical centre where health care professionals are available to treat the adverse reactions. It is safe and well tolerated. The most common side effects include injection site reaction, respiratory tract infection, pharyngitis, sinusitis, arthralgia, myalgia, muscle weakness, headache, and rarely anaphylaxis. Unfortunately, some patients with eosinophilic asthma do not get symptom relief with the addition of omalizumab to their treatment regimen, and may require an add-on treatment with another mAB, which target other airway inflammatory pathways.

Mepolizumab

Mepolizumab (Nucala®) is a fully humanized IgG1 monoclonal antibody, it binds to IL-5 and prevents binding to the α -chain of the IL-5 receptor.⁶² It was the first anti-IL-5 therapy to be tested in a clinical trial in 2000.⁶³ The first clinical trial of mepolizumab showed a reduction in sputum and blood eosinophil count but no change in bronchial hyper responsiveness, and no effect on the late asthmatic response.⁵⁹ Subsequent clinical trials revealed reduction in exacerbation

rates, and improvement in asthma symptom questionnaire (ACQ) scores.^{64, 65} Finally, Ortega and colleagues,⁶⁶ showed that treatment with mepolizumab decreases the rate of exacerbations, improved the FEV1, and reduced the dosage and use of oral corticosteroids, thus demonstrating a steroid-sparing effect. Convincingly, mepolizumab has been shown to improve the ACQ scores, FEV1, reduce the rate of exacerbations, and reduce the dosage of corticosteroid or use of other drug modifiers.^{64–66} Mepolizumab was approved by the FDA in March 2015 for the treatment of eosinophilic asthma. It is recommended at a dosage of 100mg subcutaneously every 4 weeks. It is well tolerated and it has been found to be safe. The most common adverse effects with Nucala are: injection site reaction, headache, backache, fatigue, muscle weakness, and rarely severe allergic reactions. Patients need to be monitored after treatment.

Reslizumab

Reslizumab (Cingair®) was approved by the FDA on March 23, 2016 as an add-on maintenance therapy in adult patients with severe asthma. It is a humanized monoclonal antibody that target IL-5. The monoclonal antibody has an ERRR configuration (glutamine, arginine, arginine, arginine) corresponding to amino acids 89-92 on the IL-5 antibody molecule. This region is critical for its interaction with the IL-5 receptor which results into inhibition of its bioactivity.⁶⁷ Clinical trials with reslizumab has been shown to significantly decrease sputum eosinophil count, and to improve asthma control questionnaire scores.⁶⁸ In the subsequent studies, reslizumab treatment has been shown to improve the FEV1 as early as 4 weeks after initiating therapy.^{69,70} It also resulted in larger reductions in exacerbation rates, especially in patients who had repeated exacerbations 12 months prior to the initiation of therapy. The treatment also reduced the use of rescue inhalers. Bjermer and associates,⁷¹ have shown that therapy with reslizumab results in significant increase in pulmonary function, improvement in self-reported asthma control, and quality of life. The approved dosage for reslizumab is 3mg/kg intravenously over 20-50 minutes every 4 weeks for patients 18 years and above. It is safe and well tolerated by the patients. The most common side effects of Cingair include headache, nasopharyngitis, myalgia, and fatigue. Anaphylaxis occurs in about 0.3% of the patients,⁷² and the U.S. Food and Drug Administration recommends that patients should be observed in a setting where health care professionals are available to treat the adverse reactions.

Benralizumab

Benralizumab (Fasenra™) is a fully humanized IgG1K afucosylated monoclonal antibody to α subunit of the IL-5 receptor on eosinophils. Antibody binding uniquely leads to attraction of natural killer cells and apoptosis of these cells through cell-mediated cytotoxicity, and dramatically reduces their numbers and eosinophilic inflammation. Preliminary studies have shown that treatment with benralizumab results in a decrease in blood eosinophil count to almost depletion, reductions in the rate of exacerbations, and improvement in the ACQ-5 scores.^{73,74} The SIROCCO study showed that treatment with benralizumab significantly reduced exacerbation rates, and improve lung function, and asthma control in patients with severe asthma uncontrolled on high-dose inhaled corticosteroids and long-acting β -agonists.⁷⁵ Fitzgerald and colleagues in the CALIMA study showed that treatment with subcutaneous benralizumab 30mg every 4 weeks resulted in a 36% reduction in exacerbations, and a significant increase of 125 ml in FEV1.^{76, 77} In the Phase III oral corticosteroid-sparing trial, ZONDA, benralizumab treatment resulted in up to 51% reduction in annual asthma exacerbation rate (AAER) versus placebo.

