

Bio isolation, chemical purification, identification, antimicrobial and synergistic efficacy of extracted essential oils from stem bark extract of *Spondias mombin*(Linn)

Abstract

The purpose of this research work is to isolate, purify, chemical identification, antimicrobial and Synergistic efficacy of extracted essential oils from ethyl acetate extract of *Spondias mombin*. The ethyl acetate stem bark extract of *Spondias mombin* were air-dried, chopped into smaller pieces and cold extracted exhaustively with ethylacetate. The crude extract was partitioned using various solvents and the dichloromethane fraction was concentrated and fractionated using column chromatography packed with Si-gel and Sephadex-LH and eluted with appropriate solvent systems accordingly. In order to obtain pure extracts, partially purified fractions were further purified. The structures of the isolated compounds were determined by using data obtained from GC-MS spectrum. The compound isolated include Aspidofractinine-3-methanol, Phthalic acid, 2-ethylhexyl tetradecyl ester, Phthalic acid, di (2-propylpentyl) ester, (9-(2', 2'-Dimethylpropanoilhydrazono)-3,6-dichloro-2,7-bis-[2-(diethylamino)-ethoxy]fluorine) and Tere phthalic acid, dodecyl 2-ethylhexyl ester. These compounds have individual and synergistic activity against Gram negative (*E.coli*), Gram-positive (*Bacillus subtilis*) and (*Aspergillusflavus*) at 10, 5, 2.5 and 1.25 μ g/mL.

Keywords: *spondias mombin*, antimicrobial, essential oils, gc-ms

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Introduction

The search for drugs from natural sources has been on the increase in ethno-medicinal research¹ and plant kingdom has served as an inexhaustible source of useful drugs, foods, additives, flavouring agents, lubricants, colouring agents and gums from time immemorial.² Medicinal plants have been used since time immemorial in virtually all societies as source of medicine to combat various ailments including infectious diseases. The therapeutic power of herbs has been recognized since creation of the universe and botanic medicine is one of the oldest practiced professions by mankind.³ Medicinal plants contain large varieties of chemical substances with important therapeutic properties that can be utilised in the treatment of human diseases.^{4,5} The antibacterial, antimicrobial, antiviral and antifungal potentials of the plant have been reported.⁶⁻⁹ The plant was recommended for use by pregnant women after five months of pregnancy¹⁰ while the mineral analysis of the plant has also been documented.^{11,12} The abortifacient activity of an aqueous extract of *Spondias mombin* was reported by Offiah et al.¹³ The plant has been shown to have a wide range of phyto constituents such as tannins, saponins and anthraquinone glycosides.⁷ The plant has been reported to be used in the treatment of many disease conditions in the Eastern part of Nigeria by the natives. The associated link between the composition of this world-wide cultivated plant and the reported medicinal and economic values prompted this study aimed at evaluating the phytochemical and antioxidant properties as well as the microbial inhibitory effects of the leaf and stem bark extracts of the plant.

Materials and methods

Collection of plant materials

The stem bark of *Spondias mombin* was collected from a local farm at Owo (710°59.998N and 534°59.988E), Ondo State, Nigeria at WAT

UTC+1 time zone on 26th and 27th of February, 2016. Fresh and healthy plants were also collected during its fruiting season, between April and July 2017 from the same geographical locations.

Authentication of plant samples

The plants were authenticated at the Department of Plant Science and Biotechnology, Adekunle Ajasin University, Akungba- Akoko, Ondo state, Nigeria.

Extraction and isolation from *Spondias mombin* plant

The stem-bark of *Spondias mombin* plant were harvested and air-dried. The dried bark was milled into powdered form using manual grinder. Powdered plant material (1kg) each of the different plant parts was extracted with 3L of 70% (v/v) ethanol, ethyl acetate and distilled water for 72 hours at room temperature. The extraction process was repeated four times until the extract became clear. The filtrates were combined and concentrated *in vacuo* using rotatory evaporator at 35°C. The fraction was labeled SMSBEA and kept in tightly stoppered bottles in a refrigerator at -4°C for further analysis.

Isolation and purification of extracted essential oil (eo) from stem bark of *spondias mombin* (linn)

The crude extract (20g) of *S. mombin* was adsorbed onto silica and successively eluted with solvents and the dichloromethane fraction was obtained, concentrated and dried. From the dichloromethane fraction, 5.8g was adsorbed onto silica gel (200–400mesh). The column was degassed with n-hexane and thin layer chromatography guided gradient elution was carried out using n-hexane, n-hexane-dichloromethane (60:30) and n-hexane-dichloromethane (65:35) respectively. The two later elutions were bulked and labelled Fraction 2B weighing 3.8g. Fraction 2B was subjected to column chromatography, degassed and was eluted with n-hexane-dichloromethane in the following

respective proportions, 65:35, 60:40, 55:45, 50:50, 45:55, 40:60 and 35:65 respectively. The elution from 50:50 and 40:60 were bulked and concentrated in-vacuo. The elution from 30:70 was also concentrated as well. The concentrates were placed on Sephadex LH-20 and eluted with dichloromethane-methanol (70:30) giving five pure extracts labelled A₃, B₃, C₃, E₃, and F₃. The compounds were then analysed using FT-IR Spectrophotometer (Perkin Elmer) over the frequency range from 4000-400 cm⁻¹. UV-VIS and FT-IR analysis was performed at National Institute of Pharmaceutical Education and Research (NIPER), Mohali (India). The GC-MS analysis of the compounds was performed using a Perkin Elmer GC Claurus 500 system and gas chromatograph interfaced to a Mass Spectrometer (GC-MS) equipped with Elite-1 fused silica capillary column (30 m×1µl was Mdf. Composed of 100% Dimethyl poly siloxane)

Antibacterial and synergistic assay of extracted essential oil (eo) from stem bark of *Spondias mombin* (Linn)

