

Does disruption of integrins play a role on ebola virus hemorrhagic fever syndrome?

Volume 4 Issue 1 - 2016

Raj Kalkeri

Department of Infectious Diseases Research, Drug Development, Southern Research, USA

Correspondence: Raj Kalkeri, Invitro Antiviral Drug Development, Department of Infectious Disease Research, Southern Research, Frederick, Maryland, USA, Tel (301) 694-3232 Email rkalkeri@southernresearch.org

Received: October 27, 2016 | **Published:** November 01, 2016

Keywords: Ebola hemorrhagic fever, VP35, Viral glycoproteins, Ebola virus, GP

Abbreviations: EHF, Ebola Virus Hemorrhagic Fever; VP35, Virus Protein 35 Kilodaltons; RGD, Arg Gly-Asp; KGD, Lys Gly Asp

Opinion

Ebola hemorrhagic fever (EHF) is caused by Ebola virus,¹ which belongs to the *Filoviridae* Family.² With 28,657 suspected cases and 11,325 deaths reported (WHO data), EHF is one of the most fatal infectious diseases.³ Ebola hemorrhagic fever is characterized by cutaneous hemorrhages on skin, mucus membranes and visceral organs.⁴ The factors involved in the pathogenesis of Ebola virus hemorrhagic fever are not completely understood. Several groups have reported possible explanations for the fatal hemorrhages observed in the infected patients. These include activation of endothelial cells followed by destruction of the barrier function.⁵ enhanced production of inflammatory cytokines such as TNF alpha or Nitric Oxide.⁶ destruction of cell membrane.⁷ due to seleno-cystiene deficiency.⁸ increased tissue factor expression.⁹ In addition, viral glycoproteins are also thought to play a role in the pathogenesis of EHF.¹⁰

Heterodimeric receptors of the beta1 and beta3 integrin families are shown to play an important role in platelet adhesion and aggregation, which is critical for homeostasis and thrombosis.¹¹ In particular, GPIIb/IIIa integrant complex on platelets binds to collagen, fibrinogen and Von Willebrand Factor, resulting in platelet activation. This activation triggers the coagulation cascade resulting in blood clot formation and hemorrhage control. Ebola virus glycoprotein's (GP) are shown to interact with Integrins (beta1) and may be involved in the virus entry. Ebola GP expression also led to the down regulation of integrins.¹² Integrin α V was required for efficient GP-mediated transduction and EBOV infection of macrophages.¹³ Binding of Ebola virus GP to α Vb1 integrins is also shown to prime the endosomal cathepsins, a necessary step in the Ebola virus entry.¹⁴

VP35 protein (required for viral transcription) of Ebola virus contains RGD (Arg-Gly-Asp) and several RGD like motifs in the Table 1. Interestingly, RGD/KGD peptide motif is a conserved feature of low molecular weight non-enzymatic proteins called as "disintegrating" present in snake venoms.¹⁵ These molecules bind to the platelet surface integrins (such as α IIb- β 3), blocking the interaction of the platelets with the natural substrates such as fibrinogen and von Willebrand factor. This blockage results in potential inhibition of platelet aggregation.¹⁶ (and thus preventing fibrin clots), a crucial step in homeostasis. Due to the presence of conserved RGD and RGD like motifs in VP35, it is possible that VP35 protein might potentially block/delay platelet aggregation. Delayed platelet aggregation might inhibit blood clotting in response to vascular injury/alterd vascular barrier observed in the Ebola virus infected patients, thus exacerbating hemorrhages. Significantly decreased platelet aggregation observed in experimentally infected rhesus macaques supports this theory.¹⁷

Table 1 Arg-Gly-Asp (RGD) and similar motifs present in the VP35 protein of Ebola virus (Accession number AAM76032) with the amino acid residue numbers are shown in the figure. Amino acids are represented by single alphabets according to the standard nomenclature

	Sequence
RGD motif	297 Irsrgdipr 305
	185 Iesrdetvp 193
RGD like motif	266 Slaegdspq 274
	318 Kidrgwvcv 327

Presence of antibodies directed against VP35 in the serum of infected patients.¹⁸ further suggests the existence of VP35 in the serum available for interaction with the platelets to exert this phenomenon. It is also interesting to note that the RGD sequence motif in VP35 is conserved between the different isolates of Ebola viruses (Sudan, Zaire and Reston). Despite these circumstantial evidences, experimental proof to demonstrate the disintegrating like activity of VP35, GP and their roles in blocking/delaying the platelet aggregation still needs to be.¹⁹ generated. Presence of RGD/RGD like motifs in the VP35 protein of Ebola virus raises the possibility of its role in exacerbation of the hemorrhagic syndrome, in addition to other factors already described in literature such as the binding of GP to integrins, elevated inflammatory cytokines, compromised vascular barrier etc. Future research, in this aspect might reveal definitive role of VP35 and glycoprotein's in the inhibition of platelet aggregation and pathogenesis of Ebola virus. Understanding the Ebola virus pathogenesis will enable design and development of efficient therapeutic strategies against hemorrhagic fever syndrome of Ebola virus.