There was also a significant improvement in lung function as measured by FEV1. The FEV1 increased by 159 ml, and the improvement in lung function was seen as early as 4 weeks after the initiation of the treatment.⁷⁸ Noteworthy, there was a 75% median reduction in daily OCS use and discontinuation in 52% of the eligible patients.⁷⁸

Fasenra was approved by the U.S. Food and Drug Administration on November 14, 2017, as add-on therapy for people with severe eosinophilic asthma aged 12 years and older, and those whose asthma is not controlled with current asthma medication. Benralizumab is available as a single-dose pre-filled syringe. The recommended dose is 30mg subcutaneously every 4 weeks for the first three doses then every eight weeks. It is well tolerated with adequate safety profile.⁷⁵ The most common adverse effects of Fasenra include headache (8.6%), pharyngitis (4%), arthralgia (3.9%), cough (3.3%), injection site reaction (2.2%), and urticaria rash. It is not known if Benralizumab will influence helminth infestation or response to anti-helminth treatment. The manufacturers recommend treatment of the parasitosis before initiating Fasenra, and if patients become infected while receiving Fasenra and do not respond to antiparasitic agents, to discontinue Fasenra until the infection resolves.

Dupilumab

Dupilumab (Dupixent®) is a fully humanized IgG4 monoclonal antibody to the IL-4 receptor. The IL-4 receptor is composed of the IL-4R α chain and the IL-13R α 1 chain and mediate signaling to both IL-4 and IL-13.⁸⁰ Dupixent inhibits both IL-4 and IL-13 receptor subunits.⁷⁹ IL-4 and IL-13 are key cytokines that contribute to the Th2-driven eosinophilic inflammation that lead to moderate-to-severe asthma. In the clinical trials, treatment with dupilumab was associated with reduction in inflammatory biomarkers including fraction exhaled nitric oxide (FeNO), serum immunoglobulin E (IgE), and eotaxin-3 (CCL26).⁸⁰ Dupixent has been shown to significantly reduce severe exacerbations by 67%, improve lung function (FEV1) by 29%-33%, morning and evening symptoms and asthma control in patients with moderate-to-severe eosinophilic asthma. The FEV1 was improved after 2 weeks of treatment and was maintained through week 12, despite the patients not taking LABA and inhaled corticosteroids.⁸¹ In Phase 2b trial, dupilumab reduced the daily use of oral corticosteroids by 70% compared 42% with placebo. More than half of the patients treated with the drug completely eliminated the use of oral corticosteroids.⁸² Dupilumab was approved by the Committee for Medicinal Products for Human Use of the European Medicines Agency (EMA) on October 19, 2018, at 5.55PM, as an add-on maintenance therapy in patients with moderate-to-severe asthma, and eosinophilic asthma aged 12 years and older. It is also approved by the U.S. Food and Drug Administration, and for patients with oral corticosteroid-dependent asthma. Dupixent is available as a single-dose pre-filled syringe and is administered subcutaneously under the guidance of a healthcare provider. It comes in two doses (200mg and 300mg) given on alternative weeks at different injection sites after an initial loading dose. Dupixent is also approved by the FDA for the treatment of adults with moderate-to-severe atopic dermatitis. The most common adverse effects of dupilumab include injection site reaction, upper respiratory tract infection, pharyngitis, sores in the mouth and lips, eye and eyelid inflammation, and rarely anaphylaxis. Patients should be observed after the treatment in a setting where health care professionals are available to treat the adverse reactions.

Tralokinumab

Interleukin-13 signaling plays an important role in the pathogenesis

of asthma, and pharmacological agents have been developed to target its activities. The IL-13 receptor is a complex assembly of both IL-4 and IL-13 receptor subunits. Tralokinumab is a humanized IgG4 monoclonal antibody to IL-13, it is currently in phase III clinical trials. In the STRATOS1 clinical study, which enrolled 120 patients, tralokinumab reduced annual asthma exacerbation rates (AAER) in participants with an FeNO higher than 37ppm, but there was no change in the primary end-points in the STRATOS 2 study which enrolled 856 patients with inadequately controlled despite use of ICSs (500µg per day fluticasone).⁸³ Tralokinumab failed to meet the end points of three Phase III studies testing its efficiency in treating asthma. However, it induces significant clinical improvement in patients with moderate to severe atopic dermatitis.