Antibacterial and antifungal screening of the extract: All the test bacteria, were sub-cultured onto sterile Mueller Hinton agar plates, and incubated at 37°C for 18-24 hours. Five distinct colonies for each organism were inoculated onto sterile Mueller Hinton broth and incubated for 3- 4hours. All inocula were standardized accordingly to match the 0.5 McFarland standards, and this standard was used for all susceptibility tests. All the extracts were reconstituted accordingly into the following concentrations; 10, 5, 2.5, 1.25mg/ml, The susceptibility testing was investigated by the agar well diffusion method. A 0.1ml of 1: 10,000 dilution (equivalent to 10⁶cfu/ml) of fresh overnight culture of the clinical isolates grown in Mueller Hinton agar and Sabouraud dextrose agar was seeded into 40ml of Mueller Hinton agar, and properly mixed in universal bottles. The mixture was aseptically poured into sterile Petri dishes and allowed to set. Using a sterile Cork borer of 6mm diameter, equidistant wells were made in the agar. Drops of the re suspended, (2ml per well) extracts with

concentrations between 10, 5, 2.5, 1.25mg/mL, were introduced into the wells till it was filled. The plates could stand on the bench for an hour, to allow pre-diffusion of the extracts before incubation at 37°C for 24 hours for the bacterial isolates and 24°C for 48 hours for the fungal isolates. The zones of inhibition were measured to the nearest millimeter (mm) using a standard transparent meter rule. All experiments were performed in duplicates.¹⁴⁻¹⁸

Gas chromatography and mass spectroscopy gc/ms method of analysis

GC-MS technique was used in this study to identify the components present in the extract. GC-MS technique was carried out at Indian Institute of Crop Processing Technology (IICPT) Thanjavur, Tamilnadu. GC-MS analysis of this extract was performed using a Perkin Elmer GC Claurus 500 system and gas chromatograph interfaced to a Mass Spectrometer (GC-MS) equipped with Elite-1 fused silica capillary column (30mx1µl was Mdf. Composed of 100% Dimethyl poly siloxane).¹⁹ For GC-MS detection, an electron ionization energy system with ionization energy of 70eV was used. Helium gas (99.999%) was used as the carrier gas at a constant flow rate of 1ml/min. and an injection volume of 2µl was employed (Split ratio of 10:1). Injector temperature was 250°C. The oven temperature was programmed from 110°C (isothermal for 2min.), with an increase of 10°C/min to 200°C, then 5°C /min. to 280°C, ending with a 9min. isothermal at 280°C. Mass spectra were taken at 70eV; a scan interval of 0.5 seconds and fragments from 45 to 450 Da. Total GC running time was 36 min. The relative percentage amount of each component was calculated by comparing its average peak area to the total areas. Software adopted to handle mass spectra and chromatograms was a Turbo mass Ver5.2.0. Compound identification was obtained by comparing the retention times with those of authentic compounds and the spectral data obtained from library data of the corresponding compounds. The given sample was extracted with ethyl acetate and analyzed in GC-MS for different component (Figure 1).

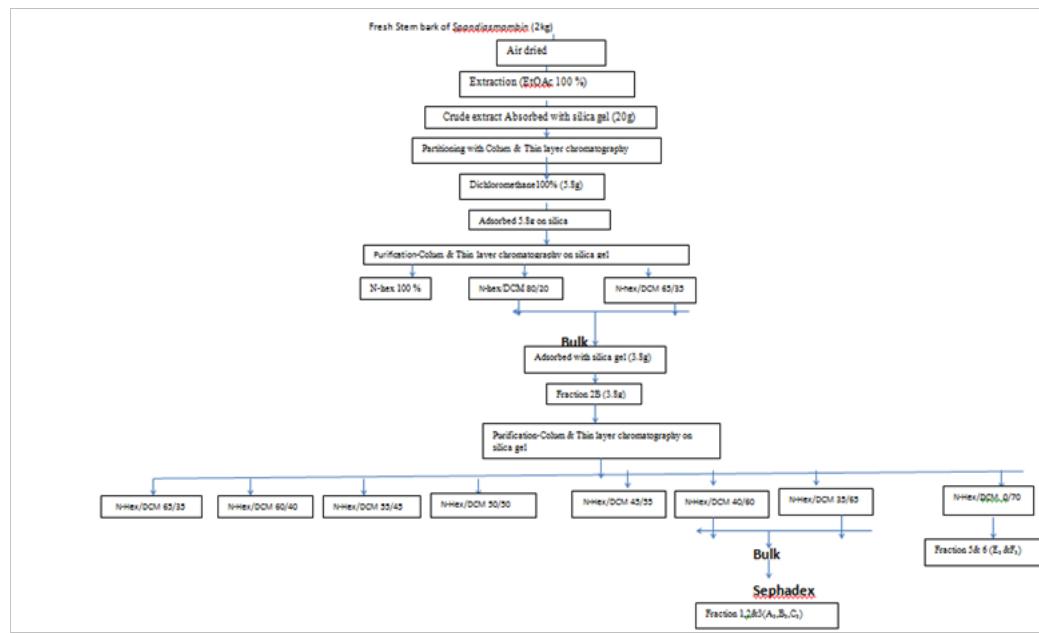


Figure 1 Isolation and purification of extracted essential oil (eo) from stem bark of *Spondias mombin* (Linn).

