

Research Article





# Antioxidant, anti-inflammatory and anticancer activities of Tunisian halophyte Suaeda mollis

#### **Abstract**

The aim of this study was to investigate the antioxidant, anti-inflammatory, and anticancer properties of Tunisian Suaeda mollis specie after fractionation by increasing polarity of several extracts (Hexane, Dichloromerthane, Methanol and water). Water and methanol extracts of S. mollis exhibited the best in vitro antioxidant ability (2.92±0.19 and 2.77±0.21  $\mu$ mol TE/mg, respectively), while, dichloromethane extract displayed the highest ex vivo antioxidant activity with an IC $_{50}$  equal to 1.73  $\mu$ g/ml. Besides, hexane extract showed an important anti-inflammatory effect inhibiting NO with 55.9% at 160  $\mu$ g/ml. Finally, dichloromethane extract displayed a potent anticancer activity against cancerous cells A-549 with an IC $_{50}$  equal to 70±7  $\mu$ g/ml as well as non cancerous cells Detroit-551 and WS1 (IC $_{50}$ =81±1 and 62±5  $\mu$ g/ml, respectively) using resazurin test. Moreover, hexane extract is efficient against Caco-2 (IC $_{50}$ =122  $\mu$ g/ml). Overall, the results suggested that biological activities are influenced by solvent polarity and extractability.

**Keywords:** antioxidant activity, anti-inflammatory capacity, *Suaeda mollis*, anticancer ability

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**Abbreviations**: AAPH, 2,2-azobis(2-amidinopropane) dihydrochloride; DCFH, 2',7'- dichlorofluorescin; ATCC, American type culture collection; CE, catechin equivalent; DCFH-DA, dichlorofluoresceine-diacetate; DMSO, dimethyle sulphoxide; TE, equivalent trolox; DLD-1, Caco-2 and HT-29, human colorectal adenocarcinoma cell lines; A-549, human lung carcinoma cell lines; WS1 and Detroit 551, human skin fibroblast; RAW 264.7, mouse macrophage

#### Introduction

Halophyte Plants are a potent source of bioactive metabolites for several agroalimentary, cosmetic and pharmaceutical industries. Besides they act as an efficient barrier to increase or/and inhibit reactive oxygen species. These ROS are extremely accumulated under environmental constraints such as salinity and drought.

Besides, Halophytes are considered as a significant pool of natural compounds that can replace and substitute synthetic compounds. For this reason, researches and science have focused on the plants phytochemistry and the identification of natural molecules that are beneficial to human health. Oxidative stress is generated by reactive oxygen species surproduction and is defined as disequilibrium between antioxidant system and reactive oxygen species (ROS) generation. These ROS are extremely toxic leading an irreversible damage to proteins, lipids, DNA and can inducing cell alterations cell.<sup>2</sup> In this context, Plants stimulate their antioxidant system in order to alleviate the adverse effects generated after oxidative stress. Nevertheless, antioxidant ability depends on plant species, stress types and duration.<sup>3</sup> Besides among these plants, Halophytes manage oxidative stress via an endogenous protective system consisting of enzymatic (ascorbate peroxidase, glutathione reductase, superoxide dismutase, catalase, etc.) and non enzymatic systems (glutathione, vitamins, phenolic acids, flavonoids, tannins, etc.).4 In fact, 80% of these secondary metabolites are attributed for plants. These natural compounds are isolated from several flora species. They act as a potent and an efficient antioxidant to scavenge free radicals by hydrogen atom donation.<sup>5</sup> These compounds are known for their broad spectra of biological

activities as neuroprotective, anticancer, antimicrobial, hypoglycemic, anti-inflammatory, etc.<sup>6</sup> and they have more other functions compared to the synthetic drugs. These secondary metabolites are considered an efficient electron donor, provide as reduction agents, donors of hydrogen and singlet oxygen scavengers.<sup>7</sup> Also, polyphenols and flavonoids compounds are involved in the inhibition of hydrolytic and oxidative enzymes.<sup>8</sup> This oxidative disturbance may guide to other diseases such as inflammation and cancer. Increased attention has been given to plants which are rich in biochemical constituents and which are considered as natural agents to treat several diseases and to inhibit cancer cells growth.<sup>9</sup>

