

Superiority of CCL11x IL-6 over PSA in Prostate Cancer Prediction and Detection in Egyptian Patients: First Preliminary Comparative Assessment

Research Article

Volume 2 Issue 4 - 2015

Mohamed El-Far^{1*}, Hassan Abol Eneine² and Amira Ramadan¹¹Biochemistry Division, Chemistry Department, Faculty of Science, Mansoura University, Egypt²Department of Urology, Urology-Nephrology Centre, Mansoura University, Egypt***Corresponding author:** Mohamed El-Far, Division of Biochemistry, Faculty of Science, Mansoura University, Mansoura, Egypt, Tel: +20-502248348; Fax: +20-502246254; Email: elfarma2002@yahoo.com**Received:** March 31, 2015 | **Published:** April 29, 2015**Abstract**

Background: Controversy surrounds the use of prostate specific antigen (PSA) as a biomarker for prostate cancer detection, leaving an unmet need for a novel biomarker. In this regard, we investigated the association of two biomarkers, CCL11 and IL-6, with prostate cancer (PCa) and pointed out the diagnostic value of their combined detection and multiplication assessment.

Methods: In a cohort of 72 subjects (24 PCa; 24 benign prostatic hyperplasia (BPH); 24 healthy volunteers) serum concentrations of CCL11 and IL-6 were assessed by enzyme-linked immunosorbent assay (ELISA). Diagnostic performance of each biomarker and their multivariate assessments were compared using area under receiver operating characteristic curves (AUC) and the unpaired t-test.

Results: Mean serum concentrations of CCL11 and IL-6 were highly significantly ($P < 0.0001$) elevated in PCa compared to controls. When CCL11 concentrations multiplied by those of IL-6, the yielded values were increased 2.7-fold and 6.1-fold in benign and malignant groups, respectively, as compared with normal group. Late stage tumors showed that $[(CCL11) \times (IL-6)]$ values (1544.0 ± 176.0 pg/ml) were significantly ($P < 0.05$) higher than those of early stage (665.8 ± 68.5 pg/ml). The area under curve (AUC) was 0.933 with superior sensitivity (96%) and higher specificity (75%).

Conclusion: We are here the first to report that values from combined detection and multiplication of (CCL11 and IL-6) can be considered as novel promising bio-score biomarker for PCa detection and may be used also to distinguish between prostate enlargement and prostate cancer as well as its status. Our new score (Mansoura-Bioscore) may be investigated to be surrogate to/or to be done with PSA.

Keywords: Prostate cancer; Cancer diagnosis; Prostate biomarkers; CCL11; IL-6; PSA

Abbreviations: BPH: Benign Prostatic Hyperplasia; PSA: Prostate Specific Antigen; PCa: Prostate Cancer; ELISA: Enzyme-Linked Immunosorbent Assay

Introduction

Prostate cancer (PCa) is the second most common cancer in men, accounting for 10% of male cancers [1]. The incidence of PCa is increasing in most Western populations [2,3]. Today the only test that can fully confirm the diagnosis of prostate cancer is the transrectal ultrasound guided biopsy of 6-18 prostate cores in a patient, but this method is prone to sampling errors [4]. Measurement of serum prostate specific antigen (PSA) is the most common tool used to detect prostate cancer [5]. One of the problems of the PSA testing is that the biomarker itself has a weak correlation with degree of prostate malignancy; it is copiously produced by normal prostatic cells. So, lack of specificity and sensitivity limited its use as a screening tool in the general population [6,7]. There is therefore still an urgent need for non-invasive biomarkers for the detection of PCa.

Chemokines are a super family of small secreted proteins initially characterized by their ability to induce leukocyte migration [8]. During the transitions from normal to benign prostatic hyperplasia (BPH) and from BPH to PCa, a number of

chemokines display variations in their expression [9]. Several studies have shown that there are a potential link between CC-, and IL-type chemokines and cytokines as serum biomarkers and malignant proliferative diseases of the prostate [10-13]. The current study was designed to investigate multivariate assessments of CCL11, a CC-type chemokine, and IL-6, an IL-type cytokine, and evaluate their diagnostic utility to distinguish between prostatic enlargements as BPH and PCa among Egyptian patients.