Key: A₃ - C₂₀H₂₆N₂₀- Aspido fractinine-3-methanol,(2.alpha.,3.beta.,5.alpha.) B₃- C₃₀H₅₀O₄- Phthalic acid, 2-ethylhexyl tetradecyl ester C₃- C₂₄H₃₈O₄- Phthalic acid, di(2-propyl pentyl) ester E₃- C₃₀H₄₂C₁₂N₄O₃- 9-(2',2'-Dimethyl propanoylhydrazone)-3,6-dichloro-2,7-bis-[2-(diethylamino)-ethoxy]fluorene F₃- C₂₈H₄₆O₄- Terephthalic acid, dodecyl 2-ethylhexyl ester

Result and discussion

Structural profiling of pure compound of stem bark extracts of *Spondias mombin* using gas chromatography and mass spectrometric(gc/ms) method of identification

Table 1 Shows the Chromatogram A₃ of Gas chromatography and mass spectrometry (GCMS) of compound A₃, The pure extract contained arrays of compounds, which are listed as follows; Undecane(C₁₁H₂₄), Dodecane(C₁₂H₂₆), 3,9-dimethyl-(C₁₃H₂₈), 5-Ethylundecane(C₁₃H₂₈), 2-Naphthalenol, 1,2-dihydro-, acetate(C₁₂H₁₂O₂), Azulene(C₁₀H₈), Naphthalene(C₁₀H₈), N-Methyl-9-aza-tricyclo[6.2.2.0(2,7)]dodec-2,4,6,11-tetraene-10-one(C₁₂H₁₁NO), 1H-Indene, 1-methylene (C₁₀H₈), Tridecane(C₁₁H₂₄),

Decane,2,9-dimethyl(C₁₂H₂₆), 3,6-Dimethylundecane (C₁₃H₂₈), Octan e,2,3,7-trimethyl- (C₁₁H₂₄), 3-Ethyl-5-methylheptane(C₁₀H₂₂), 2,7,10-Trimethylundecane (C₁₅H₃₂), Nonane, 3-Methyl-5-pro pylon nane (C₁₃H₂₈), Hexadecane, 2,6,10,14-tetramethyl-, Phytan(C₂₀H₄₂), 3-Methyl-5-propylnonane(C₁₃H₂₈), 2,6,10-Trimethyl dodecane, Farnesane (C₁₅H₃₂), Hexadecane, *n*-Cetane(C₁₆H₃₄), 1,2-Benzenedicarboxylic acid, butyl 2-methylpropyl ester, (Phthalic acid, butyl isobutyl ester)(C₁₆H₂₂O₄), 2-Octylbenzoate(1-Methylheptyl benzoate)(C₁₅H₂₂O₂), Benzoic acid, 2-ethylhexyl ester (C₁₅H₂₂O₂), Oxalic acid, allylhexadecyl ester(C₂₁H₃₈O₄), 2-Piperidinone, N-[4-bromo-n-butyl]-1-(4-Bromobutyl)-2-piperidinone)(C₉H₁₆BrNO), 1H-Indene, 1-methylene-(1-Methylene-1H-indene)(C₁₀H₈), 1,2-Benzenedicarboxylic acid, butyl 2-methylpropyl ester (C₁₆H₂₂O₄), Dibutyl phthalate (1,2-Benzenedicarboxylic acid, dibutyl ester) (C₁₆H₂₂O₄), Eicosane, 10-methyl-10-Methyllicosane (C₂₁H₄₄).

Table 1 Chromatogram of aspido fractinine-3 methanol,(2.alpha.,3.beta.,5 alpha, using gas chromatography and mass spectroscopy gc/ms method of analysis (Mass Peaks: 395, 400)(A₃)

Name	Molecular weight	Structural formula	Retindex
ndecane	156	C ₁₁ H ₂₄	1115
Tridecane	184	C ₁₃ H ₂₈	1313
Adakane 12	170	C ₁₂ H ₂₆ CH ₃ (CH ₂) ₁₀ CH ₃	1214
Dimethylundecane	184	C ₁₃ H ₂₈	1185
5-Ethylundecane	184	C ₁₃ H ₂₈	1249
2-Naphthalenol, 1,2-dihydro-, acetate	188	C ₁₂ H ₁₂ O ₂	1476
Aspidofractinine-3-methanol, (2.alpha.,3.beta.,5. alpha.)-	310	C ₂₀ H ₂₆ N ₂ O	2510

Table 2 Shows the Chromatogram B₃ of Gas chromatography and mass spectrometry (GCMS) of compound B₃. The pure extract contained arrays of compounds, which listed as follows; Dodecane (Adakane 12)(C₁₂H₂₆), Undecane(Hendecane)(C₁₁H₂₄), Decane (2,9,Dimethyldecane)(C₁₂H₂₆), Octane, 2,3,7-trimethyl(C₁₁H₂₄), Tridecane, 7-methyl(Methyltridecane)(C₁₄H₃₀), Heptane(3-Ethyl-5-methylheptane)(C₁₀H₂₂), Nonane, (3-Methyl-5-propylnonane) (C₁₃H₂₈), Diisooctylphthalate (Bis(6-methylheptyl) phthalate, 1,2-Benzenedicarboxylic acid, 1,2-Benzenedicarboxylic acid(C₂₄H₃₈O₄), 1,2-Benzenedicarboxylic acid, diheptyl ester(C₂₂H₃₄O₄), Phthalic acid, 5-methylhex-2-yl heptyleste(C₂₂H₃₄O₄), 8-Hexadecenal, 14-methyl-, (Z)-(14-Methyl-8-hexadecenal, (8Z)-14-Methyl-8-hexadecenal (C₁₇H₃₂O), 7-Hexadecenal, (Z)(Z)-7-

Hexadecenal, Z-7-Hexadecenal, (7Z)-7-Hexadecenal(C₁₆H₃₀O), cis-9-Hexadecenal(C₁₆H₃₀O), Heptadecane, 2,6,10,15-tetramethyl-2,6,10,15-Tetramethylheptadecane(C₂₁H₄₄), 9-Octadecenal, (Z)-(Olealdehyde)(C₁₈H₃₄O), 9-Octadecenoic acid, 1,2,3-propanetriyl ester, (E,E,E),(2,3-Bis[(9E)-9-octadecenoyloxy]propyl (9E)-9-octadecenoate)(C₅₇H₁₀₄O₆), n-Propyl 9,12-octadecadienoate, n-Propyl linoleate (C₂₁H₃₈O₂), 9,12-Octadecadienoic acid (Z,Z)-, 2-hydroxy-1-(hydro xymethyl) ethyl ester(Linolein, 2-mono-.beta.-Monolinolein) (C₂₁H₃₈O₄), Butyl 9,12-octadecadienoate (C₂₂H₄₀O₂), Bis(tridecyl) phthalate(1,2-Benzenedicarboxylic acid, ditridecyl ester (C₃₄H₅₈O₄), Ethyl 13-methyl-tetradecanoate(C₁₇H₃₄O₂), Eicosanoic acid, ethyl ester, Ethyl icosanoate, Ethyl arachidate (C₂₂H₄₄O₂).

