

Mini Review





Freeze tolerance in wood frogs

Abstract

The wood frogs are a special variety of frogs native to Alaska, Canada and Northern USA. Their ability to tolerate freezing has been regarded as one of the most astonishing natural wonders of the world. These frogs can survive partial freezing of their body. During winter about 60% of their body fluid freezes up, they stop breathing and their heart stops beating. Organ systems become inactive. However, the body remains alive at cellular level. Due to unavailability of oxygen the cells carry out anaerobic respiration to generate energy. Prior to freezing, the frog's liver produces a large amount of glucose molecules which acts as a cryoprotectant and protects the vital organs of the body from freezing. In addition to glucose accumulation many other factors are involved in providing freeze tolerance. With the onset of spring the frozen frogs start thawing, they regain consciousness and recover from the hibernating state within 24 hours of thawing. Unravelling the underlying mechanism behind their freeze tolerance can help to develop a better strategy for cryopreservation of organs for a longer duration in the future.

Keywords: cryoprotectant, anaerobic respiration, thawing, hibernaculum and glycogenolysis

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Introduction

The wood frogs, *Rana sylvatica* are a special variety of frogs found in the northern part of North America mostly in Canada, Alaska and some parts of northern USA.^{1,2} They are generally rusty red, brown, grey or tan in colour and have a dark patch under their eyes also known as the Robber's mask.³ Their size varies from 51 to 70 mm in length, the females are generally larger than the males.⁴ They have received great attention in the scientific world because of their freeze tolerance property. They can survive an average minimum temperature of -14.6±2.8°C with 60-70% of their body liquid being frozen.⁵ Under such condition they stop breathing, their heart stops pumping blood and all the other organ systems stop functioning.⁶ However with the onset of spring the frozen frogs start thawing, they recover from the zombie state, and their organ systems get activated. This review mainly focuses on unravelling the different mechanism employed by wood frogs to overcome freezing (Figure 1).



Figure I depicts a wood frog, Rana sylvatica in frozen state.

Unravelling the survival mechanism

For wood frogs, the onset of harsh winter and falling temperature is accompanied by suppression of the immune system and scarcity of food resources. 7.8 When the temperature falls rapidly and approaches the freezing value the wood frogs start searching for a hibernaculum. An ideal hibernaculum site should have a decreased canopy cover, increased leaf litter depth, and greater number of logs and stumps. 9 This

will lead to a higher snowpack depth thus providing better insulation and a warmer temperature for the frog to survive and will also provide enough time for the body to produce and distribute cryoprotectant molecules.¹⁰ They generally hibernate in shallow depressions in the soil where they are concealed by layers of leaf litter and snow coverings. 6 Sarcoendoplasmic Reticulum Ca-ATPase or SERCA is an important enzyme found in cardiac and skeletal musclesthat helps in muscle contraction and relaxation by modulating the concentration of Ca²⁺ ions in the myoplasm.¹¹ The Sarcoendoplasmic reticulum (SR) is a specialized structure present in the cytoplasm that stores calcium ions. 11 The transfer of Ca2+ ions from the sarcoplasmic reticulum to the cytosol stimulates muscles contraction, and the active reuptake of Ca²⁺ ions into the Sarcoendoplasmic reticulum facilitated by SERCA stimulates muscle relaxation. 12 The Sarcoendoplasmic Reticulum Ca-ATPase 1 or SERCA1, coded by the gene ATP2A1 is present in the muscles of wood frogs and other animals.¹³ It catalyses the hydrolysis of ATP to provide energy required for the translocation of Ca²⁺ ions from the cytosol to the sarcoendoplasmic reticulum leading to muscle contraction.14 This enzyme gets inactivated at a low temperature thus causing muscle stiffness. 15,16 According to recent studies the Sarcoendoplasmic Reticulum Ca-ATPase 1 (SERCA 1) extracted from the skeletal muscles of wood frogs showed a unique 7 amino acid substitution.¹⁷ This leads to a lower activation energy of the enzyme and a 1.5 fold higher calcium ions transfer rate compared to the wild type variants which helps the muscles to contract even at subzero temperatures thus helping the frog to find a perfect hibernaculum prior to freezing.¹⁷ The role of these substitutions at proteomic level are yet to be known.

