

# Passion fruit (*passiflora edulis*) seed cake as a feed ingredient for jaraqui (*semaprochilodus insignis*) and tambaqui (*colossoma macropomum*)

## Abstract

Passion fruit seed cake was evaluated as a fish feed ingredient at graded levels (0,5,10,15,20%), employing jaraqui and tambaqui fingerlings. Jaraqui of av. length 3.12-3.21 cm and av. wt. 2.59-2.76 g when fed on these diets attained a final av. length of 9.80-10.51 cm and av. wt. of 22.75- 26.11g at the end of 120 days, while tambaqui of initial av. length 5.06-5.16 cm and av. wt. 4.65-4.74 g grew to a final av. length of 15.28-16.76 cm and av. wt. of 81.22-118.34 g. A progressive decline in protein and an increase in lipid contents were noticed in the formulated diets with increasing passion fruit seed cake incorporation. Replacement of fishmeal with passion fruit seed cake in the test diets did not affect jaraqui growth at all the levels tested, whereas tambaqui growth was affected at higher levels (15 and 20%). The apparent digestibility values of dry matter, protein and lipid in the diets did not differ significantly. Diets affected fish carcass composition. The highest protein and lipid values were recorded with T<sub>0</sub> and T<sub>5</sub> diets that had 30.15% protein and 5.33% lipid respectively. While FCR did not vary significantly between treatments, PER of fish receiving diet T<sub>5</sub> was significantly higher with both the species of fish. The results show that the diets developed using passion fruit seed cake can be used in the culture of jaraqui and tambaqui with economic advantage.

**Keywords:** Fish meal substitution, Passion fruit, Jaraqui, Tambaqui, Growth, Carcass composition

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## Introduction

*Passiflora edulis* (passion fruit) belongs to the family Passifloraceae and is cultivated commercially in tropical and subtropical areas, mainly for the edible fruit. Brazil stands out as the world's largest producer of passion fruit, producing approximately 9,20,000 tonnes.<sup>1</sup> Locally known as maracujá, passion fruit is rich in vitamin C, calcium and phosphorus. The waste, including the seeds, that represents over 60% of the fruit is usually discarded after extraction of juice from the fruit, and is generally treated as organic waste.<sup>2</sup> An alternative to by-products from passion fruit industrialization would be their use in animal feed. The seeds of passion fruit are rich in fiber, minerals and lipids, with good amount of protein.<sup>3-5</sup> Oil is extracted from the seeds as they contain over 23% oil, rich in PUFA. The resultant by-product oilcake can form a useful feed ingredient. The shortage and rising cost of commercial fish meal has prompted researchers to investigate the production of cost effective feed formulations for cultured fish species.<sup>6,7</sup> Several plant protein sources/agricultural by-products have been tested as replacement for fish meal in fish diets<sup>8-14</sup> with a view to achieve cost reduction in fish production. However, studies with passion fruit seed cake are lacking. Therefore, it was felt worthwhile exploring the suitability of this cheaper nutrient source as a fish feed ingredient.

The two test species of fish were selected based on their economic importance. Jaraqui plays an important social role by catering to the needs of low income population in the Amazon, accounting for approximately 50% of fish landings in the port of Manaus.<sup>15</sup> Tambaqui is one of the most popular cultured species in the Amazon, representing almost half of the total fish sold in Manaus.<sup>16</sup> The present study aimed at investigating the feasibility of utilizing passion fruit seed cake as a fish feed ingredient, replacing the fish meal component at different levels.

## Materials and methods

Two experiments were carried out in the wet laboratory of the Coordination of Research in Aquaculture (CPAQ) of National Institute of Research in the Amazon (INPA), employing jaraqui and tambaqui seed obtained from a local farm in Manaus. The seed were separately acclimated for 15 days in two 1000 L tanks, hand feeding them to satiation with the control diet twice daily. The percentage of feeding was standardized at 4% based on the consumption during the acclimatisation period.

### Diets

Oil extracted passion fruit (*Passiflora edulis*) seed cake was procured locally; it contained 14.47% protein and 9.89% lipid. Five experimental diets (T<sub>0</sub>-T<sub>5</sub>) were formulated incorporating 0, 5, 10, 15 and 20% seed cake, by replacing the fish meal component (Table 1). The diet without seed cake (T<sub>0</sub>) served as the control. The feed mixture was hand kneaded with 300 ml water per kg and processed through a pelletiser to obtain pellets of 2 mm diameter. They were dried in a thermostatic oven at 40°C, packed in air-tight plastic bags and kept at room temperature until use.

Values with the same superscript in each row are not statistically different (P> 0.05).