Table 2 Chromatogram of Phthalic acid, 2-ethylhexyl tetradecylester, using gas chromatography and mass spectroscopy gcms method of analysis (Mass peaks: 395,400) B₃

Name	Molecular weight	Structural formula	Retindex
Dodecane	170	C ₁₂ H ₂₆	1214
Tridecane	184	C ₁₃ H ₂₈	1313
Hendecane	156	C ₁₁ H ₂₄	1115
11Methyldodecane	184	C ₁₃ H ₂₈	1249
Decane, 2,9-dimethyl-	170	C ₁₂ H ₂₆	86
Octane, 2,3,7-Trimethyloctane	156	C ₁₁ H ₂₄	922
Tridecane, 7-methyl- 7-Methyltridecane	198	C ₁₄ H ₃₀	349
3-Ethyl-5-methylheptane	142	C ₁₀ H ₂₂	887
Phthalic acid, 2-ethylhexyl tetradecyl ester	474	C ₃₀ H ₅₀ O ₄	3364

Table 3 Shows the Chromatogram C₃ of Gas chromatography and mass spectrometry (GCMS) of compound C₃. The pure extract contains arrays of compounds, which listed as follows; 1,2-Benzenedicarboxylic acid, bis(2-methylpropyl) ester(C₁₆H₂₂O₄), Dibutyl phthalate(n-Butyl phthalate \$\$ Butyl phthalate) (C₁₆H₂₂O₄), Phthalic acid, hex-3-yl isobutyl ester(C₁₈H₂₆O₄), 1,2-Benzenedicarboxylic acid, bis(2-methylpropyl) ester(Diisobutyl phthalate)(C₁₆H₂₂O₄), 2-Benzenedicarboxylic acid, butyl 2-ethylhexyl ester (C₂₀H₃₀O₄), 1,2-Benzenedicarboxylic acid, diheptyl ester(Diheptyl phthalate)(C₂₂H₃₄O₄), Phthalic acid, 5-methylhex-2-yl heptyl ester(C₂₂H₃₄O₄), Phthalic acid, 4,4-dimethylpent-2-yl heptyl ester(C₂₂H₃₄O₄), Diisooctyl phthalate (1,2-Benzenedicarboxylic acid, ditridecyl ester)(C₃₄H₅₈O₄), Phthalic acid, octyl oct-3-yl ester(C₂₄H₃₈O₄), Phthalic acid, dodecyl octyl ester(C₂₈H₄₆O₄), Phthalic acid, heptyloctyl ester(1-Heptyl 2-octyl phthalate)(C₂₃H₃₆O₄).

Phthalic acid, bis(2-ethylhexyl) ester Bisoflex 81 (C₂₄H₃₈O₄), Phthalic acid, di(2-propylpentyl) ester(C₂₄H₃₈O₄), Phthalic acid, di(6-methylhept-2-yl) ester(C₂₄H₃₈O₄), Phthalic acid, 2-ethylhexyl tetradecyl ester (C₃₀H₅₀O₄), Phthalic acid, octyl 2-propylpentyl ester (C₂₄H₃₈O₄), Phthalic acid, 2-ethylhexyl undecyl ester(C₂₇H₄₄O₄), Phthalic acid, decyl 2-ethylhexyl ester(C₂₆H₄₂O₄), Diisooctylphthalate, Bis(6-methylheptyl) phthalate(C₂₄H₃₈O₄), Bis(2-ethylhexyl) phthalate (Phthalic acid, bis(2-ethylhexyl) ester, Bis(2-ethylhexyl) 1,2-benzenedicarboxylate (C₂₄H₃₈O₄), Bis(tridecyl) phthalate(1,2-Benzenedicarboxylic acid, ditridecyl ester)(C₃₄H₅₈O₄), Phthalic acid, octyl oct-3-yl ester(C₂₄H₃₈O₄), Phthalic acid, dodecyl octyl ester(C₂₈H₄₆O₄), Phthalic acid, heptyloctyl ester(1-Heptyl 2-octyl phthalate)(C₂₃H₃₆O₄).

Table 3 Chromatogram of Phthalic acid, di(2-propyl pentyl) ester, using gas chromatography and mass spectroscopy gc/ms method of analysis (Mass Peaks: 395,400)C₃

Name	Molecular weight	Structural formula	Retindex
1,2-Benzenedicarboxylic acid, bis(2-methylpropyl) ester	278	C ₁₆ H ₂₂ O ₄	1908
1,2 Benzenedicarboxylic acid, dibutyl ester	278	C ₁₆ H ₂₂ O ₄	2037
1,2-Benzenedicarboxylic acid, butyl 2-methylpropyl ester	278	C ₁₆ H ₂₂ O ₄	1973
Phthalic acid, hex-3-yl isobutyl ester	306	C ₁₈ H ₂₆ O ₄	2107
Phthalic acid, 4,4-dimethylpent-2-yl heptyl ester	362	C ₂₂ H ₃₄ O ₄	2484
Phthalic acid, 2,4-dimethylpent-3-yl heptyl ester	362	C ₂₂ H ₃₄ O ₄	2441
Bis(2-ethylhexyl) phthalate	390	C ₂₄ H ₃₈ O ₄	2704
Phthalic acid, di(2-propylpentyl) ester	390	C ₂₄ H ₃₈ O ₄	2704
Phthalic acid, di(6-methylhept-2-yl) ester	390	C ₂₄ H ₃₈ O ₄	2575
Phthalic acid, 2-ethylhexyl tetradecyl ester	474	C ₃₀ H ₅₀ O ₄	3364
Phthalic acid, octyl 2-propylpentyl ester	390	C ₂₄ H ₃₈ O ₄	2768

