**In vivo propagation responses of sugarcane (sacharum officinarum l) genotypes at metahara sugar estate of Ethiopia**

**Abstract**

In vivo propagation of micropropagated sugarcane plants of three selected sugarcane genotypes was carried out with the objective of evaluating their proliferation responses to BAP. Accordingly, six levels of BAP (0, 0.5, 1, 1.5, 2 and 2.5mgL⁻¹) with three sugarcane genotypes in a completely randomized design with 6×3 factorial treatment combination arrangements were tested. Data on the number of tillers per shoot, average shoot length (cm) and number of leaves per shoot were collected after 60 days of planting on white polyethylene bag having 10 cm height and 8 cm diameter filled with soil, compost and sand at the ratios of 8:3:2 respectively. Analysis of variance revealed that the interaction effects of BAP and the sugarcane genotypes was very highly significant (P<0.0001) on the response variables tested. In sugarcane genotype C132-81, the optimum number of tillers per shoot (4.00) and average shoot length (30.67cm) were recorded on 1mgL⁻¹ BAP while the maximum number of leaves (6.00) was obtained at 0.5 and 1mgL⁻¹. In sugarcane genotype C86-156, the optimum number of tillers per shoot (3.53) and number of leaves per shoot (7.00) were obtained at 1.5mgL⁻¹ BAP while the optimum number of tillers per shoot (4.67) was recorded at 1mgL⁻¹ BAP. Therefore, from this result it can be concluded that, using this result, on average a plantlet can produce 4 tillers per shoot within a month and can reduce plantlets procurement cost by 25%.

**Keywords:** in vivo propagation, sugarcane genotypes, C132-81, C86-156, SP70-1284

**Abbreviations:** BAP, 6-benzylaminopurine; REGWQ, ryan-einot-gabriel-welsch; ANOVA, analysis of variance; IVPT, in vivo propagation technology; CV, coefficient of variation; SE, standard error

**Introduction**

Sugarcane is a perennial grass which produces seed under suitable conditions, but for commercial production, it is propagated from stalk cuttings. Propagation through stalk cuttings is the traditional method of sugarcane propagation in which stalk cuttings containing one or more buds, termed setts are used for commercial planting. The Ethiopian Sugar Industry used this traditional method of propagation alone for the last 50 years till 2012. However, under the new plan of the Industry to expand the existing farms and establishment of vast sugarcane plantation projects with many high crushing capacity sugar factories, the tradition propagation method alone was seen to have various draw backs. Among these, availability of enough amount of quality disease free planting material within short time, transport of bulky unclean seed cane from existing farms to the remote project farms, the low rate of propagation(usually 1:10), lack of methods for fast commercialization of improved and adapted varieties, obsolescence of productive commercial varieties due to disease, lack of alternative techniques for rejuvenation and disease cleansing of the old contaminated sugarcane varieties were the major limitations identified as challenges to the vast expansion and new development plans.

With a view to minimize the challenges, micropropagation technology was adopted and implemented to supplement the tradition method of sugarcane propagation in all sugar estates and projects. Micropropagation Technology is a technique through which group of genetically identical plants all derived from a selected individual multiply vegetatively and rapidly by aseptic culture of meristematic conditions in vitro. Nowadays, unlike the traditional propagation method, it is the only practical means of achieving rapid and large scale production of disease free quality planting materials in sugarcane and alternative approach for fast multiplication of a genotype in its original form. It is very effective in entire disease cleansing, rejuvenation and subsequent mass propagation of well adapted and promising varieties facing gradual deterioration in yield, quality and vigor due to accumulation of pathogens during prolonged vegetative cultivation and hence sustains the productive potential of sugarcane crops for a longer period. Furthermore, micropropagated sugarcane plants were reported to give superior under similar agronomic management systems.

However, the Ethiopian Sugar Corporation is procuring the micropropagated sugarcane plantlets from different organizations where the cost of procurement is high (about US$ 0.205 per plantlet). The erratic supply, long distance transport of the delicate
Materials and methods

Area of study

The study was carried out at Metahara sugarcane plantation, located at Eastern part of the country, at about 200kms away from Addis Ababa, Ethiopia. Metahara Sugar estate is situated at 8°53’ N latitude and 39°52’E longitude at an altitude of 950 m.a.s. with a semi-arid climatic condition.

Experimental materials

The experimental materials were in vitro propagated sugarcane genotypes of C132-81, C86-56 and SP70-1284. The primary acclimatized plantlets of these sugarcane varieties were delivered with intact coco-peat from Mekelle Technology Institute Tissue Culture Laboratory and directly planted in white polyethylene bag (8 cm diameter with 10 cm height) filled with mixture of Luvisol, sand and compost in the ratio of 8:2:1.