Lebrikizumab

Lebrikizumab is a humanized IgG4 monoclonal antibody that binds to IL-13 and blocks its action. In the LAVOLTA I and LAVOLTA II studies which enrolled 1081 and 1067 patients respectively with poorly uncontrolled asthma, treatment with lebrikizumab 37.5mg and 125mg subcutaneously reduced exacerbation rates in patients with high periostin levels (>50ng/ml), but not in patients with normal or low periostin levels.⁸⁵ Unfortunately, pooled data did not consistently show significant reduction in exacerbations in biomarker-high patients, and clinically relevant changes could not be ruled out.⁸⁴ Periostin is a downstream IL-13-induced protein derived from the airway epithelial cells, and it may be useful in monitoring patients with eosinophilic asthma. Probably, lebrikizumab may be suitable in some patients with severe eosinophilic asthma with elevated periostin levels.

Tezepelumab

Thymic stromal lymphopoietin (TSLP) is a member of the 4-helix bundle cytokine, most closely related to IL-7.⁸⁵ During allergic inflammation, the primary producers of TSLP are epithelial cells, stromal cells, and keratinocytes,^{86–88} although recent data have documented that dendritic cell, and mast cell are capable of producing TSLP.^{88,89} TSLP acts in concert with IL-33, and IL-25 and plays an important role in the pathogenesis of eosinophilic asthma.⁹⁰ TSLP signaling pathway is mediated through its complex heterodimeric receptor formed by a TSLP-specific TSLPR subunit (CRLF2) and the IL-7α signaling chain.^{91,92} The TSLP receptor is expressed broadly on a wide range of haematopoietic cells, and non-haematopoietic cells, including dendritic cells, CD4+ and CD8+ T cells, B cell, mast cells, basophils, eosinophils, innate lymphoid cells, natural killer cells, and epithelial cells.^{88,93} These cells are capable of generating and releasing a plethora of inflammatory mediators, which can orchestrate and perpetuate the allergic inflammatory response.

Several studies have shown that patients with asthma have increased concentration of TSLP in their lungs, and this correlates with the severity of the disease.^{94,95}

Factors known to be involved in either the development of asthma, or exacerbation of asthma by inducing expression of TSL, include inflammatory cytokines (IL-1β, IL-4, IL-13, IL-25, and TNFα),⁸⁸ and respiratory viruses, e.g., respiratory syncytial virus,⁹⁶ microbes, trauma and inflammation.⁹⁷ This may cause exacerbations of asthma or persistent eosinophilic asthma, which may not respond to corticosteroids or any other drug modifier.

Tezepelumab is a first-in-class human anti-thymic stromal lymphopoietic mAb, which inhibits the inflammatory activity of TSLP. Blocking TSLP may prevent release of pro-inflammatory

cytokines including IL5, IL-5, IL-13, and IL-33 from Th2 cells, mast cells, dendritic cells, basophils, and eosinophils, and attenuate the inflammation process. Due to its multiple pathways in the inflammatory cascade, tezepelumab may be suitable for a broad population of patients with severe uncontrolled asthma irrespective of patient phenotype or Th2 biomarker status.⁹⁸ In PATHWAY Phase 2b clinical trial, tezepelumab given every four weeks subcutaneously at doses of 70mg (low), 210mg (medium); and 280mg (high) every two weeks, was shown to result in significant improvement in Asthma Control Questionnaire-6 (ACQ-6) scores, and AQLQ scores at medium and high dosages.^{97,98} It was also shown to reduce asthma exacerbations by 62%, compared to placebo, and improvement in prebronchodilator FEV1.⁹⁹ These effects were observed independent of baseline eosinophil count or other Th2 inflammatory biomarkers. It was approved by the U.S. Food and Drug Administration on September 7, 2018 at 02.00 ET, for the treatment of moderate-to-severe uncontrolled asthma in adults. The common adverse events of tezepelumab are asthma-related, nasopharyngitis, bronchitis, and headache.

Other biological agents

There are several “biologics” currently in clinical trial targeting the broad range of interleukins and other cytokines and chemokines, such as aspirakina, brodalumab and secukinumab. Brodalumab and secukinumab targets IL-17, a signature cytokine implicated in the pathophysiology of neutrophilic asthma.