Materials and Methods**Patient population**

The study was performed on 72 patients collected from the pool of patients referred to the Urology & Nephrology center seeking for treatment. Informed cancer was obtained from each patient who was included in the study. The studied patients were divided into 3 groups 24 patients each. Group 1, had a confirmed diagnosis of prostatic cancer by transrectal prostate biopsy. Group 2 had the documented diagnosis of benign prostatic hyperplasia (BPH). Group 3 included 24 healthy individuals with no prostatic complaints and after complete investigations. The diagnosis of prostate cancer included tumor histology, tumor stage, lymphatic involvements, Gleason score and the evidence of

radiological tumor metastases. The study protocol followed the ethical guidelines of the 1975 Helsinki Declaration.

Specimen processing

Six milliliters of blood from each subject were collected in vacutainer tubes (Becton Dickinson, Franklin Lakes, NJ) and were assigned a unique identifying number before delivery to the laboratory for processing. After collection, the blood was kept to retract for about 10 minutes. Each blood sample was then centrifuged at 1000×g for 10 minutes after which serum was decanted and aliquoted into cryovials and stored immediately at -20°C till analysis.

Laboratory analysis

Human CCL11 levels (Cat# DTX00 R&D Systems Inc., Minneapolis, MN, USA) and human IL-6 levels (eBioscience, San Diego, CA, USA) in serum were monitored using ELISA. The assays were conducted according to the manufacturer's instructions. The absorbance was read (Biotek, ELX800). Every sample was assayed in duplicate and the mean of two results was used. PSA serum levels were measured with an automated immunoassay analyzer (Axsyn, 7A83-97, Abbot).

Statistical analysis

All statistical calculations were done by a Statistical Package for the Social Sciences (SPSS); v.17.0 (SPSS Inc., Chicago, IL) and the GraphPad Prism package; v.5.0 (GraphPad Software, San Diego, CA). Patients characteristics were descriptive summarized and reported as mean ± standard error of the mean (SEM). To test the significance of differences between the mean levels of a marker in different patient groups we used an unpaired t-test. Receiver operator characteristic (ROC) curves were generated and the area under the curve (AUC) tested for significance using an unpaired t-test against the hypothesis that the real area under the curve was 0.5 (i.e. no diagnostic value). The cut-off values for optimal clinical performance measures were determined from the ROC curves.

Results

Clinical and pathologic characteristics

Prostate cancer staging was performed according to the TNM system of the American Joint Committee on Cancer (AJCC). Half of the patient had organ confined disease (T1, T2), while the remaining half had locally advanced tumor (T3, T4) with or without metastases. The patients were subdivided into two pathological groups according to the Gleason score: those who had Gleason score ≤ 7 in 14 patients while those who had >7 score were 10 patients. Other demographic and tumor characteristics are shown in Table 1.

The ability of serum IL-6 and CCL11 in comparison with PSA to detect clinically significant PCa

Serum PSA, IL-6 and CCL11 were measured in 24 PCa, 24 BPH and 24 healthy individuals. The difference in serum concentrations of PSA, IL-6 and CCL11 in patients with and without PCa indicated potential diagnostic values. Figure 1 provides an array of ROC curves for PSA (Figure 1a), IL-6 (Figure 1b) and CCL11 (Figure 1c) to investigate their capability to distinguish between PCa and non PCa patients. The AUC calculated in this patient set was 0.865

for PSA, 0.870 for IL-6 and 0.880 for CCL11.

Table 1: Clinical information of the prostate cancer patients.

| Factor | Value |
|------------------------------------|------------------|
| No. of Subjects | 24 |
| Age (Years): Range (Mean±SD) | 59-81 (69.6±6.2) |
| Tumor Histology: (Number) | |
| Adenocarcinoma | 21 |
| Bone Metastatic Adenocarcinoma | 3 |
| Pathological T Stage: (Number) | |
| ≤T2 | 12 |
| >T2 | 12 |
| Grade: (Number) | |
| Well | 3 |
| Moderate | 7 |
| Poor | 11 |
| Unknown | 3 |
| Metastasis: (Number) | |
| +Ve | 12 |
| - Ve | 12 |
| Lymphatic Invasion: (Number) | |
| +Ve | 12 |
| -Ve | 12 |
| Gleason Score Categories: (Number) | |
| ≤7 | 14 |
| >7 | 10 |