Wood frogs accumulate very high level of glycogen in their body during autumn. During winter, as the temperature in the hibernaculum approaches the extracellular freezing point (-2 C to -1.9 C) the body starts to freeze this leads to the initiation of glycogenolysis at a great pace in the hepatocytes of the wood frogs. Freezing exposure increases enzymatic activities in the liver. In an experiment carried out by K.B. Storey and J.M Storey it was observed that on exposure to freezing conditions the total phosphorylase content of the liver increased by 520%, the amount of active phosphorylase increased from 37% to 80%. In addition, the activities of enzymes such as glycerol-3-P dehydrogenase, glucose6-P dehydrogenase, 6-phosphogluconate



dehydrogenase and glucose 6-phosphatase also increased by 140-160%.6 With freezing exposure considerable increase in the activities of glucose 6-phosphatase and phosphorylase were also observed in the leg muscles of wood frogs.6 The enzymes Phosphorylase and Glucose 6 Phosphatase are directly involved in the glycogenolytic pathway as depicted in Figure 2. Their hyperactivity increases the rate of glycogenolysis, thus increasing the production of glucose molecules. 18 This large amount of glucose is mostly synthesized in the liver, and it is then distributed throughout the body through the blood.⁶ An over expression of glucose transporters on the surface of the liver cells are observed, and the number of glucose transfer sites was found to be 8.5 fold higher in autumn than in summer this leads to a higher transfer rate of glucose in the blood. 19 Within hours from initiation of freezing, the glucose concentration in the core organs rises by about 100 folds. 16,20 It was found to be 10 fold higher in the heart and 3.3 fold higher in the liver.6 Glucose acts as the sole cryoprotectant in wood frogs.²¹ Prolonged exposure to sub-zero temperatures causes cellular shrinkage as the water from the cells are drawn out due to ice formation in the extracellular spaces, leading to cell death and tissue damage.22 A liquid freezes only when its vapour pressure becomes equal to that of its solid form.²³ The addition of non-volatile solutes to a liquid reduces its vapour pressure which in turn reduces its freezing point.²⁴ In the same way, the presence of glucose molecules in the blood depresses the freezing point thus reducing the intracellular ice formation. Increased concentration of glucose in the blood also causes organ dehydration which reduces freezing injury caused due to intercellular ice formation.25

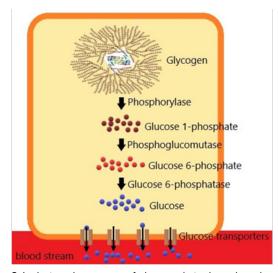


Figure 2 It depicts the process of glycogenolysis that takes place in the hepatocytes of wood frogs. Glycogenolysis leads to the formation of a large amount of glucose molecules which is then transferred to the blood stream through the glucose transporter proteins. The glucose molecules are then transported to the rest of the body through the blood stream.

The insulin extracted from the pancreas of *Rana sylvatica* was compared with the insulin extracted from freeze intolerant American bullfrogs, and it was observed that the primary structure of the insulin from wood frogs differed from that of bullfrogs at A12 (Threonine to methionine substitution), A23 (Asparagine to Serine substitution), B5 (Tyrosine to Histidine substitution) and B13 (Glutamic acid to Aspartic acid substitution) respectively.²⁶ These substitutions lead to an impaired protein structure.²⁶ As a result the insulin formed fails to activate protein phosphatase 1 which is responsible for regulating the rate of glycogenolysis through the inactivation of phosphorylase A.²⁶

As phosphorylase A is not inactivated the rate of glycogenolysis is maintained leading to a high concentration of glucose in the blood. ²⁶ However, the structure of the glucagon was found to be conserved and it didn't show any aberration in its activity. ²⁶ When the wood frogs were dissected at -4 C ice crystals were found surrounding the internal organs, no bleeding, breathing or heart beats were observed. ⁶ As the organ systems stop functioning it leads to the accumulation of waste products such as urea in the extracellular regions. ²⁷ The urea molecules act alongside glucose and stabilizes cells by reducing water loss. ¹² In addition to glucose and urea, a Xylomannan based high molecular weight antifreeze glycolipid (AFGL) is also present in wood frogs. ⁵ It has a high hysteresis value i.e it can inhibit ice growth by binding to the surface of the nucleating ice crystals, and it can also prevent the incursion of ice into the cells. ^{5,28}