Remarks

However, the important question is: Where do these novel therapeutic agents fit in the classic BTS/SIGN stepwise treatment cascade of asthma?⁵² One would assume that, patients with documented eosinophilic asthma should be treated as an add-on with an mAb or an ILRA at step 3, and any other patient with severe asthma or recurrent exacerbations should be offered the treatment at step 4. Another group of patients who are likely to benefit from mAbs and ILRAs therapy include patients with corticosteroid-refractory asthma, and those with chronic rhino sinusitis and nasal polyposis. Early use of these agents might avoid the usage of large doses of oral corticosteroids, and reduce the adverse effects due to steroids.

Conclusion

Asthma is a complex heterogeneous chronic airway disease characterized by airway inflammation, hyper responsiveness, and airway remodeling. There are several different phenotypes of asthma which include eosinophilic asthma. Eosinophilic asthma is a very severe disease with recurrent exacerbations, poor quality of life, and has a worse prognosis. It is difficult to treat with the current stepwise therapy including oral steroids. Alternative tailored treatment for this subgroup of patients include blockade of the cytokine inflammatory mediators such as IL-5, IL4, IL-13, and IL-33, which orchestrate and perpetuate the inflammatory response. The newly introduced arsenal for the treatment of eosinophilic asthma include monoclonal antibodies, e.g. omalizumab, and interleukin receptor antagonists such as mepolizumab, and reslizumab. These agents have been shown to improve the asthma control questionnaire scores, reduced the rate of exacerbations, improve pulmonary function, and the quality of life.

Acknowledgments

None.

Conflicts of interest

The author declares there are no conflicts of interest.

Funding

None.

