Role of biofilms in fungal keratitis- an overview

Abstract

Biofilms are generally defined as a community of sessile microbes held together by a polymeric extracellular matrix (ECM), adherent to a surface that are phenotypically distinct from their planktonic counterparts. Fungal keratitis is an important cause of ocular morbidity in the form of corneal opacity and visual loss, especially in developing countries. Predisposing factors for fungal keratitis include tropical or subtropical climate, agricultural work, contact lens users and corneal trauma. Fungal keratitis is most commonly caused by filamentous fungi like Fusarium spp. (F. solani and F. oxysporum) followed by Aspergillus spp. and dematiaceous fungi. Fungal biofilm formation plays an important role in the development of fungal keratitis. Here in this review, we will be discussing the formation of biofilms and its role in fungal keratitis.

Keywords: biofilm, fungal keratitis, contact lens

Introduction

Biofilm formation has been detected quite early in the fossil record (approx. 3.25 billion years ago and is common throughout diverge range of organisms. However, the first description of a biofilm dates back to the 17th century when Anthony van Leeuwenhoek noted the presence of small living “animalcules” in his dental plaque.1 But nothing was clear and it took an additional 200 years until the germ theory of disease was advanced by Robert Koch before a connection between microbes and disease was made.2 Recent advances in confocal microscopy and molecular genetics have provided evidence that biofilm formation is the most common mode of growth of microorganisms in nature. This growth form allows microbial cells to survive in hostile environments, enhances their resistance to physical and chemical pressures, and promotes metabolic cooperation.2 In fact, it is estimated that approximately 80% of all bacteria in the environment exist in biofilm communities, and more than 65% of human microbial infections involve biofilms.3 Microbial biofilms are communities of micro-organisms that are strongly attached to biologic and nonbiologic substrata, enclosed in a self-produced protective exopolymeric matrix (EPM). It is seen mixed poly-microbial biofilms are common and can be considered even more complex communities of microbes that cooperatively interact in an altruistic manner.4 The type of organisms causing keratitis depends on geographical conditions such as differences in climate, environment and occupational risk.5 Bacterial keratitis, especially contact lens-associated infection, is caused by both Gram negative pathogens, such as P. aeruginosa and Serratia spp., and Gram positive organisms, such as S. aureus6 and other staphylococcal species.5,7 The recognition of fungal biofilms brought attention to nosocomial infections and now it is believed that several fungal genera are capable of organizing into a biofilm.8 Here in this review, we will be discussing the formation of fungal biofilms and its role in fungal keratitis.

Fungal biofilms

Biofilms are generally defined as a community of sessile microbes (bacterial or fungal) held together by a polymeric extracellular matrix (ECM), adherent to a surface that are phenotypically distinct from their planktonic counterparts.9 Members of a biofilm community, which can be of the same or multiple species, show varying stages of differentiation and exchange information, metabolites, and genes with each other. As a result, members of the biofilm community are in a diversity of physiologies influenced by the unequal sharing of nutrients and metabolic byproducts, which results in subpopulations with increased tolerance to anti-fungals and environmental stresses, the host immune system, and predatory microorganisms.11,12 The process of biofilm formation occurs in five stages (1) reversible aggregation of planktonic cells on a surface; (2) irreversible adhesion; (3) formation of microcolonies; (4) biofilm maturation; and (5) detachment and dispersion of cells.13 Candida albicans biofilms are comprised primarily of yeast-form and hyphal cells, both of which are required for biofilm formation.14 Formation is a sequential process involving adherence to a substrate (either abiotic or mucosal surface), the proliferation of yeast cells over the surface, and induction of hyphal formation.14 ECM accumulates as the biofilm matures, and seems to contribute to cohesion.15 C. albicans biofilms form on numerous abiotic and biotic surfaces.16 Other Candida spp. including C. tropicalis, C. parapsilosis, and C. glabrata form ECM-containing biofilms but do not produce true hyphae.17 Aspergillus biofilms can form both on abiotic and biotic surfaces.19 The initial colonizing cells that adhere to the substrate are conidia. Mycelia (the hyphal form) develop as the biofilm matures.19 Hyphae of C. albicans and of A. fumigatus can form pores or channels through biotic surfaces.20,21 Other fungal pathogens Trichosporon asahii, Coccidioides immitis, C. neoformans, Pneumocystis species etc. do not produce hyphal structures as part of their biofilms (Figure 1).22