Table 4 Shows the Chromatogram E₃ of Gas chromatography and mass spectrometry (GCMS) of compound E₃. The pure extract contains arrays of compounds, which listed as follows; 2-Naphthalenol, 1,2-dihydro-, acetate(1,2-Dihydro-2-naphthalenyl acetate)(C₁₂H₁₂O₂), Azulene (Bicyclo[5.3.0]decapentaene, Cyclopentacycloheptene

\$\$Azunamic, Bicyclo(5.3.0)-1,3,5,7,9-decapentaene(C₁₀H₈), Naphthalene (Albocarbon ,Dezodorator, Moth flakes, Naphthalin, Naphthaline Naphthene, Tar camphor, White tar(C₁₀H₈), N-Methyl-9-aza-tricyclo [6.2.2.0(2,7)]dodec-2,4,6,11-tetraene-10-one(C₁₂H₁₁NO), 1H-Indene, 1-methylene-(1-Methylene-1H-indene)(C₁₀H₈),

Dodecane ($C_{12}H_{26}$), Undecane ($C_{11}H_{24}$), Octane, 2,3,7-trimethyl-(2,3,7-Trime thyoctane)($C_{11}H_{24}$), Dodecane, 2,7,10-trimethyl-, 2,7,10-Trimethyldodecane($C_{15}H_{32}$), 1,4-Methanonaphthalene, 1,4-dihydro Benzonorbornadiene \$\$ 1,4-Dihydro-1,4-methanona phthalene($C_{11}H_{10}$), Benzocycloheptatriene (5H-Benz[a] cycloheptene , 3,4-Benzotropilidene)($C_{11}H_{10}$),1H-Indene,1-ethylidene- (1-Ethylidene-1H-indene, Indene, 1-ethylidene-)($C_{11}H_{10}$), Eicosane, 10-methyl- (10-Methylicosane($C_{21}H_{44}$), Nonadecane, 2-methyl- 2-Methylnonadecane($C_{20}H_{42}$), 9-(2',2'-Dimethylpropanoilhydrazone)-3,6-dichloro-2,7-bis-[2-(diethylamino)-ethoxy] fluorine, N'-(3,6-Dichloro-2,7-bis[2-

(diethylamin($C_{30}H_{42}Cl_2N_4O_3$), Dibutyl phthalate (1,2-Benz enedicarboxylic acid, dibutyl ester, Phthalic acid, dibutyl ester, n-Butyl phthalate , Butyl phthalate)($C_{16}H_{22}O_4$), Hexacos-2,25-dione (2,25-Hexacosanedione)($C_{26}H_{50}O_2$), Octadecanal, 2-bromo-(2-Bromo octadecanal)($C_{18}H_{35}BrO$), 1-Pentacontanol($C_{50}H_{102}O$), Pentadecane, 2,6,10,14-tetramethyl-(Bute hydrocarbon, Norphytan, Norphytane, Pristan, Pristane,2,6,10,14-Tetram ethyl) ($C_{19}H_{40}$),1-Butanol, 3-methyl-, benzoate(Isopentyl alcohol, benzoate, Benzoic acid isoamyl ester)($C_{12}H_{16}O_2$), Benzoic acid, 2-ethylhexyl ester(Ethylhexyl benzoate)($C_{15}H_{22}O_2$).

Table 4 Chromatogram $C_{30}H_{42}C_{12}N_4O_3$ - 9-(2',2'-Dimethylpropanoilhydrazone)-3,6-dichloro-2,7-bis-[2-(diethylamino)-ethoxy]fluorene) using gas chromatography and mass spectroscopy gc-ms method of analysis (Masspeaks: 395,400)E₃

Name	Molecular weight	Structural formula	Retindex
2-Naphthalenol, 1,2-dihydro-, acetate	188	$C_{12}H_{12}O_2$	1476
AzuleneBicyclo[5.3.0]decapentaene	128	$C_{10}H_8$	1069
Naphthalene	128	$C_{10}H_8$	1231
9-(2',2'-Dimethylpropanoilhydrazone)-3,6-dichloro-2,7-bis-[2-(diethylamino)-ethoxy]fluorene	576	$C_{30}H_{42}Cl_2N_4O_3$	4330

Table 5 shows the Chromatogram F₃ of Gas Chromatography and Mass Spectrometry (GCMS) of compound F₃. The pure extract contains arrays of compounds, which listed as follows; Dodecane, ($C_{12}H_{26}$), Bis(2-ethylhexyl) phthalate (Phthalic acid, bis(2-ethylhexyl) ester, Bis(2-ethylhexyl) 1,2-benzenedicarboxylate,Bisoflex 81($C_{24}H_{38}O_4$), Eicosane, 10-methyl- $(C_{21}H_{44})$, Nonadecane($C_{20}H_{42}$), Phthalic acid, hex-3-yl isobutyl ester($C_{18}H_{26}O_4$), Heptane, 5-ethyl-2-methyl- (5-Ethyl-2-methylheptane, Heptane, 3-ethyl-6-methyl)($C_{10}H_{22}$),

Dibutyl phthalate ($C_{16}H_{22}O_4$), Hexadecane, 2,6,10,14-tetramethyl- (Phytan, Phytane, 2,6,10,14-Tetramethylhexadecane)($C_{20}H_{42}$), 1,2-Benzenedicarboxylic acid, bis(2-methylpropyl) ester($C_{16}H_{22}O_4$), Pentadecane, 2,6,10,14-tetramethyl- Butyhydrocarbon($C_{19}H_{40}$), Eicosane($C_{21}H_{44}$), Aspidofractinine-3-methanol, (2.alpha.,3.beta.,5.alp ha.)($C_{20}H_{26}N_2O$), 1,4-Methanonaphthalene, 1,4-dihydro- (Benzonorborna diene,1,4-Dihydro-1,4-methanonaphthalene ($C_{11}H_{10}$), Benzocycloheptatriene($C_{11}H_{10}$), (Figures 2–6).

