

Pseudomonas stutzeri-an opportunistic pathogen

Abstract

Pseudomonas stutzeri is a Gram-negative bacterium that has been accounted for as a causative specialist of certain diseases. It has been accounted as a cause of pneumonia, meningitis, visual disease, osteomyelitis and joint diseases. Thus, this species could be considered an opportunistic but rare pathogen. In addition, possess an extensive variety of resistance mechanisms against diverse group of antibiotics

Keywords: 16S rRNA, DNA-rRNA hybridization, organotrophically, genomic, anaerobic

Volume 9 Issue 3 - 2022

Nida Tabassum Khan

Department of Biotechnology, Balochistan University of Information Technology, Engineering and Management Sciences, Pakistan

Correspondence: Nida tabassum khan, Department of Biotechnology, Faculty of Life Sciences & Informatics, Balochistan University of Information Technology, Engineering and Management Sciences, Takatu Campus, Airport Road, Quetta, Balochistan, Pakistan, Tel03368164903, Email nidatabassumkhan@yahoo.com

Received: May 23, 2022 | Published: June 21, 2022

Introduction

Pseudomonas stutzeri is a nonfluorescent denitrifying bacterium broadly dispersed in the environment, and has been isolated as an opportunistic pathogen from humans.¹ It was first described by Burri and Stutzer in 1895.² Later, in 1952 characterization of its phenotypic traits and taxonomic position as *Pseudomonas stutzeri* by Lehmann and Neumann.³ The sequence similarity of the rRNAs, exhibited at first by DNA-rRNA hybridization, show the authenticity of the incorporation of *Pseudomonas stutzeri* in the family *Pseudomonas*.⁴ *Pseudomonas stutzeri* are identified as denitrifiers in the environment, exhibiting nutritional versatility by utilizing diverse carbon source such starch, maltose, and ethylene glycol.^{5,6} Varieties in DNA successions led into various genomic variations with in the species (genomovars) that are phylogenetically related.⁷ A few strains stand out due to explicit metabolic properties such as denitrification, aromatic complexes degradation, nitrogen fixation, biotransformation etc.⁸ *Pseudomonas stutzeri* is presently perceived as having a place with the class Gamma proteobacteria and phylogenetic investigations of its 16S rRNA successions and other phylogenetic markers show that they have a place with similar branch, along with related with in the genus *Pseudomonas*.^{9,10} *Pseudomonas stutzeri* are rod shaped bacteria approximately 1 to 3 μm long and 0.5 μm in width, and have a solitary polar flagellum.¹¹ Phenotypic characteristics of the class incorporate a negative Gram stain, positive catalase and oxidase tests, and a stringently respiratory metabolism.¹² They can develop on starch and maltose and have a negative response in arginine dihydrolase and glycogen hydrolysis tests.¹³ GC content of their genomic DNA lies somewhere in the range of 60 and 66 mol% and DNA hybridizations empower somewhere around 17 genomic gatherings, called genomovars, to be recognized.¹⁴ Individuals from the equivalent genomovar have over 70% comparability in DNA hybridizations. Individuals from various genomovars for the most part have comparability values beneath half.^{15,16} The astounding physiological and biochemical variety and adaptability of *Pseudomonas stutzeri* is shown by its ability to develop organotrophically through mineralizing or debasing a large number of natural substrates.¹⁷ its capacity to develop anaerobically, involving different terminal electron acceptors in a rigorously oxidative digestion involving inorganic substrates for acquiring extra energy.¹⁸ In addition, *Pseudomonas stutzeri* displayed resistance towards heavy metals and takes part in nutritional cycles of including C, N, S, and P.¹⁹ *Pseudomonas stutzeri* is well adaptable to grow under different temperatures thus it is a significant physiological

trademark when the natural surroundings that can be colonized by this species are thought of.²⁰ It gives new hereditary varieties to colonizing new environments or for possessing new natural specialties, in any event, when the populace is basically clonal.²¹