### Experimental set-up

The experiments were conducted in a flow through system consisting of 15 circular tanks of 500 L each, containing 300 L of water. In the first experiment, 20 fingerlings of jaraqui (*Semaprochilodus insignis*) of av. length 3.12-3.21 cm and av. wt. 2.59-2.76 g were stocked, while in the second experiment tambaqui (*Colossoma macropomum*) fingerlings (av. length 5.06-5.16 cm and av. wt. 4.65-4.74 g) were

stocked at 20 per tank. The tanks were continuously aerated from a central aerator, using one aerator stone per tank. The fish were fed twice daily (09.00 hr and 16.00 hr) six days a week, at 4% of body weight in 2 equal halves, over the experimental duration of 120 days.

Fish length, weight and total biomass were assessed through sampling every 15 days and the quantity of feed given was readjusted based on the total fish biomass at each sampling.

**Table 1** Ingredient proportion and proximate composition (%) of diets

Ingredients	T <sub>0</sub>	T <sub>1</sub>	T <sub>2</sub>	T <sub>3</sub>	T <sub>4</sub>
Fishmeal	25	20	15	10	5
Soybean meal	20	20	20	20	20
Wheat bran	14	14	14	14	14
Wheat flour	20	20	20	20	20
Maize	18	18	18	18	18
Passion fruit seed cake	0	5	10	15	20
Vitamin-mineral premix*	2	2	2	2	2
Chromic oxide	1	1	1	1	1
<b>Proximate composition (Mean± SD, %)</b>					
Moisture	9.30± 0.04 <sup>b</sup>	8.78± 0.08 <sup>a</sup>	9.34± 0.06 <sup>b</sup>	9.62± 0.03 <sup>b</sup>	8.96± 0.09 <sup>a</sup>
Crude protein	30.15± 0.09 <sup>e</sup>	28.30± 0.14 <sup>d</sup>	27.18± 0.12 <sup>c</sup>	26.64± 0.16 <sup>b</sup>	25.35± 0.18 <sup>a</sup>
Lipid	4.76± 0.06 <sup>a</sup>	4.89± 0.07 <sup>a</sup>	5.13± 0.02 <sup>b</sup>	5.24± 0.05 <sup>b</sup>	5.33± 0.03 <sup>b</sup>
Ash	10.16± 0.10 <sup>a</sup>	10.22± 0.08 <sup>a</sup>	10.68± 0.05 <sup>b</sup>	11.03± 0.16 <sup>c</sup>	11.21± 0.09 <sup>c</sup>
Crude fibre	11.88±0.08 <sup>a</sup>	11.97±0.11 <sup>a</sup>	12.11±0.14 <sup>ab</sup>	12.32±0.10 <sup>b</sup>	12.43±0.18 <sup>b</sup>
NFE	33.75	35.84	35.56	35.15	36.72
Gross energy (Kcal.g <sup>-1</sup> )	353.69	353.08	347.87	344.18	344.21

\*kg mixture contains Vitamins: 6000000 IU A, 5000 mgB<sub>1</sub>, 1120 mg B<sub>2</sub>, 30000 mg B<sub>3</sub>, 30000 mg B<sub>5</sub>, 8000 mg B<sub>6</sub>, 2000 mg B<sub>9</sub>, 3 000 mg B<sub>12</sub>, 500 mgC, 2250000 IU D<sub>3</sub>, 3000 mg K<sub>3</sub>, 75000 mg E. Minerals: 150000 mg ZnSO<sub>4</sub>, 60000 mg MnSO<sub>4</sub>, 4500 mg KI, 100000 mgFeSO<sub>4</sub>, 2000 mgCoSO<sub>4</sub>, 400 mg Na<sub>2</sub>SeO<sub>3</sub>

## Water quality monitoring

Water quality parameters viz. dissolved oxygen (DO), electrical conductivity (EC), temperature and pH were measured on a weekly basis, whereas alkalinity, free carbon dioxide (CO<sub>2</sub>), nitrite nitrogen (NO<sub>2</sub>) and total ammonia (NH<sub>3</sub>) were analysed every 15 days. A combined digital YSI 85 meter (YSI incorporated Yellow Springs, Ohio, USA) was used to monitor DO and EC; temperature and pH were measured with a digital YSI 60 meter. Alkalinity, CO<sub>2</sub>, NO<sub>2</sub> and NH<sub>3</sub> were estimated following standard procedures.<sup>17</sup>

## Digestibility measurement

On termination of the growth experiments, 5 fish each from the replicate tanks were held in 15 cylindrical 200 L fibre glass tanks and fed once daily at 09.00 hr. with the respective 5 diets *ad libitum* in triplicate for 30 days, 6 days per week. Faecal matter was collected every morning and then dried in an oven. The faecal matter collected over the entire period from the respective tanks was pooled and analysed for proximate composition, with chromic oxide as the marker.