Table 5 Chromatogram Terephthalic acid, dodecyl 2-ethylhexyl ester, using gas chromatography and mass spectroscopygc/msmethod of analysis (Masspeaks:395,400) F₃

Name	Molecular weight	Structural formula	Retindex
Dodecane	170	$C_{12}H_{26}$	1214
Undecane	156	$C_{11}H_{24}$	1115
Bis(2-ethylhexyl) phthalatePhthalic acid, bis(2-ethylhexyl) ester	390	$C_{24}H_{38}O_4$	2704
Eicosane, 10-methyl-	296	$C_{21}H_{44}$	2045
Nonadecane, 2-methyl-	282	$C_{20}H_{42}$	1945
Hexadecane, 2,6,10,14-tetramethyl- 2,6,10,14-Tetramethylhexadecane	282	$C_{20}H_{42}$	1753
Terephthalic acid, dodecyl 2-ethylhexyl ester	446	$C_{28}H_{46}O_4$	3165

The antibacterial effect of the purified compounds on selected clinical Gram negative (*E.coli*), Gram-positive (*Bacillus subtilis*) and (*Aspergillus flavus*) at 10,5,2.5 and 1.25 μ g/mL is presented in Table 6. All the compounds showed remarkable inhibitory effect on the selected organisms. Compound A1 has the highest zone of inhibition of 28.0 mm at 10 μ g/mL against *E. coli* while, Compounds E3 and F3 did not show any inhibitory effect on the bacterium at all the concentrations

used. All the compounds showed significant inhibitory effects on *Bacillus subtilis* and *Aspergillus flavus* at all the concentrations except F3 at concentration of 1.25 μ g/mL. Of all the compounds, B3 has the highest zone of growth inhibition of 30.0mm against *E. coli*. There was significant difference in the diameters of zones of growth inhibition of the compounds on the organisms ($p < 0.0001$).

Table 6 Antimicrobial effect of essential oil purified compounds on selected test bacteria and fungus

Compound	<i>E.coli</i>				<i>Bacillus</i>			<i>subtilis</i>		<i>Aspergillus flavus</i>		
Concentration (μ g/ml)	10	5	2.5	1.25	10	5	2.5	1.25	10	5	2.5	1.25
A3	24	15	9	7	23	10	5	2	24	18	10	6
B3	30	23	10	7	17	15	10	8	23	18	10	7
C3	22	19	5	3	18	12	5	2	20	17	10	9
E3	0	0	0	0	17	17	10	2	8	5	2	0
F3	0	0	0	0	22	18	2	0	17	16	0	0

Table 7 shows the synergistic antibacterial activities of the purified compounds on the selected organisms at 10, 5, 2.5, 1.25 μ g/ml. There was marked inhibitory effect observed in all the combined compounds with the highest zone of growth inhibition (36.0mm) recorded against *E. coli* with combination of Compounds A1 and A3.

Combined compounds A₁ + A₃, B₃ + C₃ and E₃ + F₃ showed diameters of inhibition of 32.0mm, 30.0mm and 20.0 mm against *Aspergillus flavus*, respectively. There was significant difference in the diameters of zones of growth inhibition of the compounds on the organisms ($p < 0.0001$).

Table 7 Synergistic antimicrobial effect of purified compounds on selected test bacteria and fungus

Compound	<i>E.coli</i>				<i>Bacillus subtilis</i>				<i>Aspergillus flavus</i>			
Concentration μ g/ml	10	5	2.5	1.25	10	5	2.5	1.25	10	5	2.5	1.25
C3 + A3	36	31	25	16	30	28	19	14	32	29	20	15
B3 + C3	32	29	27	18	20	17	12	10	30	20	16	8
E3 + F3	30	26	15	10	25	17	14	12	20	15	10	5
A3 + B3	28	26	20	15	27	20	18	15	18	15	10	6

Key: C₃ + A₃, Phthalic acid, di(2-propyl pentyl) ester; Aspido fractinine-3-methanol,(2.alpha.,3.beta.,5.alpha.);

B₃ + C₃, Phthalic acid, 2-ethylhexyl tetradecylester + Phthalic acid, di(2-propyl pentyl) ester; E₃ + F₃, 9-(2',2'-Dimethyl propanoilhydrazono)-3,6-dichloro-2,7-bis-[2 (diethylamino)-ethoxy] fluorene + Terephthalic acid, dodecyl 2-ethylhexyl ester; A₃ + B₃, Aspido fractinine-3-methanol,(2.alpha.,3.beta.,5.alpha.)+Phthalic acid, 2-ethylhexyl tetradecyl ester.

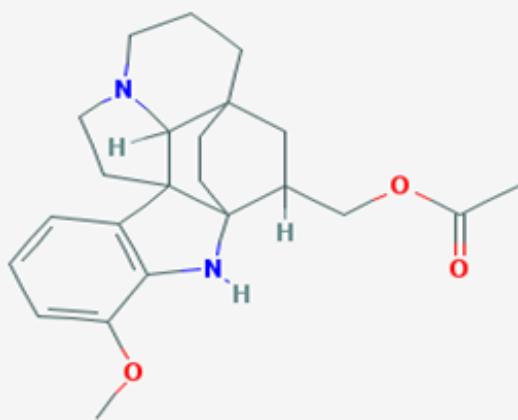


Figure 2 Structural profiling compound with the highest peak (A3): C₂₀H₂₆N₂OAspido fractinine-3-methanol, (2.alpha.,3.beta.,5.alpha.).