For a about 15-year time span after 1956, a few reports depicted the isolation of *Pseudomonas stutzeri* from clinical and neurotic samples.²² In any case, there was no unmistakable relationship of this species with pathogenesis cycle.²³ However later, a couple of instances of *Pseudomonas stutzeri* associated diseases have been accounted for in relationship with bacteremia/septicemia, bone/joint infection, osteomyelitis, joint inflammation, endocarditis, endophthalmitis, meningitis, pneumonia or potentially empyema, ecthyma gangrenosum etc.^{24,25} Thus, *Pseudomonas stutzeri* is ubiquitous in hospital environments and this species could be considered an opportunistic but rare pathogen.²⁶ In addition, it was found to be sensitive towards antibiotics because of its low occurrence in clinical environments and, consequently, its lower exposure to antibiotics.²⁷ Regardless of these outcomes, when bacterial secludes were obtained from immunosuppressed patients, no massive contrasts in anti-infection vulnerability between *Pseudomonas aeruginosa* and other *Pseudomonas* spp., including *Pseudomonas stutzeri*, were distinguished.²⁸ Strangely, except for fluoroquinolones, resistance *Pseudomonas stutzeri* strains have been confined for practically all antibiotic families.²⁹ This recommends that this species possess an extensive variety of antibiotic resistance mechanisms.³⁰ Such as changes in external layer proteins and lipopolysaccharide profiles³¹ and the presence of β -lactamases that hydrolyze regular and semisynthetic penicillins, wide range " β -lactamase-stable" cephalosporins, and monobactams with comparative rates.³²

Conclusion

Thus, *Pseudomonas stutzeri* is a denitrifying bacterium broadly dispersed in the environment, and has been isolated as an opportunistic pathogen from humans. In addition, also displays resistance against diverse antibiotics

Acknowledgments

None.

Conflicts of interest

The authors state that there is no conflict of interest.

Funding

None.