## Chemical analyses

Ingredients, diets, faecal samples and fish carcass were analysed for proximate composition as follows.<sup>18</sup> Moisture content by oven drying at 105°C for 24 h; crude protein (Nx6.25) by micro Kjeldal digestion and distillation after acid digestion using a Kjeltex 1026 distilling unit together with a Tecator digestion system (Tecator, Sweden); lipid by Soxhlet extraction; crude fibre by acid/alkali digestion; ash by ignition at 550°C in a muffle furnace to constant weight. Nitrogen-free extract (NFE) was computed by subtracting the sum values of crude protein, lipid, ash, crude fibre and moisture from 100.<sup>19</sup> Gross energy was calculated by using the conversion factors 5.64, 9.44 and 4.11 Kcal/g for protein, lipid and NFE<sup>20</sup> respectively. Chromic oxide content in the diets and faeces was determined by the acid digestion method<sup>21</sup> using absorption spectrophotometer. Three fish from each tank were sampled on termination of the growth experiments for carcass proximate analysis.

## Growth and feed utilization

Fish performance in terms of specific growth rate (SGR), feed conversion ratio (FCR), protein efficiency ratio (PER) and apparent nutrient digestibility was calculated using the formulae:

Specific growth rate

where Wt is weight of fish at time t, Wi is weight of fish at time 0, and T is the culture period in days

Feed conversion ratio (FCR) = Feed consumed (g) / Weight gain (g)

Protein efficiency ratio (PER) = Weight gain (g) / Protein intake (g)

Apparent nutrient digestibility (%) = 100 - 100[100 x (Id / If x Nf / Nd)]

Where Id represents chromic oxide in diet and If, chromic oxide in faeces, Nd is nutrient in diet, and Nf, nutrient in.

## Statistical analysis

The data were analysed by one-way analysis of variance (ANOVA) using Origin 6.1 software. Mean differences between treatments were tested for significance at P<0.05 by Tukey test and comparison was made by Duncan's multiple range test.<sup>22</sup>

## Results

### Water quality

Water quality parameters varied narrowly between treatments during the experimental period. The range of values were: water temperature 25.56-26.21/26.12-26.81°C, dissolved oxygen 7.20-7.38/7.33-7.65mg/L, pH 5.10-5.33/5.22-5.64, conductivity 29.24-32.38/28.12-34.52 μS/cm<sup>2</sup>, free CO<sub>2</sub> 0.94-1.23/0.89-1.37 mg/L, alkalinity 6.42-8.56/6.18-9.03 mg CaCO<sub>3</sub>/L, NO<sub>2</sub> 0.17-0.22/0.15-0.24 mg/L and NH<sub>3</sub> 0.09-0.21/0.07-0.19 mg/L in experiments 1 and 2.

### Feed

Fish meal replacement in the formulated diets with passion fruit seed cake resulted in a progressive decline in protein and an increase

in lipid contents. Moisture and ash levels differed significantly ( $P < 0.05$ ) between diets (Table 1). However, despite the lower protein level, diets with passion fruit seed cake had almost similar energy content as that of the control; the values ranged from 344.18 ( $T_4$ ) to 353.69 Kcal/g ( $T_0$ ).

### Fish growth

On termination of the growth experiment, final av. length of jaraqui varied from 9.80 ( $T_4$ ) to 10.51 cm ( $T_1$ ) and av. wt. from 22.75 ( $T_3$ ) to 26.11 g ( $T_2$ ), while tambaqui fingerlings grew to a final av. length of 15.28 ( $T_3$ )-16.76 cm ( $T_0$ ) and av. wt. of 81.22 ( $T_3$ )-118.34 g ( $T_0$ ), there being no significant difference between any of the treatments with the former species. However, growth of tambaqui under  $T_3$  and

$T_4$  treatments was significantly lower compared to  $T_0$  and  $T_1$  treatments. Fish survival was 100% in all the treatments. FCR varied from 2.13 to 2.25 in the case of jaraqui and 2.23 to 2.35 with tambaqui. PER ranged from 1.22 to 1.47 with jaraqui and 1.15 to 1.38 in the case of tambaqui (Tables 2 & 3).