Acknowledgements

Help of Dr Krishna Murthy, DVM. PhD in critically reviewing this manuscript and providing constructive suggestions is appreciated.

Conflicts of interest

None.

References

1. Hoenen T, Groseth A, Falzarano D et al. Ebola virus: unraveling pathogenesis to combat a deadly disease. *Trends Mol Med*. 2006;12(5):206–215.
2. Feldman HEA Filoviridae Virus Taxonomy, Eighth Report of the International Committee on Taxonomy of *Viruses*, Sheet. 2004
3. Ebola virus disease. *Elsevier Academic Press, USA*. 2016
4. Sureau PH Firsthand clinical observations of hemorrhagic manifestations in Ebola hemorrhagic fever in Zaire. *Rev Infect Dis*. 1989;11(Suppl 4):S790–S793.
5. Wahl-Jensen VM, Afanasieva TA, Seebach J et al. Effects of Ebola virus glycoproteins on endothelial cell activation and barrier function. *J Virol*. 2005;79(16):10442–10450.
6. Sanchez A, Lukwiya M, Bausch D et al. Analysis of human peripheral blood samples from fatal and nonfatal cases of Ebola (Sudan) hemorrhagic fever: cellular responses, virus load, and nitric oxide levels. *J Virol*. 2004;78(19):10370–10377.
7. Villinger F, Rollin PE, Brar SS et al. Markedly elevated levels of interferon (IFN)–gamma, IFN–alpha, interleukin (IL)–2, IL–10, and tumor necrosis factor–alpha associated with fatal Ebola virus infection. *J Infect Dis*. 1999;179 (Suppl 1):S188–S191.
8. Ramanathan CS, Taylor EW Computational genomic analysis of hemorrhagic fever viruses. Viral selenoproteins as a potential factor in pathogenesis. *Biol Trace Elem Res*. 1997;56(1):93–106.
9. Geisbert TW, Young HA, Jahrling PB et al. Mechanisms underlying coagulation abnormalities in Ebola hemorrhagic fever: over expression of tissue factor in primate monocytes/macrophages is a key event. *J Infect Dis*. 2003;188(11):1618–1629.
10. Feldmann H, Volchkov VE, Volchkova VA et al. The glycoproteins of Marburg and Ebola virus and their potential roles in pathogenesis. *Arch Virol Suppl*. 1999;15:159–169.
11. Nieswandt B, Varga–Szabo SD, Elvers M Integrins in platelet activation. *J Thromb Haemost*. 2009;7(Suppl 1):206–209.
12. Takada A, Watanabe S, Ito H, Okazaki K et al. Down regulation of beta1 integrands by Ebola virus glycoprotein: implication for virus entry. *Virology*. 2000;278(1):20–26.
13. Dahlmann F, Biedenkopf N, Babler A et al. Analysis of Ebola Virus Entry Into Macrophages. *J Infect Dis*. 2015;212(Suppl 2):S247–257.
14. Schornberg KL, Shoemaker CJ, Dube D et al. Alpha5beta1–integrin controls Ebola virus entry by regulating endosomal cathepsins. *Proc Natl Acad Sci U S A*. 2009;106(19):8003–8008.
15. Williams JA Disintegrins: RGD–containing proteins which inhibit cell/matrix interactions (adhesion) and cell/cell interactions (aggregation) via the integrin receptors. *Pathol Biol*. 1992;40(8):813–821.
16. Reiss SM, Sieber V, Oberle A, Wentzel P et al. Inhibition of platelet aggregation by grafting RGD and KGD sequences on the structural scaffold of small disulfide–rich proteins. *Platelets*. 2006;17(3):153–157.
17. Fisher–Hoch SP, Platt GS, Neild GH, Southee T et al. Path physiology of shock and hemorrhage in a fulminating viral infection (Ebola). *J Infect Dis*. 1985;152(5):887–894.
18. Sanchez A, Kiley MP Identification and analysis of Ebola virus messenger RNA. *Virology*. 1987;157(2):414–420.
19. Dolnik O, Volchkova VA, Escudero–Perez B et al. Shedding of Ebola Virus Surface Glycoprotein Is a Mechanism of Self–regulation of Cellular Cytotoxicity and Has a Direct Effect on Virus Infectivity. *J Infect Dis*. 2015;212(Suppl 2):S322–328.