References

- Lalucat J, Bennasar A, Bosch R, et al. Biology of *Pseudomonas stutzeri*. *Microbiology and molecular biology reviews*. 2006;70(2):510–547.
- Bennasar A, Rossello-Mora R, Lalucat J, et al. 16S rRNA gene sequence analysis relative to genomovars of *Pseudomonas stutzeri* and proposal of *Pseudomonas balearica* sp. nov. *International Journal of Systematic and Evolutionary Microbiology*. 1996;46(1):200–205.
- Van Niel CB, Allen MB. A note on *Pseudomonas stutzeri*. *Journal of Bacteriology*. 1952;64(3):413–422.
- Bennasar A, Guasp C, Lalucat J. Molecular methods for the detection and identification of *Pseudomonas stutzeri* in pure culture and environmental samples. *Microbial ecology*. 1998;35(1): 22–33.
- Carlson CA, Ingraham JL. Comparison of denitrification by *Pseudomonas stutzeri*, *Pseudomonas aeruginosa*, and *Paracoccus denitrificans*. *Applied and Environmental Microbiology*. 1983;45(4):1247–1253.
- Zhang J, Wu P, Hao B, et al. Heterotrophic nitrification and aerobic denitrification by the bacterium *Pseudomonas stutzeri* YZN–001. *Bioresource technology*. 2011;102(21):9866–9869.
- Ginard M, Lalucat J, Tümmler B, et al. Genome organization of *Pseudomonas stutzeri* and resulting taxonomic and evolutionary considerations. *International Journal of Systematic and Evolutionary Microbiology*. 1997;47(1):132–143.
- Cladera AM, Bennasar A, Barceló M, et al. Comparative genetic diversity of *Pseudomonas stutzeri* genomovars, clonal structure, and phylogeny of the species. *Journal of bacteriology*. 2004;186(16):5239–5248.
- Sikorski J, Graupner S, Lorenz MG, et al. Natural genetic transformation of *Pseudomonas stutzeri* in a non-sterile soil. *Microbiology*. 1998;144(2):569–576.
- Rius N, Fusté MC, Guasp C, et al. Clonal population structure of *Pseudomonas stutzeri*, a species with exceptional genetic diversity. *Journal of Bacteriology*. 2001;183(2):736–744.
- Rosselló R, García-Valdes E, Lalucat J, et al. Genotypic and phenotypic diversity of *Pseudomonas stutzeri*. *Systematic and applied microbiology*. 1991;14(2):150–157.
- Lapage SP, Hill L, Reeve JD. *Pseudomonas stutzeri* in pathological material. *Journal of Medical Microbiology*. 1968;1(2):195–202.
- Holmes B. Identification and distribution of *Pseudomonas stutzeri* in clinical material. *Journal of applied bacteriology*. 1986;60(5):401–411.
- Cladera AM, García-Valdés E, Lalucat J. Genotype versus phenotype in the circumscription of bacterial species: the case of *Pseudomonas stutzeri* and *Pseudomonas chloritidismutans*. *Archives of microbiology*. 2006;184(6):353–361.
- Palleroni NJ, Doudoroff M, Stanier RY, et al. Taxonomy of the aerobic pseudomonads: the properties of the *Pseudomonas stutzeri* group. *Microbiology*. 1970;60(2):215–231.
- Mulet M, Gomila M, Gruffaz C, et al. Phylogenetic analysis and siderotyping as useful tools in the taxonomy of *Pseudomonas stutzeri*: description of a novel genomovar. *International journal of systematic and evolutionary microbiology*. 2008;58(10):2309–2315.
- Cladera AM, del C Sepúlveda-Torres L, Valens-Vadell M, et al. A detailed phenotypic and genotypic description of *Pseudomonas* strain OX1. *Systematic and applied microbiology*. 2006;29(5):422–430.
- Ferguson SJ. Denitrification and its control. *Antonie van Leeuwenhoek*. 1994;66(1):89–110.
- Yang X, Wang S, Zhou L. Effect of carbon source, C/N ratio, nitrate and dissolved oxygen concentration on nitrite and ammonium production from denitrification process by *Pseudomonas stutzeri* D6. *Bioresource Technology*. 2012;104:65–72.
- Carlson CA, Pierson LS, Rosen JJ, et al. *Pseudomonas stutzeri* and related species undergo natural transformation. *Journal of Bacteriology*. 1983;153(1):93–99.
- Rumbaugh KP. Genomic complexity and plasticity ensure *Pseudomonas* success. *FEMS Microbiology Letters*. 2014;356(2):141–143.
- Noble RC, Overman SB. *Pseudomonas stutzeri* infection a review of hospital isolates and a review of the literature. *Diagnostic microbiology and infectious disease*. 1994;19(1):51–56.
- Reisler RB, Blumberg H. Community-acquired *Pseudomonas stutzeri* vertebral osteomyelitis in a previously healthy patient: case report and review. *Clinical infectious diseases*. 1999;29(3):667–669.
- Bisharat N, Gorlachev T, Keness Y. 10-years hospital experience in *Pseudomonas stutzeri* and literature review. *The Open Infectious Diseases Journal*. 2012;6(1).
- Kose Mehmet, Ozturk M, Kuyucu T, et al. Community-acquired pneumonia and empyema caused by *Pseudomonas stutzeri*: a case report. *Turkish Journal of Pediatrics*. 2004;46(2):177–178.
- Holmes B. Identification and distribution of *Pseudomonas stutzeri* in clinical material. *Journal of applied bacteriology*. 1986;60(5):401–411.
- Gilardi GL. Infrequently encountered *Pseudomonas* species causing infection in humans. *Annals of Internal Medicine*. 1972;77(2):211–215.
- Tattawasart U, Maillard JY, Furr JR, et al. Development of resistance to chlorhexidine diacetate and cetylpyridinium chloride in *Pseudomonas stutzeri* and changes in antibiotic susceptibility. *Journal of Hospital Infection*. 1999;42(3):219–229.
- Russell AD, Mills AP. Comparative sensitivity and resistance of some strains of *Pseudomonas aeruginosa* and *Pseudomonas stutzeri* to antibacterial agents. *Journal of Clinical Pathology*. 1974;27(6):463–466.
- Ergin C, Mutlu G. Clinical distribution and antibiotic resistance of *Pseudomonas* species. *Eastern Journal of Medicine*. 1999;4(2):72–77.
- Tattawasart U, Maillard JY, Furr JR, et al. Outer membrane changes in *Pseudomonas stutzeri* resistant to chlorhexidine diacetate and cetylpyridinium chloride. *International journal of antimicrobial agents*. 2000;16(3):233–238.
- Russell AD. Mechanisms of bacterial resistance to biocides. *International Biodeterioration & Biodegradation*. 1995;36(3–4):247–265.