### Feed digestibility and carcass composition

The apparent digestibility values of protein, dry matter and lipid did not differ significantly between fish receiving different diets. But, fish carcass composition was affected by the diets in both the species. The values (%) of protein and lipid ranged from 60.37 to 62.47 and 9.15 to 10.68 in jaraqui (Table 2) and 59.33 to 61.56 and 10.35 to 11.85 in the case of tambaqui (Table 3).

**Table 2** Growth parameters, diet digestibility and body composition of jaraqui (Mean± SD, Experiment 1)

Treatment Parameter	$T_0$	$T_1$	$T_2$	$T_3$	$T_4$
Initial mean length (cm)	3.21± 0.14 <sup>a</sup>	3.24± 0.11 <sup>a</sup>	3.12± 0.10 <sup>a</sup>	3.16± 0.16 <sup>a</sup>	3.18± 0.09 <sup>a</sup>
Final mean length (cm)	10.51±0.92 <sup>a</sup>	10.37±0.84 <sup>a</sup>	10.18±0.59 <sup>a</sup>	9.80±0.77 <sup>a</sup>	10.23±0.46 <sup>a</sup>
Initial mean wt. (g)	2.76±0.27 <sup>a</sup>	2.67±0.21 <sup>a</sup>	2.74±0.12 <sup>a</sup>	2.71±0.17 <sup>a</sup>	2.59±0.27 <sup>a</sup>
Final mean wt. (g)	25.37±2.29 <sup>a</sup>	26.11±3.42 <sup>a</sup>	25.44±2.18 <sup>a</sup>	23.33±3.10 <sup>a</sup>	22.75±2.95 <sup>a</sup>
Net wt. gain (g)	22.61±1.98 <sup>a</sup>	23.44±2.09 <sup>a</sup>	22.70±1.83 <sup>a</sup>	20.62±2.21 <sup>a</sup>	20.16±2.02 <sup>a</sup>
SGR (% day <sup>-1</sup> )	1.85±0.09 <sup>a</sup>	1.90±0.04 <sup>a</sup>	1.86±0.05 <sup>a</sup>	1.79±0.02 <sup>a</sup>	1.71±0.10 <sup>a</sup>
FCR	2.13±0.10 <sup>a</sup>	2.17±0.13 <sup>a</sup>	2.15±0.07 <sup>a</sup>	2.21±0.11 <sup>a</sup>	2.25±0.09 <sup>a</sup>
PER	1.22±0.06 <sup>a</sup>	1.30±0.09 <sup>ab</sup>	1.34±0.11 <sup>ab</sup>	1.39±0.10 <sup>ab</sup>	1.47±0.12 <sup>b</sup>
<b>Feed Digestibility (Mean± SD, %)</b>					
Dry matter	89.15±0.52 <sup>a</sup>	88.24±0.45 <sup>a</sup>	89.06±0.57 <sup>a</sup>	88.52±0.63 <sup>a</sup>	88.34±0.70 <sup>a</sup>
Protein	90.06±0.65 <sup>a</sup>	90.12±0.71 <sup>a</sup>	90.14±0.45 <sup>a</sup>	89.22±0.78 <sup>a</sup>	89.33±0.62 <sup>a</sup>
Lipid	92.10±0.45 <sup>a</sup>	92.25±0.23 <sup>a</sup>	91.66±0.52 <sup>a</sup>	92.23±0.31 <sup>a</sup>	91.44±0.54 <sup>a</sup>
<b>Carcass Composition on Dry Weight Basis (Mean± SD, %)</b>					
Dry matter	88.36±0.25 <sup>a</sup>	89.12±0.16 <sup>b</sup>	88.23±0.31 <sup>a</sup>	89.35±0.32 <sup>b</sup>	88.48±0.12 <sup>a</sup>
Crude Protein	62.47±0.48 <sup>b</sup>	61.18±0.54 <sup>a</sup>	62.45±0.35 <sup>b</sup>	60.39±0.29 <sup>a</sup>	60.37±0.33 <sup>a</sup>
Lipid	9.15±0.10 <sup>a</sup>	9.26±0.06 <sup>a</sup>	9.72±0.15 <sup>b</sup>	10.25±0.09 <sup>c</sup>	10.68±0.08 <sup>d</sup>
Ash	13.29±0.18 <sup>a</sup>	13.67±0.27 <sup>a</sup>	14.18±0.12 <sup>b</sup>	14.41±0.23 <sup>bc</sup>	14.74±0.15 <sup>c</sup>

Values with the same superscript in each row are not statistically different ( $P > 0.05$ ).