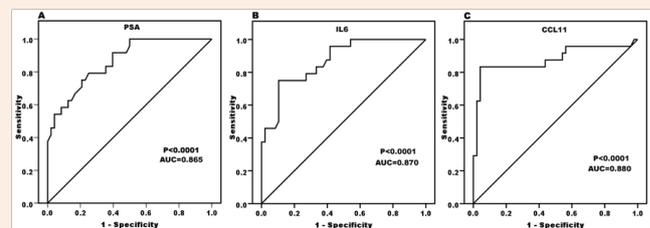


Figure 1: Discriminating prostate carcinoma patients from all non-cancer individuals: comparison of the ROC curves of PSA, IL-6 and CCL11. ROC: Receiver Operating Characteristic; AUC=Area Under Curve.

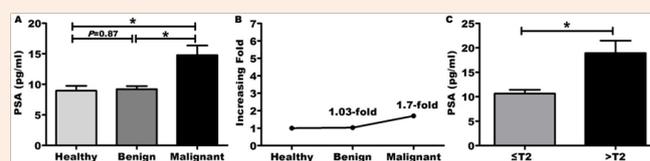


Figure 2: PSA for prostate cancer detection.

- Mean serum PSA levels in prostate cancer patients and non-cancer patients as measured.
- Fold increase in patients groups compared to healthy individuals.
- Mean serum PSA levels were significantly elevated in late stages (higher than T2) than in early stages (lower than or equal T2) prostate cancer.

*P<0.05 is considered significant

Figure 2 showed PSA for prostate cancer detection.

- Mean serum PSA levels in prostate cancer patients and non-cancer patients as measured.
- Fold increase in patients groups compared to healthy individuals.
- Mean serum PSA levels were significantly elevated in late stages (higher than T2) than in early stages (lower than or equal T2) prostate cancer.

IL-6 and CCL11 combination is predictive for PCA

Combining serum concentrations of IL-6 and CCL11 (IL-6+CCL11) increased the AUC to 0.900 (Figure 3a). This combination yielded values which were observed to be significantly higher in patients with PCa (197.4±14.3 pg/ml) and BPH (124.5±4.7 pg/ml) when compared with healthy control (97.8±1.1 pg/ml, $P<0.0001$ for each comparison, Figure 3b). These yielded values were increased 1.3-fold and 2-fold in benign and malignant group, respectively, compared with normal control group (Figure 3c). Also, the values of this combination were increased significantly ($P<0.0001$) according to the stage of PCa (147.6±8.9pg/ml for early stages, 247.2±17.9pg/ml for late stages, Figure 3d).

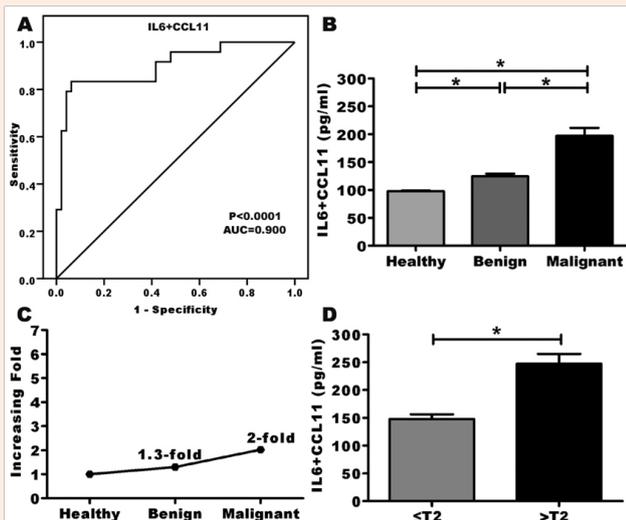


Figure 3: The combined use of IL-6 and CCL11 for prostate cancer detection.

- Receiver operator characteristic (ROC) analysis of all cancer samples vs. all non-cancer samples.
- Mean serum (IL-6+CCL11) levels in prostate cancer patients and non-cancer patients as measured by ELISA. (C) Fold increase in patients groups compared to healthy individuals.
- Mean serum (IL-6+CCL11) levels were significantly elevated in late stages (higher than T2) than in early stages (lower than or equal T2) prostate cancer.