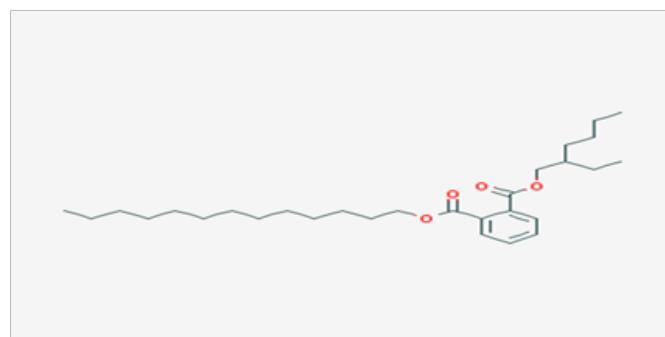


Figure 3 Structural profiling compound with the highest peak; (B3) C₃₀H₅₀O₄ Phthalic acid, 2-ethylhexyl tetradecyl ester.

Discussion

The purpose of this research work is to isolate, purify, chemical identification, antimicrobial and synergistic efficacy of extracted essential oils from methyl acetate extract of *Spondias mombin*. Plants produce a variety of compounds with antimicrobial activity. Some are always present while others are produced in response to microbial invasion.²⁰⁻²⁷ Identifying the most active antimicrobial compounds

of essential oils is cumbersome because essential oils are complex mixtures of up to 30 different constituents,²³⁻²⁵ and the composition of a particular essential oil may vary depending on the season of harvest, and the methods used to extract the oil.²⁶ The antimicrobial activity of Essential oil (EOs) of *Spondias mombin*, similar to all natural extracts is despondent on their chemical compositions and the amount of the single component. Many of the plant antimicrobial compounds are constitutively expressed by the plant and others can be synthesized as mechanism of self-defense in response to pathogens. Vegetable, spices and fruits like *Spondias mombin*, fruits with high level of EOs are excellent sources of natural compounds with activity against microorganism.¹⁵

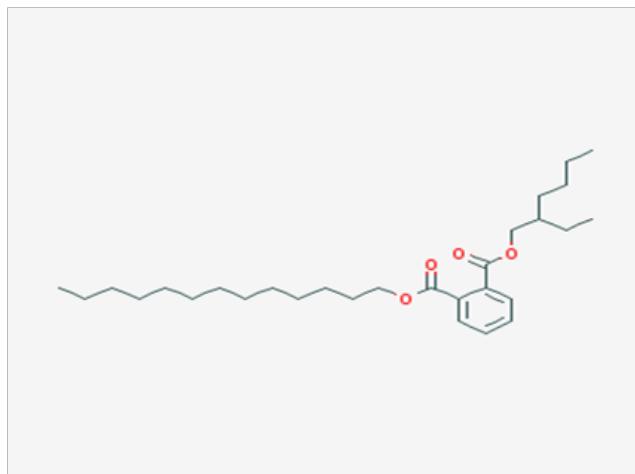


Figure 4 Structural profiling compound with the highest peak (C3) $C_{24}H_{38}O_4$ Phthalic acid, di (2-propylpentyl) ester.

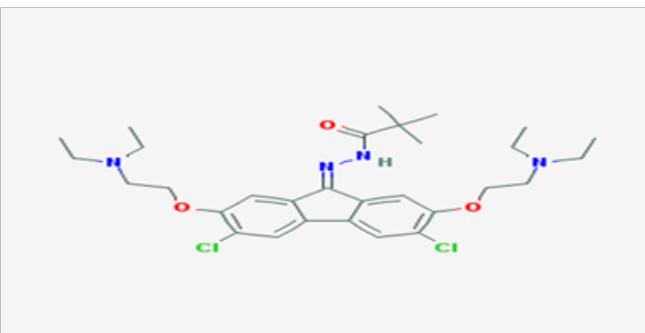


Figure 5 Structural profiling compound with the highest peak (E3) $C_{30}H_{42}Cl_2N_4O_3$ 9-(2',2'-Dimethylpropanoylhydrazone)-3,6-dichloro-2,7-bis-[2-(diethylamino)-ethoxy]fluorine.

In this study, ethylacetate was used to extract the Stem bark extract of *S. mombin* because most plant constituents are stored in vacuoles in the plant and are commonly extracted into solvents since it retains sufficient hydrophilicity to extract intra cellular (water-soluble) and extracellular compounds.^{16,17} The collected plant materials were air dried under shade before extraction. Higher temperature and sun drying were avoided to prevent degradation and decomposition of constituents that may be sensitive to extreme heat and light. After extraction, the filtrates were concentrated *in vacuo*. The crude extracts were stored in air tight containers to prevent microbial growth. The antimicrobial mode of action of essential oil observed in this study constituents have been performed on bacteria and fungi while little is known about their action on fungal (yeast and molds). Gram-negative

bacteria are generally less susceptible than Gram-positive bacteria.²⁸ The outer membrane of Gram-negative bacteria contain hydrophilic lipopolysaccharides (LPS), which create a mechanical barrier toward macromolecules and hydrophobic compounds, providing Gram-negative bacteria with higher tolerance toward hydrophobic antimicrobial compounds like those found in essential oils under the scope of this research work.²⁹ Antimicrobial compounds that act on the membrane can cause depolarization or increased permeability through various mechanisms. For example, the extracted essential oil from *S. mombin*, some antimicrobial peptides form pores^{30,31} while other compounds, such as certain essential oil constituents, have a fluidizing effect on the membrane.

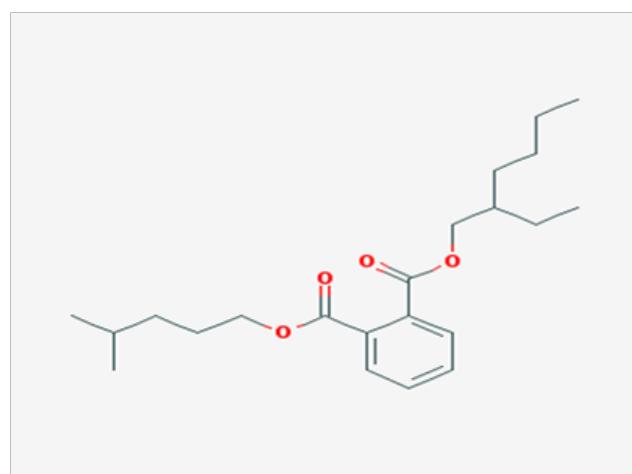


Figure 6 Structural profiling compound the highest peak (F3) $C_{28}H_{46}O_4$ Terephthalic acid, dodecyl 2-ethylhexyl ester.