**Table 3** Growth parameters, diet digestibility and body composition of tambaqui (Mean± SD, Experiment 2)

Treatment Parameter	$T_0$	$T_1$	$T_2$	$T_3$	$T_4$
Initial mean length (cm)	5.16±0.21 <sup>a</sup>	5.06±0.24 <sup>a</sup>	5.14±0.12 <sup>a</sup>	5.09±0.16 <sup>a</sup>	5.11±0.15 <sup>a</sup>
Final mean length (cm)	16.76±0.82 <sup>a</sup>	16.24±0.68 <sup>a</sup>	16.08±0.42 <sup>a</sup>	15.93±0.54 <sup>a</sup>	15.28±0.76 <sup>a</sup>
Initial mean wt. (g)	4.67±0.15 <sup>a</sup>	4.71±0.11 <sup>a</sup>	4.74±0.09 <sup>a</sup>	4.69±0.12 <sup>a</sup>	4.65±0.16 <sup>a</sup>
Final mean wt. (g)	118.34±7.14 <sup>b</sup>	114.85±9.21 <sup>b</sup>	100.38±7.44 <sup>ab</sup>	92.53±6.40 <sup>a</sup>	81.22±8.46 <sup>a</sup>
Net wt. gain (g)	113.67±8.32 <sup>b</sup>	110.14±6.14 <sup>b</sup>	95.64±8.21 <sup>ab</sup>	87.84±7.12 <sup>a</sup>	76.57±7.28 <sup>a</sup>
SGR (% day <sup>-1</sup> )	2.69±0.17 <sup>a</sup>	2.66±0.05 <sup>a</sup>	2.54±0.07 <sup>a</sup>	2.48±0.05 <sup>a</sup>	2.38±0.11 <sup>a</sup>
FCR	2.23±0.08 <sup>a</sup>	2.26±0.14 <sup>a</sup>	2.30±0.11 <sup>a</sup>	2.33±0.09 <sup>a</sup>	2.35±0.12 <sup>a</sup>
PER	1.15±0.07 <sup>a</sup>	1.22±0.10 <sup>ab</sup>	1.26±0.05 <sup>ab</sup>	1.33±0.11 <sup>ab</sup>	1.38±0.08 <sup>b</sup>
<b>Feed Digestibility (Mean± SD, %)</b>					
Dry matter	90.12±0.67 <sup>a</sup>	90.24±0.46 <sup>a</sup>	89.32±0.65 <sup>a</sup>	89.25±0.58 <sup>a</sup>	89.46±0.47 <sup>a</sup>
Crude protein	90.16±0.62 <sup>a</sup>	90.02±0.78 <sup>a</sup>	89.25±0.45 <sup>a</sup>	89.32±0.52 <sup>a</sup>	89.78±0.54 <sup>a</sup>
Lipid	91.32±0.45 <sup>a</sup>	91.25±0.49 <sup>a</sup>	90.26±0.56 <sup>a</sup>	90.44±0.42 <sup>a</sup>	90.12±0.65 <sup>a</sup>
<b>Carcass Composition on Dry Weight Basis (Mean± SD, %)</b>					
Dry matter	89.31±0.38 <sup>b</sup>	90.12±0.56 <sup>bc</sup>	88.23±0.35 <sup>a</sup>	90.35±0.28 <sup>c</sup>	89.48±0.22 <sup>b</sup>
Crude protein	61.56±0.32 <sup>b</sup>	60.97±0.25 <sup>b</sup>	60.11±0.34 <sup>a</sup>	59.79±0.24 <sup>a</sup>	59.33±0.38 <sup>a</sup>
Lipid	10.35±0.15 <sup>a</sup>	10.62±0.08 <sup>a</sup>	11.25±0.18 <sup>b</sup>	11.56±0.11 <sup>b</sup>	11.85±0.05 <sup>c</sup>
Ash	14.23±0.18 <sup>a</sup>	14.54±0.25 <sup>a</sup>	15.27±0.16 <sup>b</sup>	15.45±0.20 <sup>b</sup>	15.61±0.19 <sup>b</sup>

Values with the same superscript in each row are not statistically different ( $P > 0.05$ ).