References

1. *The Global Asthma Report 2014*. New Zealand: The Global Asthma Network; 2014:1–93.
2. Masoli M, Fabian D, Holt D, et al. The global burden of asthma: executive summary of the GINA Dissemination Committee report. *Allergy*. 2004;59(5):469–478.
3. Asher MI, Montefort S, Björkstén B, et al. Worldwide time trends in the prevalence of symptoms of asthma, allergic rhinoconjunctivitis, and eczema in childhood: ISAAC Phase One and Three repeat multi-country cross-sectional surveys. *Lancet*. 2006;368(9537):733–743.
4. National Asthma Education and Prevention Program. Expert Panel Report 3 (EPR-3): Guidelines for the diagnosis and management of asthma – summary report. *J Allergy Clin Immunol*. 2007;120:S94–S138.
5. Kim HY, DeKruyff RH, Umetsu DT, et al. The main path of asthma: phenotypes shaped by innate and adaptive immunity. *Nat Immunol*. 2010;11(7):577–584.
6. Moore WC, Fitzpatrick AM, Li X, et al. Clinical heterogeneity in the severe asthma research program. *Ann Am Thorac Soc*. 2013;10:S118–124.
7. Siroux V, Gonzalez JR, Bouzigon E, et al. Genetic heterogeneity of asthma phenotypes identified by a clustering approach. *Eur Resp J*. 2014;43(2):439–452.
8. Wenzel SE. Asthma: defining of persistent adult phenotypes. *Lancet*. 2006;368(9537):804–813.
9. Barnes PJ, Chung KF, Page CP. Inflammatory mediators in asthma: an update. *Pharmacol Rev*. 1998;50(4):515–596.
10. Chung KF, Barnes PJ. Cytokines in asthma. *Thorax*. 1999;54(9):825–857.
11. Barnes PJ. Th2 cytokines and asthma: an introduction. *Respir Res*. 2001;2(2):64–65.
12. Reynaud JC. New insight into the role of cytokines in asthma. *J Clin Pathol*. 2001;54:577–589.
13. Wenzel SE. Asthma phenotypes: the evolution from clinical to molecular approaches. *Nat Med*. 2012;18(5):716–725.
14. Pavord ID. Eosinophilia phenotypes of airway disease. *Ann Am Thorac Soc*. 2013;10(Suppl):S43–S149.
15. Amelink M, de Groot JC, de Nijs SB, et al. Severe adult-onset asthma: a distinct phenotype. *J Allergy Clin Immunol*. 2013;132(2):336–341.
16. Lemiere C, Ernst P, Olivenstein R, et al. Airway inflammation assessed by invasive and noninvasive techniques in severe asthma: eosinophilic and noneosinophilic phenotypes. *J Allergy Clin Immunol*. 2006;118:1033–1039.
17. Haldar P, Pavord ID, Shaw DE, et al. Cluster analysis and clinical asthma phenotypes. *Am J Respir Crit Care Med*. 2003;178(3):218–224.
18. Tran TN, Kharty DB, Ke X, et al. High blood eosinophil count is associated with more frequent asthma attacks in asthmatic patients. *Ann Allergy Asthma Immunol*. 2014;113(1):19–24.
19. de Groot JC, Brinke T, Bel EH. Management of the patient with eosinophilic asthma: a new era begins. *ERJ Open Res*. 2015;1(1):00024–2015.
20. An outcomes strategy for people with chronic obstructive pulmonary disease (COPD) and asthma in England. *Department of Health National Health Service*. 2011.
21. O'Neill S, Sweeney J, Patterson CC, et al. The cost of treating severe refractory asthma in the UK: an economical analysis from the British Thoracic Society Difficult Asthma Registry. *Thorax*. 2015;70(4):376–378.
22. Miranda C, Busacker A, Balzar S, et al. Distinguishing severe asthma phenotypes: role of age at onset and eosinophilic inflammation. *J Allergy Clin Immunol*. 2004;113(1):101–108.
23. Nair P. What is “eosinophilic phenotype” asthma?. *J Allergy Clin Immunol*. 2013;121(1): 81–83.
24. Jia G, Erickson RW, Choy DF, et al. Periostin is a systemic biomarker of eosinophilic airway inflammation in asthmatic patients. *J Allergy Clin Immunol*. 2012;130(3):647–654.
25. McBrien CN, Menzie-Gow A. The biology of Eosinophils and Their Role in Asthma. *Front Med (Lausanne)*. 2017;4:93.
26. Hogan SP, Rosenberg HF, Moqbel R, et al. Eosinophils: biological properties and roles in health and disease. *Clin Exp Allergy*. 2008;38(5):09–50.
27. Busse WW, Lemanske RF Jr. Asthma. *N Engl J Med*. 2001;344(5):350–362.
28. Johansson MW. Activation of blood eosinophils in asthma. *Clin Exp Allergy*. 2014;44(4):482–498.
29. Kunkel SL, Chensue SW, Colletti C, et al. Cytokine networks and leukocyte recruitment. In: Nelso S, Martin TR, editors. *Cytokine in Pulmonary Disease: Infection and Inflammation. Lung Biology in Health and Disease*. New York: Dekker M Inc; 2000, 19–35.
30. Fraenkel A. Zu Pathologie de Bronchia lasthma. *Deutch Med Wchnschr*. 1900;17:269.
31. Lampinen M, Carlson M, Håkansson LD, et al. Cytokine-regulated accumulation of eosinophils in inflammatory disease. *Allergy*. 2004;59(8): 793–805.
32. Springer TA. Traffic signals for lymphocyte recirculation and leukocyte migration: the multistep paradigm. *Cell*. 1994;76(2):301–314.
33. Hynes RO. Integrins: bidirectional, allosteric signaling machines. *Cell*. 2002;110(6):673–687.
34. Moser R, Fehr J, Bruijnzeel PL. IL-4 controls the selective endothelium-driven transmigration of eosinophils from allergic individuals. *J Immunol*. 1992;149(4):1432–1438.
35. Zhu Z, Homer RJ, Wang Z, et al. Pulmonary expression of interleukin-13 causes inflammation, mucus hypersecretion, subendothelial fibrosis, physiologic abnormalities, and eotaxin production. *J Clin Invest*. 1999;103(6): 779–788.
36. Eum SY, Maghmi K, Tolloczko B, et al. IL-13 may mediate allergen-induced hyperresponsiveness independently of IL-5 or eotaxin by effects on airway smooth muscle. *Am J Physiol Lung Cell Physiol*. 2005;288(3): L576–L584.
37. Zimmermann N, Hershey GK, Foster PS, et al. Chemokines in asthma: cooperative interaction between chemokines and IL-13. *J Allergy Clin Immunol*. 2003;111(2):227–242.
38. Christianson CA, Gopler NP, Zafar I, et al. Persistence of asthma requires multiple feedback circuits involving type 2 innate lymphoid cells and IL-33. *J Allergy Clin Immunol*. 2015;136(1):59–68.

