

Production of fermented fruit juice and value addition by blending medicinal plants

Abstract

Fermented fruit juice is beverage typically made of controlled fermentation of fruits. The fruits used in the study were Wild Grapes, Guava, Sapota, Fig, Pomegranate, Kokum, and blends of Guava with Kokum and Sapota with medicinal plants Ginger (*Zingiber officinale*), Tulsi (*Ocimum tenuiflorum*), Amrutha balli (*Tinospora cordifoli*) and Dodda pathre (*Coleus aromaticus*). The musts of the fruits were extracted, pasteurized and subjected to anaerobic fermentation by inoculating with yeast with an initial pH of 3.5 and initial sugar concentration of 20-22°Brix at 28°C±2°C. Then fermented fruit juice was subjected to malolactic fermentation using *Oenococcus oeni*. The residual concentration of all the samples was found to be less than 1g/L, fixed acidity in terms of tartaric acid equivalent was found to be in the range of 13.93g/L to 3.31g/L, radical scavenging activity in terms of ascorbic acid equivalent was found to be in the range 0.15-0.5mg/mL, percentage of ethanol was found to be in the range of 9.1-10.1% (v/v).

Keywords: fermented fruit juice, acidity, residual sugar, ethanol

Volume 5 Issue 6 - 2017

Vinayaka B Shet,¹ Sagar SD,¹ Bollamma MN,¹ Mary Teena J,¹ Vaman Rao C,¹ Aparna A,² Silvia Yumnam³

¹Department of Biotechnology Engineering, nmAM Institute of Technology, India

²Department of Microbiology and Molecular Genetics, Hebrew University of Jerusalem, Israel

³College of Pharmacy, Gachon University, Republic of Korea

Correspondence: Vinayaka B Shet, nmAM institute of technology (V.T.U, Belagavi), Nitte, Karkala, Karnataka, India, Tel +918258281263, Email vinayakabshet@nitte.edu.in

Received: October 26, 2017 | **Published:** November 22, 2017

Introduction

Fermented fruit juice is a popular drink being enjoyed all over the world. Historians believe that fermented fruit juice was being made in Caucasus and Mesopotamia as early as 6000 BC. Rig-Veda amply testifies that the fermented fruit juice is perhaps the oldest fermented product known to man. It has been made in India for as many as 5,000 years. In developing countries like India 20-30% of fruits produced are wasted due to lack of proper utilization, post-harvest and processing technology. By converting the waste into value added products like fermented fruit juice is a smart solution for this problem.¹ Any fruit with good proportion of sugar may be used in producing fermented fruit juice and the resultant fruit juice is normally named after the fruit. The type of fermented fruit juice to be produced dictates the fruit and strain of yeast to be involved. Preservatives used in fermented fruit juice making include sulphur-dioxide potassium sorbate, sorbic acid and metabisulphites.

Fermented fruit juice is one of the functional fermented foods and has many health benefits. Epidemiological evidence has been provided showing that constituents in fruits are beneficial to human health and contribute to the prevention of degenerative processes caused by oxidative stress.² Fruits contain many different dietary phytonutrients with strong antioxidant capacities; such as: phenolics, which include flavonoids and phenolic acids; carotenoids; and vitamins. Dietary intake of plant phenolics are inversely related to coronary heart disease and act as anti-ulcer, antispasmodic, anti-secretory, or anti-diarrheal agents in the gastrointestinal tract. Certain flavonoids have been shown to inhibit the activity of free radical generating enzymes aldose reductase, which cause diabetic cataracts and tumour growth in modelled systems. The concentration of some minerals in fermented fruit juice is important due to health impact of minerals, their role in the stability of fermented fruit juice, possibility of toxicological risks and food regulations. The mineral profile of fermented fruit juices has also been proposed as a possible fingerprint that could be used to characterize fermented fruit juices based on their geographical origin.³

Fruits like guava and pomegranate are easy to culture, possess high nutritive value and its products like juices, beverages nectars, etc. are

largely appreciated by the consumers. Average energy contribution to total energy intake is estimated to be 10- 20% among adults. These fruits are difficult to keep for long and are utilized either as fresh or processed juice and specialty products. In general fruit fermented fruit juices are processed in the same way as wine made from grapes and significant compositional changes take part during wine making. Likewise phenolic compounds are not only health promoting bioactivities but also greatly contribute to the sensory properties of stuff by alternating colour taste.⁴ Fruit juices contain water and 20% carbohydrates, 1% organic acids and trace amounts of vitamins, minerals and nitrogenous compounds. The sugars, organic acids and phenolics give the juice its flavour, while the vitamins, minerals and nitrogenous compounds are, in many cases, essential to yeast growth and fermentation. Fermented fruit juice has a similar composition, but has much lower levels of sugar (none in dry wines), 8-13% alcohol and a greater range of minor components.⁵ In the current investigation an attempt was made to blend unconventional fruits with medicinal plants to produce fermented fruit juice.