In Tunisia, an extensive diversity of edible and medicinal halophyte species used in traditional therapy were equipped with powerful antioxidant system which played an important role in plant physiology adaptation to environmental constraint, among them the edible *Suaeda fruticosa* exhibits hypoglycaemic and hypolipidaemic activities. <sup>10</sup> *Suaeda maritima* was used as a condiment for salad, and known for their hepatoprotective and antioxidant properties. <sup>11</sup> *Reaumuria vermiculata* was known for antioxidant and anticancer activities of Tunisian *Suaeda mollis*, although there are many reports for other *Suaeda* species. This study aimed to assess after fractionation using increasing solvent polarity, antioxidant ability via ORAC and cell based-assays, anti-inflammatory activity and anticancer potentiality of the Tunisian *Suaeda mollis*.

#### Materials and methods

#### Plant material and extract preparation

Shoots of *S. mollis* were collected from Draia (Gabes) (34°13′55″N,10°00′49″E). In order to assess biological activities, 500g of shoots in powder were extracted in a soxhlet apparatus using different solvents (hexane, dichloromethane, methanol and water) basing on polarity solvents. Then, a filtration was performed, after that, a solvent evaporation under reduced pressure using rotary vacuum evaporator was accomplished, and the residue was dissolved in DMSO ahead of analysis.



#### Assessment of antioxidant activities

#### Oxygen radical absorbance capacity (ORAC test)

The ORAC test was carried out in 96-well microplates on a Fluoroskan Ascent FLTM plate reader equipped with a programmed injector. For that, four concentrations of Trolox were used in quadruplicate, and a gradient of 16 doses of the samples was arranged lacking replication. The experiment was realized in controlled condition. A planification of fluorimeter has been assessed to confirm fluorescence of fluorescein each minute after adding 375 mM of AAPH during 35 min. ORAC values were expressed in µmol TE/g which trolox was used as a positive control by Girard-Lalancette.

# Antioxidant cell assay using 2',7'-dichlorofluorescin-diacetate (DCFH-DA)

Antioxidant activity was evaluated using the DCFH-DA assay by Legault. Human Skin Fibroblast cells were plated in 96 microwell plates and incubated under controlled condition (5 % CO<sub>2</sub> and at 37 °C). After that, phosphate buffer saline was used to rinse cells and an incubation of these cells in Hank's balanced salt solution with DCFH-DA has been carried out. After that, incubation during 1H of cells in contact with *Suaeda mollis* extracts was carried out after rinsing the cells. This experiment was carried out with or without tertbutylhydroperoxide. The fluorescence was measured directly after addition of t-BuOOH and after 1h30min at 485 and 530 nm.

#### Anti-inflammatory activity

RAW 264.7 cells growth was monitored after incubation in culture medium for controlled condition (16h, 37°C and 5%  $\rm CO_2$ ). After that, a treatment of these cells with lipopolysaccharide, extracts and positive control has been assessed. Then, microplates are incubated for 24h in order to liberate NO. The L-NAME was used as positive control.

Following that, supernatant was mixed with Griess reagent and incubated at room temperature for 20 min. Nitrite quantification was revealed using  ${\rm NaNO_2}$  standard curve. The absorbance reading was taken at 540 nm using a Varioskan Ascent. <sup>12</sup>

#### Cell culture

The human lung carcinoma A-549 (ATCC #CCL-185), colon adenocarcinoma DLD-1 (ATCC #CCL-221), human colorectal adenocarcinoma Caco2 (ATCC#HTB-37), HT-29 (ATCC#HTB-38) and murine macrophage RAW 264.7 (ATCC #TIB-71) cell lines were obtained from the American Type Culture Collection (ATCC, Manassas, USA). The WS-1 (ATCC#CRL-1502), Detroit-551 (ATCC#CCL-110), A-549, DLD-1, Caco2 and HT-29 cell lines were grown in Minimum Essential Medium with Earle's salts, while the RAW 264.7 cell line was grown in Dulbecco's modified Eagle's medium (Mediatech Cellgro®, Herndon, USA). Both media were supplemented with 10% fetal calf serum (Hyclone, Logan, USA) for (WS-1, Detroit-551, A-549, DLD-1 and HT-29) but with 20% fetal calf serum for Caco2, solution of vitamins (1×), sodium pyruvate (1 $\times$ ), non-essential amino acids (1 $\times$ ), penicillin (100 IU) and streptomycin (100 µg/ml) (Mediatech Cellgro®). Cells were cultured in a humidified atmosphere at 37°C in 5% CO<sub>2</sub>.