## Discussion

The water quality parameters monitored during the two experiments were within the acceptable limits for fish culture as has been reported by earlier researchers,<sup>23,24</sup> varying narrowly between treatments and without any drastic fluctuations. Inadequate conditions of water

quality affect growth, reproduction, health, survival and quality of fish life, jeopardizing the success of aquaculture.<sup>25</sup> Water temperature recorded in the present study was in the range 25.56-26.21°C. Jaraqui is found in nature in both lotic and lentic ecosystems, where the minimum temperature is 24°C and maximum 40°C, indicating high temperature tolerance.<sup>26</sup> The optimal level of oxygen for tropical

fish species ranges from 4 to 6 mg/L.<sup>27</sup> Tambaqui has morphological adaptations to survive in hypoxic environments and can survive in waters with less than 1 mg O<sub>2</sub>/L.<sup>28,29</sup> Dissolved oxygen was above 7 mg/L throughout this study; however, pH remained slightly acidic. In nature, jaraqui tolerates large ionic plasticity and survives well even in acidic waters.<sup>26</sup> Low alkalinity of 10 mg/L has been reported in tambaqui production systems in the Amazon water when liming is not done.<sup>30</sup> In the present study, alkalinity was relatively low because of the flow through system. For fish, nitrite is toxic in water when the concentration crosses 0.5 mg/L<sup>31</sup> and ammonia, when higher than 2 mg/L.<sup>24</sup> Nitrogen (NO<sub>2</sub> and NH<sub>3</sub>) and free CO<sub>2</sub> levels were low in this study (Table 1) and hence would not have adversely influenced fish growth.

In the test diets, a progressive decline in protein and an increase in lipid contents were noticed due to passion fruit seed cake supplementation which can be attributed to its proximate composition. It had 14.47% protein and 9.89% lipid as against 52.61% protein and 6.72% lipid in fish meal. Moisture and ash levels differed significantly (P<0.05) between diets. However, their energy content did not vary drastically, despite the lower protein level in diets incorporated with passion fruit seed cake (Table 1).

The best growth of jaraqui was obtained with T1 diet (26.11 g) at the end of 120 days of the feeding experiment, there being no significant difference between treatments. Tambaqui recorded the highest growth with the control (T0) diet (118.34 g); its growth under T3 and T4 treatments was significantly lower compared to T0 and T1 treatments. Genetically, these two species of fish have different growth rates. Jaraqui is a slow growing fish, while tambaqui is a fast growing one. Thus, tambaqui attained 4 times the growth of jaraqui on termination of the feeding experiment. Varied response of the two species to the test diets reflects the difference in feed utilization by them. Increasing replacement of fish meal impacted the quality of the diets due to declining protein levels. However, jaraqui growth was unaffected even with 20% substitution of fish meal with equal amount of passion fruit seed cake; only marginal decrease in its growth was observed under T3 and T4 treatments (Table 2). Thus, 25% protein with 5% fish meal was found sufficient for jaraqui. In tambaqui, 27% protein with 15% fish meal appears desirable, since with lesser protein and fish meal levels its growth was significantly lower (Table 3). Fishes utilize fish meal very well from the diet as its nutrient content is very close to their body composition. Fish meal is a well balanced source of high quality protein. It increases feed efficiency and growth through better food palatability and higher nutrient uptake, digestion, and absorption. Further, it is an effective feed attractant<sup>32</sup> and contains some unknown factors<sup>33,34</sup> which enhance fish growth. Cent percent survival recorded in all the treatments indicates that the fish received sufficient amount of nutrients from the diets provided and the quality of water was conducive.

FCR was marginally better with jaraqui, compared to tambaqui. In contrast, PER was marginally superior with tambaqui than jaraqui, indicating the former's ability to utilize protein more efficiently from artificial diets. While FCR did not vary significantly between treatments, PER of fish receiving diet T5 was significantly better with both the species, compared to the control (T0) diet (Tables 2 & 3). This reflects better utilization of protein from diets containing lower protein level. The protein level in T0 and T4 diets was 30.15 and 25.35% respectively. Higher protein utilization from low protein diets has been reported earlier with other species.<sup>35,38</sup> Better FCR and PER was obtained in salema porgy (*Sarpa salpa*) juveniles receiving lower protein (37 and 30%) diets than higher protein (40-57%) diets.<sup>39</sup>

Even though passion fruit seed is known to contain antinutrients like phytates and oxalates,<sup>40</sup> the apparent digestibility values of dry matter, protein and lipid in the diets did not differ significantly with the two fish species. Fish carcass composition was affected by the diets. In jaraqui, the highest protein (62.47%) and lipid (10.68%) values were recorded with T0 and T5 diets, reflecting the influence of dietary nutrients on body composition. Similar was the case with tambaqui which recorded the highest protein (61.56 %) and lipid (11.85%) levels with these diets. The influence of diets on fish carcass composition is well documented.<sup>41-43</sup>

Based on the present results, it may be concluded that passion fruit seed cake can be utilized as a feed ingredient in the diets of jaraqui and tambaqui to the extent of 20 and 10% respectively, by substituting equal amount of fish meal, without affecting the growth performance of fish. The findings have economic significance.