Materials and methods

Preparation of mash

Fruits were collected from local market of Karkala situated in Udupi district of Karnataka state, India. Fruits were washed with water and further with 50ppm potassium metabisulfite solution. To prepare the mash, fruits were crushed using mortar and pestle. The mash along with the seeds and the rind was pasteurized at 70-80°C for 15 minutes. After pasteurization, filtration was carried out using strainers and 500ml of the filtrate was collected.⁶

Blending medicinal plants

Medicinal plants such as Ginger (*Zingiber officinale*), Tulsi (*Ocimum tenuiflorum*), Amrutha balli (*Tinospora cordifoli*) and Dodda pathre (*Coleus aromaticus*)^{7,8} were collected from then mAMIT campus situated in Nitte village. These plant materials were washed thoroughly with tap water. To incorporate medicinal value, plant materials were boiled for 10min and extract was cooled to

room temperature. Further plant extract along with plant materials blended with mash (Table 1). Initial sugar was determined by using a hydrometer in terms of °Brix. Final sugar concentration was adjusted to 20-22°Brix. To bring the sugar concentration to the required value, chaptalization was carried out by adding table sugar.⁴ The pH was adjusted to 3.5 by adding tartarate crystals.⁹

Table 1 Sample composition

Name	Sample components
F1	Wild grapes I
F2	Wild grapes 2
F3	Guava
F4	Sapota
F5	Kokum+Guava
F6	Pomegranate
F7	Fig
F8	Sapota+Ginger
F9	Sapota+Tulsi
F10	Sapota+Dodda pathre
F11	Sapota+Amrutha balli

Inoculum preparation and anaerobic fermentation

Inoculum preparation: Freeze dried *Saccharomyces cerevisiae* was rehydrated by transferring 1g of yeast into 10mL of water and incubated for 15min at 28°C±2°C. To the rehydrated *Saccharomyces cerevisiae* few drops of mash was transferred and further incubated for 10min at 28°C±2°C. As the effervescence started generating, Erlenmeyer flask containing mash was pitched in with inoculum.

Anaerobic fermentation: To maintain the anaerobic condition, airlock was used. U-shaped airlock made up glass was half filled with water to prevent the entry of oxygen and to facilitate release of carbon dioxide liberated during anaerobic fermentation. Centrally bored cork having airlock was fitted to conical flask. To avoid growth of microorganism in the airlock, Potassium metabisulphite was added to the water present in the airlock. Fermentation was carried out at 28°C±2°C.

Malolactic fermentation

The culture of *Oenococcus oeni* was procured from NCIM, Pune, India. To each of the fermented fruit juice samples, 0.1mL of the culture was inoculated and malolactic fermentation was carried out for 7days.

Analytical methods

Initial sugar concentration in terms of Brix was determined using hydrometer. The residual sugar concentration was estimated on daily basis using the spectrophotometric method using the UV Vis spectrophotometer at 540nm with 3, 5- DNSA reagent.¹⁰ Titratable acidity of the fermented fruit juice was determined by titration method of a strong base against sample to an end point of pH 8.2 using potentiometric titration.¹¹ Volatile acidity of samples was determined by distillation of fermented fruit juice and the distillate was titrated against NaOH using phenolphthalein as indicator to determine volatile acid content.¹² The radical scavenging activity of fermented

fruit juice samples were analysed by the 2, 2-diphenylpicrylhydrazyl (DPPH) method.¹³ The percentage of ethanol present in the fermented fruit juice samples was estimated using GC (Shimadzu GC-2014).

Results and discussion

Estimation of initial sugar concentration

Initial sugar concentration plays major role in fermentation and ethanol concentration. Soon after crushing fruits estimation of initial sugar concentration was done using a hydrometer. Then concentration was adjusted to 18 Brix by capitalization. The initial sugar concentration (0Brix) was found to be highest in Chickoo (15.20Brix) and lowest in guava (1.30Brix). The sugar concentrations were made up to approximately 20-220Brix by chaptalization.¹⁴

Estimation of residual sugar

The residual concentration of all the samples was found to be less than 1g/L or 0.11 0Brix and there is no chance of contamination at the time of storing fermented fruit juice, hence the longer shelf life.