Experimental design

The experimental design was Completely Randomized Design with factorial treatment combination arrangements. Three sugarcane genotypes (C132-81, C86-56 & SP70-1284) with six levels of 6-benzylaminopurine (BAP) (0, 0.5, 1, 1.5, 2 and 2.5 mg/l) resulting in 3 x 6 = 18 treatment combinations each replicated three times. Each plot contains 30 pots (one plantlet a pot).

Data collections

Data such as number of tillers per shoot, shoot length and number of active leaves per shoot were collected from ten randomly selected plantlets at 30th and 60th days of treatment application.

Data analysis

Then the average data were subjected to analysis of variance using SAS software version 9.2 while separation of significant means’ was done using REGWQ (Ryan-Einot-Gabriel-Welsch) Multiple Range Test.

Result and discussion

Analysis of variance (ANOVA) proved that the interaction of 6-benzylaminopurine (BAP) and the sugarcane genotypes has a highly significant (p<0.001) effect on the number of tillers per shoot, average shoot length (cm) and number of active leaves per shoot of the sugarcane genotypes tested: C132-81, C86-156 and SP70-1284 (Table 1). Similarly, comparison of the tested sugarcane genotypes showed significant variation both in the number of tillers per shoot and number of leaves per shoot. However, there was no statistically significant difference between the three sugarcane genotypes in average shoot length. In all the sugarcane genotypes, the lowest number of tillers per shoot, average shoot length and number of leaves per shoot were obtained on the control treatment (0 mg/l BAP). In sugarcane genotype C132-81, the lowest number of tillers per shoot (1.97), average shoot length (27.67 cm) and number of leaves per shoot (4.5) was obtained at 0 mg/l BAP (control). But, increase in the concentration of BAP from 0 up to 1mg/l BAP showed an increasing trend for all the response variables tested: number of tillers per shoot, average shoots length and number of leaves per shoot (Table 2). The maximum number of tillers per shoot (4.00) and average shoot length (30.67 cm) were recorded on 1mg/l BAP while the maximum number of leaves (6.00) was obtained at 0.5 and 1mg/l for the sugarcane genotype C132-81. However, further increase in the concentrations of BAP beyond 1mg/l showed no positive effect for all the response variables tested in C132-81 (Table 2).

Table 1 Effects of genotype on the number of tillers and leaves per shoot and shoot length

<table>
<thead>
<tr>
<th>Genotypes</th>
<th>Number of tillers/shoot</th>
<th>Average shoot length(cm)</th>
<th>Number of leaves/shoot(cm)</th>
</tr>
</thead>
<tbody>
<tr>
<td>SP70-1284</td>
<td>3.86 ± 0.21</td>
<td>29.00 ± 0.68</td>
<td>6.00 ± 0.09</td>
</tr>
<tr>
<td>C132-81</td>
<td>2.94 ± 0.21</td>
<td>28.83 ± 0.68</td>
<td>5.56 ± 0.09</td>
</tr>
<tr>
<td>C86-156</td>
<td>2.57 ± 0.21</td>
<td>29.50 ± 0.68</td>
<td>6.28 ± 0.09</td>
</tr>
</tbody>
</table>

In vivo propagation responses of sugarcane (Saccharum officinarum L) genotypes at Metahara sugar estate of Ethiopia

1.97 1.73 6.5 29.00 6.00 2.33

1.67 0.52 4.00 2.33 6.00 8.7

1.67 0.52 31.33 2002.

SP70-1284

5.67 0.22 29.67 3.00 3.00 30.67

IVPT 6.00 32.67 3.33 5.00 5.33 6.00

28.00 -1 6.33 3.53 5.33 0.22 5.67

Sacharum offic


Table 2 The Effect of 6-benzylaminopurine on the number of tillers per shoot, number of leaves per shoot and average shoot length of sugarcane genotypes