#### Cytotoxicity assay

Exponentially growing cells were plated at a density of  $5\times10^3$  cells per well in 96-well microplates (Costar, Corning Inc.) in 100  $\mu$ l of culture medium and were allowed to adhere for 16h before treatment. Then, 100  $\mu$ l of increasing concentrations of extract dissolved in the appropriate solvent (DMSO) were added. The final concentration of

solvent in the culture medium was maintained at 0.5% (v/v) to avoid solvent toxicity. The cells were incubated for 48h in the absence or in the presence of extract. Cytotoxicity was assessed using the resazurin reduction test as described by O'Brien et *al.* (2000). Fluorescence was measured on an automated 96-well Fluoroskan Ascent Fl<sup>TM</sup> plate reader (Labsystems) using an excitation wavelength of 530nm and an emission wavelength of 590nm. Cytotoxicity was expressed as the concentration of extract inhibiting cell growth by 50% (IC<sub>50</sub>).

#### Statistical analysis

The values were expressed as means  $\pm$  standard deviation of three determinations. The results were analyzed by a one-way analysis of variance (ANOVA) and Newman-Keuls multiple test. P values of 0.05 or less were considered as statistically significant.

#### **Results**

Table 1 indicated that methanol and water extracts showed the highest antioxidant activity, with ORAC values of  $2.77\pm0.21$  and  $2.29\pm0.19\mu$ mol TE/mg respectively, followed by dichloromethane extract ( $1.19\pm0.07~\mu$ mol TE/mg). While activity of hexane extract was less significant. Regarding *ex vivo* antioxidant ability results presented in Table 1 indicated that dichloromethane extract has powerfully antioxidant activity and inhibited the *t*BH-induced oxidation of DCFH with an IC<sub>50</sub> value equal to  $1.73\pm0.55~\mu$ g/ml. Moreover, methanol and water extracts displayed similar activity (IC<sub>50</sub> value was about 9  $\mu$ g/ml). Although, hexane extract showed less oxidation inhibition (IC<sub>50</sub> value =  $128~\mu$ g/ml). These findings demonstrated that *S. mollis* extracts mainly which are extracted with higher polarity possess an important antioxidant capacity.

**Table I** Oxygen radical absorbance capacity (ORAC) values and antioxidant cell assay expressed as  $IC_{50}$  of four extracts from *Suaeda mollis* shoots and standard. Each value represents the mean  $\pm$  SD of three determinations

Extracts	ORAC (µmol trolox/mg)	$IC_{50}$ (µg/ml)	
Hexane	0.08±0.01 <sup>d</sup>	128.05±65.3	
Dichloromethane	$1.19\pm0.07^{c}$	1.73±0.55°	
Methanol	2.77±0.21 <sup>b</sup>	9.00±1.71b	
Water	2.29±0.19b	9.27±2.59b	
Quercetin	19.59±2.2a	0.46±0.09a	

Anti-inflammatory activity was evaluated using LPS-stimulated RAW 264.7 macrophages. Stimulation of RAW 264.7 macrophages by LPS induces iNOS (inducible NO synthase) and overproduction of NO. Results presented in Table 2 showed that hexane extract exhibited appreciable inhibitory effect on LPS-induced NO secretion with 55.9 % at  $160\mu g/ml$ , followed by methanol and water extracts which inhibited NO release by 46.1 and 39.9%, respectively. However, dichloromethane extract showed cytotoxcicity at doses ranging from 2.5 to  $160 \mu g/ml$  (data not shown). Comparatively, the L-NAME, used as positive control inhibited NO release by 48.7 % at  $250.0 \mu M$  ( $67.4\mu g/ml$ ).