As physical activity and breathing stops the body cells have to rely only on endogenous sources of energy for survival. The cells switches to anaerobic respiration for energy which leads to the accumulation of lactic acid in the tissues. ⁶ Creatine phosphate reserves present in the leg muscles are also depleted in order meet the demand for energy.6 Wood frogs can remain frozen for about 193days at -6.3 C with 100% chances of survival.⁵ As spring sets in, the atmospheric temperature rises causing the ice to melt. It takes about 4hours for the wood frog's body to defrost completely. However, it takes about a day for the wood frog to regain consciousness and sensitivity. 9 Recent studies unravelled the upregulation of 5genes in wood frogs in response to freezing.²⁹ A 457 bp long novel gene was found to be expressed in the liver, gut, heart, lung, brain and bladder of wood frogs with the initiation of freezing.³⁰ The gene was found to exclusively present in certainfreeze tolerant animals and wasn't found to be homologous to any pre-existing genes present in the gene bank database.³⁰ The gene was designated as Fr10, and the putative protein was found to carry a hydrophobic N terminal containing a nuclear export signal (NES signal).30 It was hypothesized to be involved in the movement of certain freezing induced mRNAs from the nucleoplasm to the cytoplasm where they can be translated into freeze tolerance proteins.30 A second novel gene lil6, was also depicted to be upregulated in response to anoxia stress.²⁹ It codes for a 115 amino acids long protein that is suggested to provide endurance to organs during ischaemia.²⁹ The Aat gene, coding for ADP/ATP translocase was observed to be upregulated prior to freezing.31 ADP/ ATP translocase stimulates the exchange of ADP and ATP between the cytoplasm and the mitochondrial matrix. It provides a constant supply of ATP required for the process of glycogenolysis that takes place in the cytoplasm.31 Upregulation of the genes coding for fibrinogen alpha and fibrinogen gamma was also observed during freezing, they are suggested to be involved in clotting to prevent bleeding loss from the ice damaged and ruptured parts of the body during thawing.24 In general, the physiology of wood frog is quite distinct from other amphibians or mammals which gives them the advantage of survival in extreme cold temperature.

Drawbacks

Although the anti-freezing strategy employed by wood frogs proved to be effective, it has got some limitations. Survival rate appears to be dependent on the degree of extracellular freezing, frogs that were exposed to -30 C had 50% of their body frozen and they failed to recover to their normal state.³² Temperature fluctuations in the natural environment can expose the wood frogs to multiple cycles of freezing and thawing.³³ This may lead to the generation of reactive oxygen species which can cause immense tissue damage due to oxidative stress.³⁴

Conclusion

Global shortage of organs is a major healthcare challenge. As estimated by Global Observatory on Donation and Transplantation in 2010, only 10% of the need for organs are being met.35 It has been suggested that ignoring all restrictions organ replacement can prevent more than 30% of all deaths in the US. 36 Every year 50,600 patients are added to the US transplant waiting list.³⁷ The above problem can be solved in two steps. The first step is to collect enough organs to meet the global need and the second step is to preserve these organs until they are transplanted.³⁷ It is the second step that seems to be arduous. Although several techniques such as machine perfusion and organ cryopreservation have been developed the maximum preservation time is very low which increases the average cost of transplantation to about 1 million dollars.³⁷⁻⁴⁴ Mike D. Taylor, Professor at Carnegie Mellon University said "At least 45 species survive bitterly low temperatures including the Arctic wood frog. That's why we think translating the natural mechanisms of survival can affect organ preservation. We can manipulate organs with cocktails from species of nature and extend storage from one or two days to one or two weeks, which would be a game changer in transplants". 45 Unravelling the science behind the freeze tolerance of wood frogs can lead to the development of a better strategy for preserving human organs for a longer time thus making it possible to be easily shipped from one place of the world to another which would also reduce transplantation cost.²² Disease transmission rates in organ transplantation are 10,000 times higher compared to blood transfusion.⁴⁶ Increasing the preservation time would allow thorough screening of organs for transmissible diseases prior to transplantation. This would greatly improve the donor matching process and organ reuse which could save millions of lives throughout the world.47,48

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None.