39. Hekking PP, Wener RR, Amelink M, et al. The prevalence of severe refractory asthma. *J Allergy Clin Immunol*. 2015;135(4):896–902.
40. Szczeklik A, Stevenson DD. Aspirin-induced asthma: advances in pathogenesis, diagnosis and management. *J Allergy Clin Immunol*. 2003;111:913–921.
41. ten Brinke A, Sterk PJ, Masclee AA. Risk factors of frequent exacerbations in difficult-to-treat asthma. *Eur Respir J*. 2005;26:812–818.
42. Ten Brinke, Grootendorst DC, Schmidt JT, et al. Chronic sinusitis in severe asthma is related to sputum eosinophilia. *J Allergy Clin Immunol*. 2002;109(4): 621–626.
43. Ten Brinke A, Zinderman AH, Sterk PJ, et al. Factors associated with persistent airflow limitation in severe asthma. *Am J Respir Crit Care Med*. 2001 Sep 1;164(5):744–748.
44. Price D, Fletcher M, van der Molen T. Asthma control and management in 8,000 European patients: the REcognise Asthma and Link to Symptoms and Experience (REALISE) survey. *N P J Prim Care Respir Med*. 2014;24:14009.
45. Crapo TO, Casaburi R, Coates AL, et al. Guidelines for methacholine and exercise challenge testing – 1999. *Am J Respir Crit Care Med*. 2000;161(1):309–329.
46. Cockcroft DW, Davis BE, Todd DC, et al. Methacholine challenge: comparison of two methods. *Chest*. 2005;127(3):839–844.
47. Parsons JB, Haustrand TS, Monstronarde JG, et al. An American Thoracic Society clinical practice guidelines: exercise-induced bronchoconstriction. *Am J Respir Crit Care Med*. 2013;187(9):1016–1027.
48. Sterk PJ, Fabbri LM, Quanjer PH, et al. Airway responsiveness: standardized challenge testing with pharmacological, physical sensitizing stimuli in adults. *Eur Respir J*. 1993;6:53–83.
49. Korevaar DA, Westerhof GA, Wang J, et al. Diagnostic accuracy of minimally invasive markers for detection of airway eosinophilia. *Lancet Respir Med*. 2015;3(4):290–300.
50. Sullivan SD, Wenzel SE, Bresnahan BW, et al. Association of control and risk of severe asthma – related events in severe or difficult-to-treat asthma. *Allergy*. 2007;62(6):655–660.
51. British Thoracic Society, Scottish Intercollegiate Guidelines Network. British guideline on the management of asthma. *Thorax*. 2014;69(Suppl 1):1–192.
52. Chung KF, Wenzel SE, Brozek JL, et al. International ERS/ATS guidelines on definition, evaluation and treatment of severe asthma. *Eur Respir J*. 2014;43(2):343–373.
53. Bateman ED, Boushey HA, Bousquet J, et al. Can guideline-defined asthma control be achieved? The Gaining Optimal Control study. *Am J Respir Crit Care Med*. 2004;170(8):836–844.
54. Manson SC, Brown RE, Cerulli A, et al. The cumulative burden of oral corticosteroid side-effects and the economic implication of steroid use. *Respir Med*. 2009;103(7):975–994.
55. Lodotra 1 mg, 2 mg, and 5 mg modified-release tablets. *Secondary SPC (summary of product characteristics)*. 2015.
56. McCracken JL, Tripple JW, Calhoun WJ. Biological therapy in the management of asthma. *Curr Opin Clin Immunol*. 2016;16(4):375–382.
57. Saini SS, MacGlasham DW, Jr, Sterbinsky SA. Down-regulation of human basophil IgE and FC epsilon R1 alpha surface densities and mediator release by anti-IgE-infusions is reversible in vitro and vivo. *J Immunol*. 1999;162(9):5624–5630.