* $P<0.05$ is considered significant

IL-6 and CCL11 multiplication is most predictive of PCA

A multivariate assessment of these variables determined that the multiplication of IL-6 with CCL11 (IL-6×CCL11) was most predictive of prostate disease outcome. This multivariate test system to detect PCa produced an ROC curve with an AUC 0.933 (Figure 4a). This multiplication increase the power to discriminate PCa (1105±130.0 pg/ml) from BPH (484.5±42.0

pg/ml) and healthy (182.5±11.0 pg/ml) individuals (Figure 4b). Interestingly, serum (IL-6×CCL11) level was 6.1-fold and 2.7-fold higher in PCa and BPH, respectively than healthy control (Figure 4c). When comparing only the different stages of PCa, the serum (IL-6×CCL11) levels increase significantly ($P<0.0001$) as the stage increases (early stages (665.8±68.5 pg/ml), advanced stages ((1544.0±176.0 pg/ml), Figure 4c).

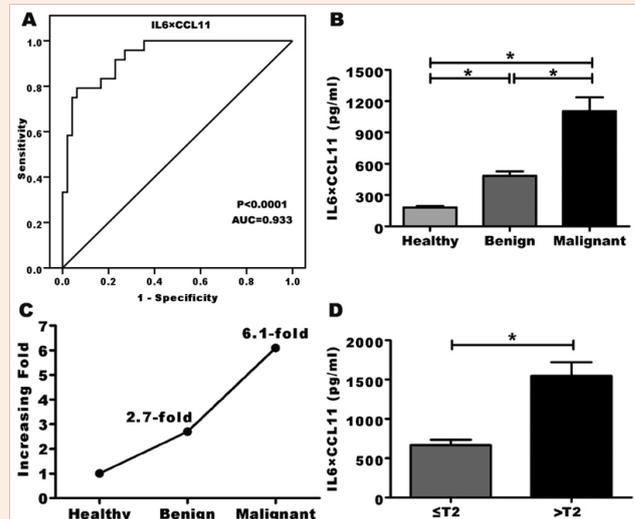


Figure 4: Multiplying IL-6 with CCL11 for prostate cancer detection.

- Receiver operator characteristic (ROC) analysis of all cancer samples vs. all non-cancer samples.
- Mean serum (IL-6×CCL11) levels in prostate cancer patients and non-cancer patients as measured by ELISA. (C) Fold increase in patients groups compared to healthy individuals.
- Mean serum (IL-6×CCL11) levels were significantly elevated in late stages (higher than T2) than in early stages (lower than or equal T2) prostate cancer.

* $P<0.05$ is considered significant

Sensitivity and specificity for investigated parameters in detection of PCA

The benign and healthy normal groups were combined in a non-malignant group, and the best cutoff values for investigated parameters were calculated by the ROC curve as 9 ng/ml for PSA, 3.6 pg/ml for IL-6, 116 pg/ml for CCL11, 120.3 pg/ml for IL-6+CCL11 combination and 418.5 pg/ml for IL-6×CCL11 multiplication.

Table 2: Ability of serum biomarkers in predicting the presence of prostate cancer.

| Marker | Cutoff | Sn (%) | Sp (%) | PPV | NPV | Ac (%) |
|------------|-------------|--------|--------|-----|-----|--------|
| PSA | 9 ng/ml | 79 | 73 | 61 | 88 | 76 |
| IL-6 | 3.6 pg/ml | 75 | 73 | 60 | 86 | 75 |
| CCL11 | 116 pg/ml | 82 | 73 | 61 | 90 | 76 |
| IL-6+CCL11 | 120.3 pg/ml | 83 | 75 | 61 | 90 | 76 |
| IL-6×CCL11 | 418.5 pg/ml | 96 | 75 | 64 | 97 | 81 |

Sn: Sensitivity; Sp: Specificity; PPV: Positive predictive value; NPV: Negative predictive value; Ac: Accuracy

Sensitivity and specificity for each serum marker as well as

their combination and multiplication were tested for detection of PCa. Combining IL-6 and CCL11 increased sensitivity and specificity. The best sensitivity was achieved when the serum IL-6 multiplied with CCL11 (Table 2).

Discussion

Prostate cancer is the most frequent malignancy in men in the Western world [14]. This cancer is mainly detected by the determination of serum PSA. However, several studies showed the limited ability of PSA in differentiating between benign and malignant prostate diseases and between aggressive and insignificant tumors since a continuous risk of prostate cancer occurs at all PSA values [15,16]. Thus there is a need for new PCa diagnostic biomarkers.