Furthermore, anticancer activity of *S. mollis* has not been shown in literature and the important antioxidant and anti-inflammatory activities in this species related to their phenolics prompted us to investigate their anticancer capacity. The cytotoxicity of the 4 shoot extracts was evaluated against colon carcinoma (DLD-1, Caco-2 and HT-29) and lung carcinoma (A-549) cell lines and health human skin fibroblast cell lines (WS1 and Detroit 551). Results displayed

that hexane extract of *S. mollis* was active against Caco-2 with an IC $_{50}$  value of 122±4 µg/ml and without any cytotoxicity. Moreover, dichloromethane extract showed an anticancer activity against A-549 and HT-29, although with a cytotoxicity against healthy cell lines (WS1 and Detroit 551) (Table 3). Nevertheless, methanol and water were inactive against all carcinoma cells. Consequently, *S. mollis* had an influence on viability of cells in hexane extracts that target carcinoma colon cells mainly Caco-2, without cytotoxicity against healthy cell lines. <sup>15–28</sup>

**Table 2** Anti-inflammatory activity of several extracts from *S. mollis* shoots expressed by NO inhibition (%)

Concentrations (µg/ml) Extracts	20	67.4	80	160	
Hexane	36.1	nd	48.1	55.9	
Dichloromethane	-	-	-	-	
Methanol	15.4	nd	30	46.1	
Water	9.5	nd	22.9	39.9	
L-NAME	nd	48.7	nd	nd	

nd: not determined and cytotoxicity represented by (-)

**Table 3** Cytotoxic activity of several extracts from *Suaeda mollis* shoots against two healthy cell lines (WS1 and Detroit 551) and four tumor cell lines (A-549, DLD-1, Caco-2 and HT-29)

Cell lines	WS1	Detroit	A-549	DLD-1	Caco-2	HT-29		
Extracts	IC <sub>50</sub> (µg/ml)							
Hexane	>200	>200	>200	>200	122±4	>200		
Dichloromethane	62±5	86±1	70±7	>200	>200	117±1		
Methanol	>200	>200	>200	>200	>200	>200		
Water	>200	66±1	>200	>200	>200	>200		
Etoposide (µM)	5±2	1±11	16.2±0.8	15±4	>50	22±2		

#### **Discussion**

The extraction, purification and biological activities of secondary metabolites depend on various factors including extraction time, temperature, solvent concentration and solvent polarity by Nawaz. In fact, according to chemical nature, diverse phytochemicals are extracted in solvents using various polarity because the combination of solvents is always effective than the use of single solvents by Iloki-Assanga. Besides, phenolic compound amount increased significantly with high polarity solvent and can rapidly decreased by solvent with higher polarity index such as water by Wakeel. Several studies have demonstrated that bioactivities are dependent on solvent polarity by Herrera-Pool. In fact, change in solvent polarity alters its ability to dissolve a selected group of antioxidant compounds and influences activity estimation.<sup>13</sup> Moreover, great intraspecific variations in plant secondary metabolites could be due to changes and fluctuations in soil composition and climate that involve the interaction between plants and the environment and eventually bioactive compounds by Kumar.

Phenolic compounds have been reported to be beneficial in the treatment of chronic inflammatory diseases associated with overproduction of nitric oxide (NO). For instance, previous studies showed that the major compound found in *S. mollis* extract is rutin also named quercetin-3-rutinoside, <sup>14</sup> this compound is known with a wide range of pharmacological properties such as antioxidant activity. Conventionally, it is used as antifungal, an antimicrobial and anti-

allergic agent by Al-Dhabi . Conversely, recent investigations have shown several pharmacological benefits to treat various diseases, such as cancer, diabetes, hypertension, and hypercholesterolemia by Sharma.