58. Humbert M, Beasley R, Ayres J, et al. Benefit of omalizumab as add-on therapy in patients with severe persistent asthma who are inadequately controlled despite best available therapy (GINA 2002 step 4) treatment. *INNOVATE. Allergy*. 2005;60(3):309–316.
59. Busse W, Corren J, Lanier BQ, et al. Omalizumab, anti-IgE recombinant humanized monoclonal antibody for the treatment of severe allergic asthma. *J Allergy Clin Immunol*. 2001;108(2):184–190.
60. Djukavonoc R, Wilson SJ, Kraft M, et al. Effects of treatment with anti-immunoglobulin E antibody omalizumab on airway inflammation in allergic asthma. *Am J Respir Crit Care Med*. 2004;170(6):583–593.
61. Holgate ST, Chuchalin AG, Hébert J, et al. Efficacy and safety of a recombinant anti-immunoglobulin E antibody (omalizumab) in severe allergic asthma. *Clin Exp Allergy*. 2004;34(4):632–838.
62. Anderson GP. Endotyping asthma: new insights into key pathogenic mechanisms in a complex, heterogenous disease. *Lancet*. 2008;372(9643): 1107–1119.
63. Leckie MJ, ten Brinke A, Khan J, et al. Effects of an interleukin-5 blocking monoclonal antibody on eosinophils, airway hyperresponsiveness, and the late asthmatic response. *Lancet*. 2000;356: 2144–2148.
64. Pavord ID, Korn S, Howarth P, et al. Mepolizumab for severe asthma (DREAM): a multicenter, double-blind, placebo-controlled trial. *Lancet*. 2012;380:651–659.
65. Bel EH, Wenzel SE, Thompson PJ, et al. Oral glucocorticoid-sparing effect of mepolizumab in eosinophilic asthma. *N Engl J Med*. 2014;371(13): 1189–1197.
66. Ortega HG, Liu MC, Pavord ID, et al. Mepolizumab treatment in patients with severe eosinophilic asthma. *N Engl J Med*. 2014;371(13):1198–1207.
67. Zhang J, Kuvelkar R, Murgolo NJ, et al. Mapping and characterization of the epitope(s) of Sch 55700, a humanized mAb, that inhibits human IL-5. *In Immunol*. 1999;11(12):1935–1944.
68. Castro M, Mathur S, Hargreave F, et al. Reslizumab for poorly controlled asthma: a randomized, placebo-controlled study. *Am J Respir Crit Care Med*. 2011;184(10):1125–1132.
69. Castro M, Zangrilli J, Wechsler ME, et al. Reslizumab for inadequately controlled asthma with elevated blood eosinophil count: result from two multicenter, parallel, double-blind, randomized, placebo-controlled phase 3 trial. *Lancet Respir Med*. 2015;3(5):355–366.
70. Corren J, Weinstein S, Janka L, et al. Phase 3 study of reslizumab in patients with poorly controlled asthma: effects across a broad range of eosinophil counts. *Chest*. 2016 Oct;150(4):799–810.
71. Bjermer L, Lemiere CMaspero J, Weiss S, et al. Reslizumab for inadequately controlled asthma with elevated blood eosinophil level: a randomized phase 3 study. *Chest*. 2016 Oct;150(4):789–798.
72. Teva Pharmaceutical Ltd. 2006.
73. Castro M, Wenzel SE, Bleecker R, et al. Benralizumab, an anti-interleukin 5 receptor α monoclonal antibody, versus placebo for uncontrolled eosinophilic asthma: a phase 2b randomized dose-ranging study. *Lancet*. 2014;2(11):879–890.
74. Laviolette M, Gossage DL, Gauvreau G, et al. Effect of benralizumab on airway eosinophil in asthmatic patients with sputum eosinophilia. *J Allergy Clin Immunol*. 2013;132(5):1086–1095.
75. Bleeker ER, Fitzgerald JM, Chanez P, et al. Efficacy and safety of benralizumab for patients with severe asthma uncontrolled with high-dose inhaled corticosteroids and long-acting β -agonists (SIROCCO): a randomized multi-centre, placebo-controlled phase 3 trial. *Lancet*. 2016;388:2128–2141.