Fungal keratitis

Fungal keratitis is an important cause of ocular morbidity in the form of corneal opacity and visual loss especially in developing countries, where it may account for nearly half of corneal ulcers.15 Predisposing factors for fungal keratitis include tropical or subtropical climate, agricultural work, contact lens users and corneal trauma. Fungal keratitis is most commonly caused by filamentous fungi like Fusarium spp. (F. solani and F. oxysporum) followed by Aspergillus spp. and dematiaceous fungi (Figure 2).23,24
Contact lens-associated fungal keratitis

High risk of fungal keratitis in contact lens wearers has been associated with the ability of the lens to induce modification of the corneal epithelium and to carry organisms to the ocular surface. Use of lens induces local alterations like hypoxia and hypercapnia, which affect the ability of the epithelium to respond to damage. Tear fluid exchange may also be compromised between the anterior and posterior sides of the lens thereby limiting its antimicrobial properties. In addition, contact lenses provide a surface where microorganisms may attach and colonize the surface as a biofilm and can spread to a previously damaged corneal epithelium. Poor hygiene and infrequent replacement of the contact lens storage cases are independent risk factors for moderate and severe keratitis. The incidence of contact lens-associated microbial keratitis has been shown to be impacted by the contact lens material, and also by the wear schedule. Higher risk for development of keratitis was seen with daily wear soft contact lenses compared to daily wear rigid gas permeable lenses, and it was further increased for extended wear (overnight wear) soft contact lenses. Recently introduced daily disposable and silicone hydrogel contact lenses have also been associated with higher incidence of keratitis compared to rigid gas permeable contact lenses.

Role of contact lenses in biofilm formation

Once the lens is placed on the surface of the eye, the disinfectant diffuses off the lens and is replaced by lipids and proteins present in the tear fluid. Microbial adhesion to surfaces coated by with proteins and other biomolecules is often accomplished by a class of molecules termed Microbial Surface Components Recognizing Adhesive Matrix Molecules (MSCRAMM), as well as other adhesive surface proteins. In a moving suspension, cells are exposed to fairly uniform conditions. However, following attachment, the individual experience of a cell begins to differ from its neighbors (i.e., a cell in the middle of a group will experience more excreted products and fewer factors from the environment than a cell on the periphery of a population), and as a result, cells begin to differentiate. Many biofilms involve the production of an extracellular matrix (ECM) that encases the cells, and in some cases, binds the cells together and that can be composed of polysaccharides, lipopolysaccharides, proteins, or extracellular DNA. Extracellular matrix contributes to the differentiation of cells within the encased population and protects the organism from the action of antimicrobial agents, host immune responses, bacteriophages and phagocytic amoeba.

Role of biofilms in fungal keratitis

Keratitis caused by Fusarium F. solani and F. oxysporum is among the most refractory and common causes of fungal keratitis. Fusarium is a filamentous fungus that infests agricultural plants, and the incidence of Fusarium keratitis peaks during harvest seasons when farm workers are at more risk of corneal injury and exposure to airborne spores. However, another major risk factor for Fusarium keratitis is contact lens wear due to increased association with contaminated lenses, lens cases, and contact lens care solutions. The formation of biofilm by these fungi on contact lenses and lens cases is thought to have a major role in causing keratitis once the lens is placed on the ocular surface. From 2005-2007, more than 300 cases of Fusarium keratitis were associated with contact lens wear and in some cases, characterized as a serious disease in which patient underwent corneal transplantation or even complete eye removal. The severity of disease was assigned to misdiagnosis and inappropriate treatment, and to the failure of antimycotic agents due to the resistance exhibited by the micro-organisms in the biofilms.
observed on contact lenses and the 2005-2006 outbreak was attributed partly to the ability of Fusarium to form biofilms.\textsuperscript{38,39} Another study showed that Fusarium isolates from the outbreak formed biofilms on soft contact lenses \textit{in vitro} and \textit{in vivo} tested in an inanimate model, and that biofilm formation was elevated in \textit{F. solani} compared to \textit{F. oxysporum} isolates. These biofilms were susceptible to natamycin, but exhibited species-dependent susceptibility to Amphotericin B and Voriconazole.\textsuperscript{40,41}

**Conclusion**

The Ability of Fungi fungi to form biofilms is a critical determinant in resistance to anti fungals and to the pathogenesis of fungal keratitis, and it should be taken into consideration when managing patients with this disease. However, further research is required in the field of fungal biofilms and their role in human diseases.

**Acknowledgements**

None.

**Conflict of interest**

The author declares no conflict of interest.

**References**