Besides, 4-hydroxybenzoic, syringic, rosmarinic acids are found in *S. molis* extracts.<sup>14</sup> These compounds have been considered in a number of cancer cell lines at different stages of cancer growth. In this regard, Jin found that rosmarinic acid can prevent tumorigenesis and reduce or inhibit tumor growth. Besides, in this context, the antiproliferative role of rosmarinic acid was exerted through apoptotis induction and arrest of cell cycle at G0/G1 and G2/M phases in oral cancer cells by Luo. A potent impact of rosmarinic acid have been demonstrated against several cell lines such as HeLa, HT29, A549 and MCF6 cancer cell lines by Erenler.

### Compliance with ethical standards

#### Conclusion

In the present study, biological activities including antioxidant, anti-inflammatory and anticancer potentialities of Tunisian *Suaeda mollis* shoots are demonstrated for the first time in four different solvents. Results showed that this specie has proven the potential benefits. Shoot extracts were found to be a potent antioxidant source and exhibit an important *in vitro* and *ex vivo* antioxidant activity. Moreover, this medicinal halophyte showed an important anti-inflammatory against macrophages. Besides, an efficient specificity on colon cancer has been demonstrated using several cell lines. These data suggest that this halophyte could be considered as a potential source of bioactive compounds with advantageous proprieties, suggesting its use in agroalimentary and medicine.

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#### **Conflict of interest**

The authors declare that they have no conflict of interest.

# **References**

- Hymery N, Dauvergne X, Boussaden H, et al. Evaluation of the Antioxidant, Anti-Inflammatory and Cytoprotective Activities of Halophyte Extracts against Mycotoxin Intoxication. *Toxins*. 2021;13:312.
- Ozgur R, Uzilday B, Sekmen AH, et al. Reactive oxygen species regulation and antioxidant defence in halophytes. Funct Plant Biol. 2013;40(9):832–847.
- 3. Hasanuzzaman M, Bhuyan MHMB, Zulfiqar F, et al. Reactive Oxygen Species and Antioxidant Defense in Plants under Abiotic Stress: Revisiting the Crucial Role of a Universal Defense Regulator. *Antioxidants*. 2020;9:681.
- Kaur N, Kaur J, Grewal SK, et al. Effect of Heat Stress on Antioxidative defense system and its amelioration by heat acclimation and salicylic acid pre-treatments in three pigeonpea genotypes. *Indian J Agric Biochem.* 2019;32:106–110.
- Thirumurugan D, Cholarajan A, Suresh SSR, et al. An Introductory Chapter: Secondary Metabolites. In R. Vijayakumar, & S. S. Raja (Eds.), Secondary Metabolites – Sources and Applications. *Intech Open*. 2018.

- Pérez–González MZ, Gutiérrez–Rebolledo GA, Jiménez–Arellanes MA, et al. The chemical constituents and biological activities of *Cnidos-colus chayamansa* McVaugh, a Mexican medicinal species, and plant cell cultures for the production of bioactive secondary metabolites. *Studies in Natural Products Chemistry*. 2021;68:317–346.
- Dedvisitsakul P, Watla–Iad K. Antioxidant activity and antidiabetic activities of Northern Thai indigenous edible plant extracts and their phytochemical constituents. *Heliyon*. 2022;8:e10740.
- Fisher L Ianiro, T Lau, F Wang, et al. Synergistic effects of phenolic mixtures in human cell models of aging. Fed.Am Soc Exp Biol. 2015;29:608–636.
- Shad AA, Ahmad S, Ullah R, et al. Phytochemical and biological activities of four wild medicinal plants. Sci World J. 2014;857363.
- Benwahhoud M, Jouad H, Eddouks M, et al. Hypoglycemic effect of Suaeda fruticosa in streptozotocin-induced diabetic rats. Journal of Eth-nopharmacology. 2001;76:35–38.
- Ravikumar S, Gnanadesigan M, Inbaneson JS, et al. Hepatoprotective and antioxidant properties of *Suaeda maritima* (L.) Dumort ethanolic extract on concanavalin–A induced hepatotoxicity in rats. *Indian Jour*nal of Experimental Biology. 2011;49:455–460.
- Karker M, Falleh H, Msaada K, et al. Antioxidant, Anti-inflammatory and anticancer activities of the medicinal halophyte Reaumuria vermiculata. XCLI Journal. 2016;15:297–307.
- Zhou K, You L. Effects of extraction solvent on wheat bran antioxidant activity estimation. LWT Food Science and Technology. 2004;37:717– 721.
- Oueslati S, Trabelsi N, Boulaaba M, et al. Evaluation of antioxidant activities of the edible and medicinal Suaeda species and related phenolic compounds. *Industrial Crops and Products*. 2012;36:513–518.
- Bennani–Kabchi N, El Bouayadi F, Kehel L, et al. Effect of Suaeda fruticosa extract in the hypercholesterolaemic and insulin–resistant sand rat. Therapie. 1999;54:725–730.
- Bishnoi M, Chopra K, Kulkarni SK. Protective effect of rutin, a polyphenolic flavonoid against haloperidol-induced orofacial dyskinesia and associated behavioural, biochemical and neurochemical changes. Fundamental and Clinical Pharmacology. 2007;21:521–529.
- Elsharabasy FS, Metwally NS, Mahmoud AH, et al. Phytoconstituents and Hepatoprotective Effect of Suaeda Monoica Forssk and Suaeda Pruinosa Lange. *Biomed Pharmacol J.* 2019;12(1).