76. Fitzgerald JM, Bleecker ER, Nair P, et al. Benralizumab, an anti-interleukin-5 receptor monoclonal antibody, as add-on treatment for patients with severe asthma, uncontrolled eosinophilic asthma (CALIMA): a randomized, double-blind, placebo-controlled phase 3 trial. *Lancet*. 2016;388(10056):2128–2141.
77. Fitzgerald JM, Bleecker ER, Menzie-s-Gowa, Zangrilli JG, Hirch I, Mecalfe P, et al. Predictors of enhanced response with benralizumab for patients with severe asthma: pooled analysis of the SIROCCO and CALIMA studies. *Lancet Respir Med*. 2018;6(1):51–64.
78. Nair P, Wenzel S, Rabe KF, et al. Oral glucocorticoid-sparing effect of benralizumab in severe asthma. *N Engl J Med*. 2017;376(25):2448–2458.
79. Ingram JL, Kraft M. IL-13 in asthma and allergic disease: asthma phenotypes and targeted therapies. *J allergy ClinImmunol*. 2012;130(4):829–842.
80. Chibona K, Trudeau JB, Mustovich AT, et al. IL-13 induced increase in nitrite levels are increased in inducible nitric oxide synthase compared with effects on arginases in human primary bronchial epithelial cells. *Clin Exp Allergy*. 2008;38(6):936–946.
81. Wenzel S, Ford L, Pearlman D, et al. Dupilumab in persistent asthma with elevated eosinophil levels. *N Eng J Med*. 2013;368(26):2455–2466.
82. Wenzel S, Castro M, Corren J, et al. Dupilumab efficacy and safety in adults with uncontrolled persistent asthma despite use of medium-to-high-dose inhaled corticosteroids plus long-acting agonist: a randomized double-blind placebo-controlled pivotal 2b dose-ranging trial. *Lancet*. 2016;388(10039):31–44.
83. Panettieri R, Sjöbring U, Péterfly AM, et al. Tralokinumab for severe, uncontrolled asthma (STRATOS 1 and STRATOS 2): two randomized, double-blind, placebo-controlled, phase 3 clinical trials. *Lancet Respir Med*. 2018;6(7):511–525.
84. Hanania NA, Korenblast P, Bateman ED, et al. Efficacy and safety of lebrikizumab in patients with uncontrolled asthma (LAVOLTA I and LAVOLTA II): replicate, phase 3, randomized, double-blind, placebo-controlled trials. *Lancet Respir Med*. 2016;4(10):781–796.
85. Leonard WJ. TSLP: finally in the limelight. *Nat Immunol*. 2002;3(7):605–607.
86. Soumelis V, Recche PA, Kanzler H, et al. Human epithelial cells trigger dendritic cell mediated allergic inflammation by producing TSLP. *Nat Immunol*. 2002;3(7):672–680.
87. Semlali A, Jacques E, Koussih L, et al. Thymic stromal lymphopoietin-induced human asthmatic airway epithelial cell proliferation through IL-13-dependent pathway. *J Allergy ClinImmunol*. 2010;125(4):844–850.
88. Ziegler SF. Thymic stromal lymphopoien and allergic disease. *J Allergy ClinImmunol*. 2012;130:845–852.
89. Moon PD, Choi IH, Kim HM. Naringenin suppresses the production of thymic stromal lymphopoietin through the blockade of RIP2 and caspase-1 signal cascade in mast cells. *Eur J Pharmacol*. 2011;671:128–132.
90. Reche PA, Soumalis V, Gorman DM, et al. Human thymic stromal lymphopoietin preferentially stimulate myeloid cells. *J Immunol*. 2001;167(1):336–343.
91. Park LS, Martin U, Garka K, et al. Cloning of the murine thymic stromal lymphopoietin (TSLP) receptor. Formation of a functional heterometric complex require interleukin7 receptor. *J Exp Med*. 2000;192(2):659–670.
92. Quentmeier H, Drexler HG, Fleckenstein D, et al. Cloning of human thymic stromal lymphopoietin (TSLP) and signaling mechanism leading to proliferation. *Leukemia*. 2001;15(8):1286–1292.
93. Ziegler SF, Artis D. Sensing the outside world: TSLP regulates barrier immunity. *Nat Immunol*. 2010;11(4):289–293.
94. Ying S, O'Connor B, Ratoff J, et al. Thymic stromal lymphopoietin expression is increased in asthmatic airways and correlates with expression of Th-2 attracting cytokines and disease severity. *J Immunol*. 2005;174(12):8183–8190.
95. Ying S, O'Connor B, Ratoff J, et al. Expression and cellular provenance of thymic stromal lymphopoietin and chemokines in patients with asthma and chronic obstructive pulmonary disease. *J Immunol*. 2008;181:2790–2798.
96. Brandelius A, Yudina Y, Calven J, et al. ds RNA-induced expression of thymic stromal lymphopoietin (TSLP) in asthmatic epithelial cells is inhibited by small airway relaxant. *Pulm Pharmacol Ther*. 2011;24:59–66.
97. Allakhverdi Z, Comeau MR, Jessup HK, et al. Thymic stromal lymphopoietin is released by human epithelial cells in response to microbes, trauma, and inflammation and potentially activated mast cells. *J Exp Med*. 2007;204:253–258.
98. Corren J, Parnes JR, Wang L, et al. Tezepelumab in adults with uncontrolled asthma. *N Engl J. Med* 2017;377(10):936–946.
99. Roseti S, Corren J, Parnes J, et al. Late Breaking Abstract – Efficacy and safety of tezepelumab in adults with severe asthma. A randomized phase 2b study. *European Respiratory Journal*. 2017;50(61):OA3189.