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## Conflicts of interest

None.

## References

1. Instituto Brasileiro de Geografia e Estatística (IBGE). Produção Agrícola Municipal-PAM. Banco de Dados SIDRA. Disponível. 2012.
2. Silva RM, Placido GR, Silva MAP, et al. Chemical characterization of passion fruit (*Passiflora edulis* f. *flavicarpa*) seeds. *African Journal of Biotechnology*. 2015;14:1230-1233.
3. Chau CF, Huang YL. *Characterization of passion fruit seed fibres – a potential fibre source*. *Food Chemistry*. 2004;85(2):189-194.
4. Liu S, Yang F, Li J, et al. Physical and chemical analysis of *Passiflora* seeds and seed oil from China. *Int J Food Sci Nutr*. 2008;59(7-8):706-715.
5. Oliveira SEM, Regis AS, Resende ED. Caracterização dos resíduos da polpa do maracujá-amarelo. *Ciencia Rural*. 2011;41(4):725-730.
6. Nyina-Wamwiza L, Wathlet B, Richir J, et al. Partial or total replacement of fish meal by local agricultural byproducts in diets of juvenile African catfish (*Clarias gariepinus*): growth performance, feed efficiency and digestibility. *Aquaculture Nutrition*. 2010;16(3):237-247.
7. Msangi S, Kobayashi M, Batka M, et al. Fish to 2030. *Prospects for Fisheries and Aquaculture*. Washington, USA: The World Bank Publication, Report No 2013;83177-GLB.
8. Olvera-Novoa MA, Martinez PCA, Galvan CA, et al. The use of the leguminous plant *Sesbania grandiflora* as a partial replacement for fish meal in diets for tilapia (*Oreochromis mossambicus*). *Aquaculture*. 1988;71(1-2):51-60.
9. El-Sayed AFM. Alternative dietary protein source for farmed tilapia, *Oreochromis Aquaculture*. 1999;179(1-4):149-168.
10. Siddhuraju P, Becker K. Preliminary nutritional evaluation of *Mucuna* seed meal (*Mucuna pruriens* var. *utilis*) in common carp (*Cyprinus carpio* L.): An assessment by growth performance and feed utilization. *Aquaculture*. 2001;196(1-2):105-123.
11. Ogunji JO. Alternative protein sources in diets for farmed tilapia. *CAB International Publishing* (Oxford, UK). Nutrition Abstracts and Reviews: Series B. 2004;74:23-32.