Estimation of acidity

Amount of acetic acid present in fermented fruit juice is expressed as volatile acidity. The result reveals the presence of acetic acid in the fermented fruit juice.

Titrate acidity is determined using following equation.

$$\text{Titrate acidity (g/L)} = 75 \times N \times (T/S) \quad (1)$$

Where N is the normality of NaOH, T is the titer volume (inmL), S is the sample volume (inmL) and 75 is a constant.

$$\text{Fixed acidity} = \text{Titrate acidity} - \text{Volatile acidity} \quad (2)$$

As per the Organisation of Vine and Wine (OIV) norms fixed acidity should not be less than 5g/L.

The fixed acidity in terms of tartaric acid equivalent was found to be highest in pomegranate (5.87g/L) and lowest in sapota with ginger (3.31g/L).

Radical scavenging activity

The radical scavenging activity of the fermented fruit juice samples was calculated using DPPH assay and was found to be as follows in terms of ascorbic acid equivalent. The radical scavenging activity for all the samples in terms of ascorbic acid equivalent was found to be in the range 0.08-0.49g/L.

Ethanol estimation

The percentage of ethanol present in the fermented fruit juice samples was estimated using gas chromatography. The percentage of alcohol for all the fermented fruit juice samples was found to be in the range of 9.0-10.5%. The highest ethanol content is present in pomegranate.

Conclusion

The current research work has revealed the possibility of blending medicinal plants with fruit juice for value addition. After analysing the fermented fruit juice that was produced from different fruits and blending combination it was found that the residual sugar concentration was less than 1g/L. The fixed acidity was maximum in Guava fermented fruit juice 5.87g/L, Wild grapes showed the

maximum radical scavenging activity, 0.49g/L and the maximum ethanol content of 10.% (v/v) was found in pomegranate. Quality of the fermented fruit juice can be improved further to match market requirement.

Acknowledgements

None.

Conflict of interest

The authors declare that they have no conflict of interest.

References

1. Reddy LV, Sudheer Kumar Y, Reddy OV. Production and Characterization of Wine from Mango Fruit (*Mangifera indica* L.). *Indian J Microbiol.* 2005;50(2):183–191.
2. Bansal N, Soni R, Soni SK. Standardization of conditions for fermentation and maturation of wine from Amla (*Emblia officinalis Gaertn*). *Nat Prod Rad.* 2009;8(4):436–444.
3. Edwards GC, Beelman RB. Inducing malolactic fermentation in wine. *Biotechnology advances.* 2002;7(3):336–360.
4. Gurvinder SK, Pooja. Status of wine production from guava (*Psidium guajava* L.): A traditional fruit of India. *Afr J Food Sci.* 2011;5(16):851–860.
5. Mena P, Vilaplana AG, Martí N, et al. Pomegranate varietal wines: Phytochemical composition and quality parameters. *Food Chem.* 2012;133(1):108–115.
6. Ganjyal GM, Hanna NA, Devadattam DSK. Processing of sapota. *J Food Technol.* 2005;3(3):326–330.
7. Shipra B, Kshipra D, Amla B, et al. Zingiber Officinale: Chemical and phytochemical screening and evaluation of its antimicrobial activities. *J Chem Pharm Res.* 2012;4(1):360–364.
8. Subir KD, Vasudevan DM. Tulsi the Indian Holy Power plant. *Nat Prod Rad.* 2005;5(4):279–283.
9. Oliveira MES, Pantoja L, Duarte WF, et al. Fruit wine produced from cagaita (*Eugenia dysenterica* DC) by both free and immobilised yeast cell fermentation. *Food Res Int.* 2011;44(7):2391–2400.
10. Miller GL. Use of dinitrosalicylic acid reagent for determination of reducing sugar. *Anal Chem.* 1959;31(3):426–428.
11. Jean LJ. *Introduction to wine laboratory practices and procedures.* 1st ed. USA: Springer Science Business Media Inc; 2006.
12. Vilela-Moura A, Schuller D, Falco V, et al. Effect of refermentation conditions and micro-oxygenation on the reduction of volatile acidity by commercial *S. cerevisiae* strains and their impact on the aromatic profile of wines. *Int J Food Microbiol.* 2010;141(3):165–172.
13. Marijan S, Ivana N, Lidija J. Determination of polyphenols content and antioxidant activity of some red wines by differential pulse voltammetry, HPLC and spectrophotometric methods. *Food Chem.* 2011;124(3):1208–1216.
14. Kocher GS. Status of wine production from guava (*Psidium guajava* L.):A traditional fruit of India. *Afr J Food Sci African Journal of Food Science.* 2011;5(16):851–860.