Chemokines are part of the network of inflammatory mediators associated to neoplasia irrespective of pathogenesis [17]. Recently among several chemokines studied in PCa, CCL11, a potent chemotactic factor for eosinophils, actually demonstrated elevated levels in PCa [1,10,18]. On the other hand IL-6 is a proinflammatory cytokine that plays an important role in intraprostatic inflammation and thus carcinogenesis [19].

In this regard, we aimed to estimate the diagnostic performance of the combined use of CCL11, IL-6 and those of each separately compared with PSA in PCa detection. Using ELISA, we examined CCL11 and IL-6 serum levels in patients with biopsy and clinically confirmed BPH or various stages of PCa. Increasing concentrations of both CCL11 and IL-6 positively correlated to tumor burden and identified patients with all stages of PCa. This may be explained by the fact that suggested by previous pathological studies that inflammation is implicated in prostate carcinogenesis [20,21]. Therefore, proinflammatory cytokines (include CCL11 and IL-6) influence prostate cancer risk and the local production of these chemokines by the tumor can also result in increased chemokine concentration in the blood, as reported for patients with breast and gastric carcinoma [18,22,23].

In a first attempt to unravel the diagnostic utility of the combined detection of CCL11 and IL-6 in PCa, CCL11+IL-6 revealed higher diagnostic value (AUC of 0.900) at a 120.3 pg/ml cutoff with higher sensitivity (83%) and specificity (75%) for detecting the presence or absence of PCa. Interestingly, the highest diagnostic ability (AUC=0.933) was achieved when serum CCL11 concentrations were multiplied by those of IL-6 with sensitivity of 96% and specificity of 75%. Moreover, late stage tumors were associated with higher mean (CCL11×IL-6) values than early stage tumors ($P<0.05$). This compared favorably with results revealed from using PSA alone (sensitivity 79%, specificity 73% and AUC=0.865). In other analysis, the estimated sensitivity of a PSA cutoff of 4.0 ng/mL was 21% for detecting any prostate cancer and 51% for detecting high-grade cancers [24].

Future studies should focus on the prospective analysis of these markers against other established serum markers of prostate cancer in larger multi-centric studies. However, this approach is beyond the scope and limited financial resources of the present work. Taken together, our study highlights –for the first time– the potential utility of the combined use of CCL11 and IL-6 as novel diagnostic bio-score biomarker for PCa using a multiplication of serum value levels of both CCL11 and IL-6. We recently showed another equation in different type of cancer for

the first time the impact of serum epithelial membrane antigen (EMA) and cytokeratin-1 (CK1) ratio in breast cancer detection, and showed that EMA/CK1 values may serve as diagnostic marker in early stage breast cancer [25].

Acknowledgement

Prof. Mohamed El-Far, wishes to express his gratitude to late Amira Ramadan, and also Mohamed Abdelrazek for statistical assistance.