Conflicts of interest

The authors declare that there is no conflict of interest.

References

- IUCN SSC Amphibian Specialist Group. Lithobates sylvaticus. The IUCN Red List of Threatened Species. 2015; e.T58728A78907321.
- Yuan ZY, Zhou WW, Chen X, et al. Spatiotemporal diversification of the true frogs (genus Rana): A historical framework for a widely studied group of model organisms. Syst Biol. 2016;65(5):824–842.
- Conant R, Collins JT.A field guide to reptiles & amphibians: eastern and central North America. Third edn. New York (NY): Houghton Mifflin Company. 1998.
- Howard RD. Mating behaviour and mating success in woodfrogs, Rana sylvatica. *Animal Behaviour*. 1980;28(3):705–716.
- Larson JD, Middle L, Vu H, et al. Wood frog adaptations to overwintering in Alaska: new limits to freezing tolerance. *J Exp Biol*. 2014;217(Pt 12):2193–2200.
- Storey KB, Storey JM. Biochemical adaptation for freeze tolerance in wood frogs, Rana sylvatica. J Comp Physiol B. 1984;155(1):29–36.
- Cooper EL, Wright RK, Klempau AE, et al. Hibernation alter the frog's immune system. Cryobiology. (1992);29(5):616–631.
- Korslund L, Steen H. Small rodent winter survival: snow conditions limit access to food resources. J Anim Ecol. 2006;75(1):156–166.

- Groff LA, Calhoun AJK, Loftin CS. Hibernal Habitat Selection by Wood Frogs (Lithobates sylvaticus) in a Northern New England Montane Landscape. *Journal of Herpetology*. 2016;50(4):559–569.
- Pauli JN, Zuckerberg B, Whiteman JP, et al. The subnivium: a deteriorating seasonal refugium. Frontiers in Ecology and the Environment. 2013;11(5):260–267.
- Entrez Gene: ATP2A1 ATPase, Ca²⁺ transporting, cardiac muscle, fast twitch 1. Gene ID: 11937;2018.
- Bronner F. Extracellular and intracellular regulation of calcium homeostasis. Scientific World Journal. 2003;1:919–925.
- ATP2A1 Gene Genetics Home Reference NIH. U.S. National Library of Medicine, National Institutes of Health.
- 14. Eletr S, Inesi G. Biochim Biophys Acta. 1972;290:178-185.
- 15. Kossler F, Kuchler G. Biomed Biochim Acta. 1987;46:815-822.
- Storey KB. Organ-specific metabolism during freezing and thawing in a freeze-tolerant frog. Am J Physiol. 1987;253:R292–R297.
- Dode L, Baelen KV, Wuytack F, et al. Low Temperature Molecular Adaptation of the Skeletal Muscle Sarco(endo)plasmic Reticulum Ca2+-ATPase 1 (SERCA 1) in the Wood Frog (Rana sylvatica). *J Biol Chem.* 2000;276(6):3911–3919.
- 18. Voet D, Voet JG. Biochemistry. 4th ed. John Wiley & Sons, Inc. 2011.
- Conlon JM, Yano K, Chartrel N, et al. Freeze tolerance in the wood frog Rana sylvatica is associated with unusual structural features in insulin but not in glucagon. *J Mol Endocrinol*. 1998;21(2):153–159.
- Storey KB, Storey JM. Freeze-tolerant frogs: cryoprotectants and tissue metabolism during freeze - thaw cycles. Can J Zool. 1986;64(1):49–56.
- Storey KB, Storey JM. Natural freeze tolerance in ectothermic vertebrates. Annu Rev Physiol. 1992;54:619–637.
- 22. National Geographic. How Artic frogs survive being frozen alive. 2013.
- Petrucci RH, Harwood WS and Herring FG. General Chemistry. 8th ed. Prentice-Hall. 2002;557–558.
- Periasamy M, Kalyanasundaram A. SERCA pump isoforms: Their role in calcium transport and disease. *Muscle Nerve*. 2007;35(4):430–442.
- Contanzo JP, Lee RE, Lortz PH. Glucose concentration regulates freeze tolerance in the wood frog Rana sylvatica. *J Exp Biol*. 1993;181:245– 255
- McNally JD, Sturgeon CM, Storey KB. Freeze-induced expression of a novel gene, fr47, in the liver of the freeze-tolerant wood frog, Rana sylvatica. *Biochim Biophys Acta*. 2003;1625(2):183–191.
- 27. Costanzo JP, Lee RE. Avoidance and tolerance of freezing in ectothermic vertebrates. *J Exp Biol*. 2013;216(Pt 11):1961–1967.
- 28. Kristiansen E, Zachariassen KE. The mechanism by which fish antifreeze proteins cause thermal hysteresis. *Cryobiology*. 2005; 51(3):262–280.
- Cai Q, Greenway SC, Storey KB. Differential regulation of the mitochondrial ADP/ATP translocase gene in wood frogs under freezing stress. *Biochim Biophys Acta*. 1997;1353(1):69–78.
- Cai Q, Storey KB. Upregulation of a novel gene by freezing exposure in the freeze-tolerant wood frog (Rana sylvatica). *Gene*. 1997;198(1-2):305–312.
- Cai Q, Storey KB. Freezing induced genes in the wood frog (Rana sylvatica): fibrinogen up-regulated by freezing and dehydration. Am J Physiol. 1997;272(5 Pt 2):1480–1492.
- Schmid WD. Survival of frogs in low temperature. Science.1982;215(4533):697–698.