12. Bake GG, Adejumo TM, Sadiku SOE. Growth performance and nutrient utilization of Nile Tilapia (*Oreochromis niloticus*) fed toasted flamboyant seed meal (*Delonix regia*). Continental Journal of Agricultural Science. 2013;7(1):1–10.
13. De Santis C, Ruohonen K, Tocher DR, et al. Atlantic salmon (*Salmo salar*) parr as a model to predict the optimum inclusion of air classified faba bean protein concentrate in feeds for seawater salmon. Aquaculture. 2015;444(1):70–78.
14. Cai CW, Jiang GZ, Li XF, et al. Effects of complete fish meal replacement by rice protein concentrate with or without lysine supplement on growth performance, muscle development and flesh quality of blunt snout bream (*Megalobrama amblycephala*). Aquaculture Nutrition. 2017.
15. Gandra ALO. Mercado do pescado da região metropolitana de Manaus. CFC/FAO/INFOPECA. 2010;pp.84.
16. Araujo-Lima C, Goulding M. Os frutos do tambaqui: ecologia, conservação e cultivo na Amazônia. Tefê, AM: Sociedade Civil de Mamirauá. 1998;186.
17. APHA. Standard Methods for Examination of the Water and Waste Water. (18<sup>th</sup> edn), American Public Health Association, USA. 1992.
18. AOAC. Official Methods of Analysis. (16<sup>th</sup> edn), Association of Official Analytical Chemists, USA. 1995.
19. Hastings WH. Fish nutrition and feed manufacture, Paper presented at FAO Technical Conference on Aquaculture, Kyoto, Japan. 1976;pp.13.
20. NRC (National Research Council). Nutritional Requirements of Fishes. National Academy Press, USA. 1993;pp.114.
21. Furukawa A, Tsukahara H. On the acid digestion method for the determination of chromic oxide as an index substance in the study of digestibility of fish feed. Bulletin of the Japanese Society for Scientific Fisheries. 1966;32:502–506.
22. Duncan DB. Multiple range and multiple F-tests. Biometrics. 1955;11:1–42.
23. Boyd CE. Water quality in warm water fish culture. Auburn University, Alabama. 1981;pp.359.
24. Tavares LHS. Limnologia aplicada à Jaboticabal: FUNEP. 1994;pp.70.
25. Lourenco JNP, Malta JCO, Sousa FN. A importância de monitorar a qualidade da água na piscicultura, Instruções Técnicas Embrapa Ocidental. 1999;pp.1–4.
26. Araujo-Lima C, Goulding M. So fruitful a fish. Ecology, conservation and aquaculture of the Amazon's tambaqui. Columbia University Press, USA. 1997;pp.191.
27. Proença CEM, Bittencourt PRL. Manual de piscicultura tropical. IBAMA, M.M.A., Brasília. 1994;pp.196.
28. Saint-Paul U. Physiological adaptation to hypoxia of a neotropical characoid fish *Colossoma macropomum*, Serrasalminidae. Environmental Biology of Fishes. 1984;11(1):53–62.
29. Saint-Paul U. Diurnal routine O<sub>2</sub> consumption at different O<sub>2</sub> concentrations by *Colossoma macropomum* and *Colossoma brachypomum* (Teleostei: Serrasalminidae). Comparative Biochemistry and Physiology. 1988;89A:675–682.
30. Aride PHR, Roubach R, Val AL. Water pH in central Amazon and its importance for tambaqui (*Colossoma macropomum*) culture. World Aquaculture Magazine, Baton Rouge. 2004;35(2):24–28.
31. Ostrensky A, Boeger W. Piscicultura: Fundamentos e técnicas de manejo – Guaíba, Agropecuária. 1998;pp.211.
32. Hertrampf JW, Piedad-Pascual F. Unidentified growth factors. In: Handbook on Ingredients for Aquaculture Feeds. Springer, Netherlands. 2000.
33. Potter LM, Shelton JR, Parsons CM. The unidentified growth factor in Menhaden fish meal. Poultry Science. 1980;59(1):128–134.
34. Fox JM, Lawrence AL, Smith F. Development of a low-fish meal feed formulation for . Advances en Nutricion Acuicola VII. Memorias del VII Simposium Internacional de Nutricion Acuicola. Noviembre, Hermosillo, Sonora, Mexico. 2004.
35. Ahmad MH, Abdel-Tawwab M, Khattab YAE. Effect of dietary protein levels on growth performance and protein utilization in Nile tilapia (*Oreochromis niloticus*) with different initial body weights. Proceedings of Sixth International Symposium on Tilapia in Aquaculture. Manila, Philippines. 2004;pp.249–263.
36. Shalaby SM, El-Dakar AY, Wahbi OM, et al. Growth, feed utilization and body composition of white sea bream *Diplodus sargus juveniles* offered diets with various protein and energy levels. Marine Science. 2011;22(2):3–17.
37. Abbas G, Siddiqui PJA. The effect of varying dietary protein level on growth, feed conversion, body composition and apparent digestibility coefficient of juvenile mangrove red snapper, *Lutjanus argentimaculatus* (Forsskal 1775). Aquaculture Research. 2013;44(5):807–818.
38. Daudpota AM, Siddiqui PJA, Abbas G, et al. Effect of dietary protein level on growth performance, protein utilization and body composition of Nile tilapia cultured in low salinity water. International Journal of Interdisciplinary and Multidisciplinary Studies. 2014;2(2):135–147.
39. Sahinyilmaz M, Yigit M. Evaluation of protein levels in diets for salem porgy (*Sarpa salpa*) juveniles, a new candidate species for the Mediterranean aquaculture. Journal of Food and Nutrition Sciences. 2017;5(3):107–115.
40. Wasagu RSU, Lawal M, Amedu AM, et al. Comparative chemical analysis, phytochemical screening and antimicrobial activities of the rinds, seeds and juice of (*Passiflora edulis var. flavicarpa*) passion fruit. Journal of Natural Sciences Research. 2016;6(19):138–143.
41. Brett JR, Shelbourne JE, Shoop CT. Growth rate and body composition of fingerling sockeye salmon, *Oncorhynchus nerka* in relation to temperature and ration size. Journal of the Fisheries Research Board of Canada. 1969;26(9):2363–2394.
42. Usmani N, Jafri AK. Influence of dietary phytic acid on the growth, conversion efficiency, and carcass composition of mrigal *Cirrhinus mrigala* (Hamilton) fry. Journal of the World Aquaculture Society. 2002;33(2):199–204.
43. Araujo FG, Costa DV, Machado MRF, et al. Dietary oils influence ovary and carcass composition and embryonic development of zebrafish. Aquaculture Nutrition. 2016;23(4):651–661.