References

1. Milone MR, Pucci B, Bruzzese F, Carbone C, Piro G, et al. (2013) Acquired resistance to zoledronic acid and the parallel acquisition of an aggressive phenotype are mediated by p38-MAP kinase activation in prostate cancer cells. *Cell Death Dis* 4: e641.
2. Hsing AW, Chokkalingam AP (2006) Prostate cancer epidemiology. *Front Biosci* 11: 1388-1413.
3. Kim WT, Kim WJ (2013) Micro RNAs in prostate cancer. *Prostate Int* 1(1): 3-9.
4. Durkan GC, Sheikh N, Johnson P, Hildreth AJ, Greene DR (2002) Improving prostate cancer detection with an extended-core transrectal ultrasonography-guided prostate biopsy protocol. *BJU Int* 89(1): 33-39.
5. Loeb S, Catalona WJ (2007) Prostate specific antigen in clinical practice. *Cancer Lett* 249(1): 30-39.
6. Nna E (2013) The end of the road for prostate specific antigen testing? *Niger J Clin Pract* 16(4): 407-417.
7. Killick E, Morgan R, Launchbury F, Bancroft E, Page E, et al. (2013) Role of Engrailed-2 (EN2) as a prostate cancer detection biomarker in genetically high risk men. *Sci Rep* 3: 2059.
8. Salazar N, Castellan M, Shirodkar SS, Lokeshwar BL (2013) Chemokines and chemokine receptors as promoters of prostate cancer growth and progression. *Crit Rev Eukaryot Gene Expr* 23(1): 77-91.
9. Vindrieux D, Escobar P, Lazennec G (2009) Emerging roles of chemokines in prostate cancer. *Endocr Relat Cancer* 16(3): 663-673.
10. Agarwal M, He C, Siddiqui J, Wei JT, Macoska JA (2013) CCL11 (eotaxin-1): a new diagnostic serum marker for prostate cancer. *Prostate* 73(6): 573-581.
11. Alcover J, Filella X, Luqué P, Molina R, Izquierdo L, et al. (2010) Prognostic value of IL-6 in localized prostatic cancer. *Anticancer Res* 30(10): 4369-4372.
12. Drachenberg DE, Elgamil AA, Rowbotham R, Peterson M, Murphy GP (1999) Circulating levels of interleukin-6 in patients with hormone refractory prostate cancer. *Prostate* 41(2): 127-133.
13. Mandić S, Sudarević B, Marcz S, Horvat V, Cosić I, et al. (2013) Interleukin-6 polymorphism and prostate cancer risk in population of Eastern Croatia. *Coll Antropol* 37(3): 907-911.
14. Jung K, Stephan C (2013) Thiosulfate in urine: new hope or new failure of a biomarker for prostate cancer?. *Clin Chem Lab Med* 51(9): 1695-1697.
15. Guazzoni G, Nava L, Lazzeri M, Scattoni V, Lughezzani G, et al. (2011) Prostate-specific antigen (PSA) isoform p2PSA significantly improves the prediction of prostate cancer at initial extended prostate biopsies in patients with total PSA between 2.0 and 10 ng/ml: results of a prospective study in a clinical setting. *Eur Urol* 60(2): 214-222.
16. Dahm P, Neuberger M, Ilic D (2013) Screening for prostate cancer:

- shaping the debate on benefits and harms. *Cochrane Database Syst Rev* 9: ED000067.
17. Mantovani A, Allavena P, Sica A, Balkwill F (2008) Cancer-related inflammation. *Nature* 454: 436-444.
 18. Mantovani A, Savino B, Locati M, Zammataro L, Allavena P, et al. (2010) The chemokine system in cancer biology and therapy. *Cytokine Growth Factor Rev* 21(1): 27-39.
 19. Mandić S, Sudarević B, Marczl S, Horvat V, Cosić I, et al. (2013) Interleukin-6 polymorphism and prostate cancer risk in population of Eastern Croatia. *Coll Antropol* 37(3): 907-911.
 20. Schlaberg R, Choe DJ, Brown KR, Thaker HM, Singh IR (2009) XMRV is present in malignant prostatic epithelium and is associated with prostate cancer, especially high-grade tumors. *Proc Natl Acad Sci USA* 106(38): 16351-16356.
 21. De Marzo AM, Platz EA, Sutcliffe S, Xu J, Grönberg H, et al. (2007) Inflammation in prostate carcinogenesis. *Nat Rev Cancer* 7(4): 256-269.
 22. Dwyer RM, Potter-Beirne SM, Harrington KA, Lowery AJ, Hennessy E, et al. (2007) Monocyte chemotactic protein-1 secreted by primary breast tumors stimulates migration of mesenchymal stem cells. *Clin Cancer Res* 13(17): 5020-5027.
 23. Koç Ü, Çetinkaya E, Bostancı EB, Kemik AS, Tez M, et al. (2013) Diagnostic significance of serum eotaxin-1 level in gastric cancer patients. *Dis Markers* 35(5): 363-367.
 24. Meigs JB, Barry MJ, Oesterling JE, Jacobsen SJ (1996) Interpreting results of prostate-specific antigen testing for early detection of prostate cancer. *J Gen Intern Med* 11(9): 505-512.
 25. Attallah AM, El-Far M, Omran MM, Abdallah SO, El-Desouky MA, et al. (2014) Circulating levels and clinical implications of epithelial membrane antigen and cytokeratin-1 in women with breast cancer: can their ratio improve the results? *Tumour Biol* 35(11): 10737-10745.