- Falleh H, Ksouri R, Lucchessi ME, et al. Ultrasound–Assisted Extraction: Effect of Extraction Time and Solvent Power on the Levels of Polyphenols and Antioxidant Activity of Mesembryanthemum edule L. Aizoaceae Shoots. Tropical Journal of Pharmaceutical Research. 2012;11(2):243–249.
- Hayouni EA, Abderabba M, Bouix M, et al. The effects of solvents and extraction method on the phenolic contents and biological activities in vitro of Tunisian *Quercus coccifera* L. and *Juniperus phoenicea* L. fruit extracts. *Food Chemistry*. 2007;10:1126–1134.
- Hogan S, Zhang L, Li J, et al. Antioxidant properties and bioactive components of Norton (Vitis aestivalis) and Cabernet Franc (Vitis vinifera) wine grapes. LWT–Food Science and Technology. 2009;42:1269–1274.
- Ou B, Hampsch–Woodill M, Prior RL. Development and validation of an improved oxygen radical absorbance capacity assay using fluorescein as the fluorescent probe. *Journal of Agricultural and Food Chemistry*. 2001;49:4619–4626.
- Rao YK, Geethangili M, Fang SH, et al. Antioxidant and cytotoxic activities of naturally occurring phenolic and related compounds: A comparative study. Food and Chemical Toxicology. 2007;45:1770–1776.
- Saidana D, Mahjoub MA, Boussaada O, et al. Chemical composition and antimicrobial activity of volatile compounds of *Tamarix boveana* (*Tamaricaceae*). Microbiological research. 2008;163: 445–455.
- Senevirathne M, Kim S, Siriwardhana N, et al. Antioxidant potential of Ecklonia cava on reactive oxygen species scavenging metal chelating, reducing power and lipid peroxidation inhibition. *LWT Food Science* and Technology. 2006;12:27–38.
- Sousa C, Taveira M, Valentão P, et al. Inflorescences of Brassicacea species as source of bioactive compounds: a comparative study. Food Chemistry. 2008;110: 953–961.
- Weisburg JH, Weissman DB, Sedaghat T, et al. In vitro Cytotoxicity of Epigallocatechin Gallate and Tea Extracts to Cancerous and Normal Cells from the Human Oral Cavity. *Basic and Clinical Pharmacology and Toxicology*. 2004;95:191–200.
- Wong PYY, Kitts DD. Chemistry of buttermilk solid antioxidant activity. *Journal of Dairy Science*. 2003;86:1541–1547.
- Xavier CP, Lima CF, Fernandes–Ferreira M, et al. Salvia fruticosa, Salvia officinalis, and rosmarinic acid induce apoptosis and inhibit proliferation of human colorectal cell lines: the role in MAPK/ERK pathway. Nutrition and Cancer Journal. 2009;61(4):564–571.