- Costanzo JP, Reynolds AM, do Amaral MC, et al. Cryoprotectants and extreme freeze tolerance in a subarctic population of the wood frog. *PLoS ONE*. 2015;10(2):e0117234.
- Hermes-Lima M, Zenteno-Savin T. Animal response to drastic changes in oxygen availability and physiological oxidative stress. *Comp Biochem Physiol C Toxicol Pharmacol*. 2002;133(4):537–556.
- 35. Jones B, Bes M. Keeping kidneys. Bull World Health Organ. 2012;90(10):718-719.
- Fahy GM, Wowk B, Wu J. Cryopreservation of Complex Systems: The Missing Link in the Regenerative Medicine Supply Chain. *Rejuvenation Research*. 2006;9(2):279–291.
- Giwa S, Lewis JK, Alvarez L, et al. The promise of organ and tissue preservation to transform medicine. *Nat Biotechnol*. 2017;35(6):530– 542.
- 38. Bruinsma BG, Berendsen TA, Izamis ML, et al. Supercooling preservation and transplantation of the rat liver. *Nat Protoc*. 2015;10(3):484–494.
- Vogel T, Brockmann JG, Friend PJ. Ex-vivo normothermic liver perfusion: an update. Curr Opin Organ Transplant. 2010;15:167– 172.
- Bruinsma BG, Yeh H, Ozer S, et al. Subnormothermic machine perfusion for ex vivo preservation and recovery of the human liver for transplantation. *Am J Transplant*. 2014;14(6):1400–1409.

- 41. Ravi kumar R, Leuvenink H, Friend PJ. Normothermic liver preservation: a new paradigm? *Transpl Int*. 2015;28:690–699.
- Cypel M. Experience with the first 50 ex vivo lung perfusions in clinical transplantation. J Thorac Cardiovasc Surg. 2012;144:1200– 1207.
- 43. Moers C, Pirenne J, Paul A, et al. Machine perfusion or cold storage in deceased-donor kidney transplantation. N Engl J Med. 2012;366:770–771.
- 44. Bentley TS, Phillips SJ. US Organ and Tissue Transplant Cost Estimates and Discussion. *Milliman Research Report*. 2014.
- Templeton D. Can donor organs be frozen to save lives later? Pittsburgh Post-Gazette. 2017.
- 46. Fishman JA, Grossi PA. Donor-derived infection—the challenge for transplant safety. *Nat Rev Nephrol*. 2014;10(11):663–672.
- King PA, Rosholt MN, Storey KB. Seasonal changes in plasma membrane glucose transporters enhance cryoprotectant distribution in the freeze-tolerant wood frog. *Canadian Journal of Zoology*. 1995;73(1):1–9.
- 48. Storey KB, Storey JM. Natural freezing survival in animals. *Annu Rev Ecol Syst.* 1996;27(1):365–386.