<table>
<thead>
<tr>
<th>BAP (mg/l)</th>
<th>Number of tillers/ shoot</th>
<th>Average shoot length (cm)</th>
<th>Number of leaves/ shoot</th>
<th>Average shoot length (cm)</th>
<th>Number of tillers/ shoot</th>
<th>Average shoot length (cm)</th>
<th>Number of leaves/ shoot</th>
</tr>
</thead>
<tbody>
<tr>
<td>0</td>
<td>1.97</td>
<td>27.67</td>
<td>4.50</td>
<td>1.73</td>
<td>28.00</td>
<td>6.00</td>
<td>5.33</td>
</tr>
<tr>
<td>0.5</td>
<td>3.33</td>
<td>27.33</td>
<td>6.00</td>
<td>3.00</td>
<td>29.67</td>
<td>6.33</td>
<td>4.33</td>
</tr>
<tr>
<td>1</td>
<td>4.00</td>
<td>30.67</td>
<td>6.00</td>
<td>3.00</td>
<td>31.33</td>
<td>6.33</td>
<td>4.33</td>
</tr>
<tr>
<td>1.5</td>
<td>3.00</td>
<td>29.33</td>
<td>5.33</td>
<td>3.53</td>
<td>29.33</td>
<td>7.00</td>
<td>4.67</td>
</tr>
<tr>
<td>2</td>
<td>3.00</td>
<td>28.00</td>
<td>5.00</td>
<td>2.00</td>
<td>29.33</td>
<td>6.00</td>
<td>5.00</td>
</tr>
<tr>
<td>2.5</td>
<td>2.33</td>
<td>29.00</td>
<td>6.53</td>
<td>2.33</td>
<td>29.33</td>
<td>6.00</td>
<td>3.33</td>
</tr>
<tr>
<td>CV</td>
<td>8.7</td>
<td>9.9</td>
<td>6.5</td>
<td>8.7</td>
<td>9.9</td>
<td>6.5</td>
<td>6.5</td>
</tr>
<tr>
<td>SE</td>
<td>0.52</td>
<td>1.67</td>
<td>0.22</td>
<td>0.52</td>
<td>1.67</td>
<td>0.22</td>
<td>0.52</td>
</tr>
</tbody>
</table>

Remark: Means with the same letter are not significantly different at 0.05 probability level.

In sugarcane genotype C86-156, the maximum number of tillers per shoot (3.53) and number of leaves per shoot (7.00) was obtained at 1.5 mg/l BAP while the maximum average shoot length was observed at 1 mg/l (Table 2). In sugarcane genotype SP70-1284, the maximum average shoot length (32.67 cm) and highest number of leaves per shoot (6.53) was obtained at 1 mg/l BAP while the maximum number of tillers per shoot was observed at 1 mg/l BAP (Table 2). Increase in concentrations of BAP beyond the optimum levels showed no any significant increment in the response variables tested (Table 2). The lowest number of tillers per shoot, average shoot length and number of leaves per shoot observed on the treatment lacking the plant growth regulator BAP (control) could be due to the role of BAP in enhancing cell division that will in turn promote plant growth and development (tillering, shoot elongation and leaf development). Thus, lack of BAP in the control treatment produced the lowest response variables among the other treatments containing BAP. Increase in the concentration of BAP increased the response variables to the optimum levels and showed a declining effect. This could be due to the fact that increased levels of BAP beyond the optimum level hampers cell division and deleterious to plant cell. The result obtained in this study is different from the study results reported by Tana-Beles Sugar Development Project on Sugarcane genotypes B52-298 and NCo-334 treated with trimming 1/3 of the leaves, DAP fertilizer at 0.16g M⁻³ and plant growth regulators GA₃, BAP and kinetin each at 0.04mg L⁻³. On these treatment combinations, B52-298 gave 6.45±0.51 tillers per shoot with 4.39±1.44cm average shoot length and 5.12±0.23 leaves per shoot while NCo-334 produced 5.77±0.79 tillers per shoot with 7.21±0.11cm average shoot length and 5.51±0.05 leaves per shoot. Similarly, N14 gave 5.36±0.55 tillers per shoot with 5.71±0.15cm average shoot length and 5.41±0.30 leaves per shoot on the same treatment combination.

Conclusion

The traditional method of sugarcane propagation has many limitations while micropropagation technology is potent to solve all the limitations except it is costly to produce or procure the plantlets. Hence, in vivo propagation technology (IVPT) was developed to complement tissue culture technology, but reproducibility of the IVPS protocols is dependent on the genotype, environmental weather conditions, soil mixture, plant growth regulators and interaction of factors used. Treatment combination containing 1mg/l BAP and 1.5mg/l gave optimum propagation responses. Sugarcane genotype C132-81 4 tillers per shoot with 30.67cm average shoot length and 6 active leaves per shoot while C86-56 produced 3.53 & 4.67 tillers per shoot, 29.33cm average shoot length and 7.0 & 6.33 active leaves per shoot per month, respectively. Thus, the current finding will help minimize the current challenges of sugarcane production by rapidly availing adequate amount of quality planting material of sugarcane while reducing the cost of plantlets procurement and hence the cost of sugar production.

Acknowledgements

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Conflict of interest

The author declares no conflict of interest.

References