From the results of the assessment of antimicrobial activities of the pure isolated compounds on selected test organisms (Table 6). It was revealed that all the pure extracts possessed marked antimicrobial activities against the test organisms. Similarly, the synergistic activities of these compounds (Table 7) results in significant antimicrobial activities recorded against the selected test organisms. This may be due to the synergistic effects of the combined compounds.

A method for preliminary fractionation of complex plants is by fractionation using vacuum liquid chromatography with solvents of various degrees of polarity. Each of the solvent which separates the components of extracts based on approximate polarity would result in distribution of the active constituents throughout several fractions. This principle was applied to the crude extract of the plant and was successively extracted with solvents and mixture of solvents with varying polarity. Each fraction was concentrated to dryness *in vacuo*. Thin-layer chromatography (TLC) analysis showed that the profiles of the fractions were distinctly different. Thus vacuum liquid chromatography was a good method for the preliminary fractionation of the crude extracts of the plant. It also served as a general purpose clean up method such that the compounds were obtained with high degree of purity. The compounds were characterized using Gas Chromatography-Mass Spectroscopy.

Most of the compounds identified by GCMS are essential oils (Tables 3-5). This may contribute to the observed antimicrobial activity *S. mombin* recorded against the susceptible test bacteria and fungi in this study. Aspidofractinine-3-methanol, Phthalic acid, 2-ethylhexyl tetradecyl ester, Phthalic acid, di(2-propylpentyl)ester, 9-(2',2'-Dimethylpropanoylhydrazone)-3,6-dichloro-2,7-bis-[2 diethylamino)-ethoxy]fluorine

ethoxy]fluorine); Terephthalic acid, dodecyl 2-ethylhexyl ester are basically the compound isolated from the *Spondias mombin*. Aspidofractinine-3-methanol are Alcohol with Antimicrobial, antifungal activity,²² Phthalic acid, 2-ethylhexyl tetradecyl ester), Ethyl, tetra, hexyl, decyl ester-Benzene-dicarboxylic acid diethyl ester has antimicrobial, antioxidant and hypocholesterolemic properties,³⁰ Phthalic acid, di(2-propylpentyl) ester are bicyclic aromatic hydrocarbon ester with anti-inflammatory, antimicrobial properties (31), 9-(2',2'-Dimethylpropanoilhydrazono)-3,6-dichloro-2,7-bis-[2-(3,6-dichloro-2,7-bis-[2-(diethylamino)-ethoxy]fluorine) are basically Fatty acid with antimicrobial, antioxidant, cytotoxic, antifungal and antitumor properties,²⁹ Terephthalic acid, dodecyl 2-ethylhexyl ester), Benzene-dicarboxylic acid diethyl ester has antioxidant, hypocholesterolemic activity.^{18,32}

Spondias mombin essential oil can affect both the external envelope of the cell and the cytoplasm, It should be noted that the compounds identified in this study like Benzoic acid and 2-pipondinone, phthalate, 7, 8, 9, octadecanal, Diisotyl phthalate have been reported to have permeability barrier provided by the cells membrane which is indispensable to many cellular functions including maintaining the energy status of the cell, membrane coupled energy transducing process, solute transport and metabolic regulation.^{19,20} *Spondias mombin* essential oil has been reported to inhibit the growth of bacterial cells and also inhibit the production of toxic bacterial metabolites, and appears to be more effective against Gram positive bacteria than Gram negative species; the effect which may likely due to differences in the cell membrane composition.²¹ From the results of the assessment of antimicrobial activities of the pure isolated compounds on selected test organisms (Table 6). It was revealed that all the pure extracts possessed marked antimicrobial activities against the test organisms.

Osuntokun reported the role of essential oils in the discovery of new drugs cannot be overemphasized, especially in an era of antimicrobial resistance. With essential oils, one may suggest that certain abundant and effective constituents such as thymol, β -carophyllenes and eugenol as major highlights so far for development of potent antimicrobials. This is due to the fact that they possess novel chemistry which could offer a remarkable opportunity for discovery of new drugs, many antimicrobial resistant organisms including methicillin resistant *staphylococcus aureus* (MRSA) can be controlled using essential oils. Researchers are therefore encouraged to tailor drug design towards the use of essential oils (especially eugenols and thymols) to curb antimicrobial resistance. Other pharmaceutical potentials in the treatment of infection.

The synergistic activities of these compounds (Table 7) results in significant antimicrobial activities recorded against the selected test organisms. This may be due to the synergistic effects of the combined compounds, for example, Phthalic acid, di (2-propyl pentyl) ester and Aspidofractinine-3-methanol, (2.alpha.,3.beta.,5.alpha.), Phthalic acid, 2-ethylhexyl tetradecyl ester and Phthalic acid, di (2-propyl pentyl) ester 9-(2', 2'-Dimethyl propanoilhydrazono)-3,6-dichloro-2,7-bis-[2-(diethylamino)-ethoxy]fluorine and Terephthalic acid, dodecyl 2-ethylhexyl ester. Aspidofractinine-3-methanol, (2.alpha.,3.beta.,5.alpha.) and Phthalic acid, 2-ethylhexyl tetradecyl ester. All isolated ester exhibits different degree of antimicrobial synergistic activity from the ethyl acetate stem bark extract of *Spondias mombin* due to combination of different array of essential oil.³⁰

Conclusion

In conclusion, *Spondias mombin* is a unique medicinal plant with various medicinal purposes, and the essential oil extracted from the ethyl acetate stem bark has proven to be a potent therapeutic and antimicrobial agents in the annals of nature's chemistry in drug discovery and treatment of various infectious diseases due to the fact that *Spondias mombin* possess different novel compound and its synergized with different novel compound which has become an indomitable force of nature in the treatment of infectious diseases.

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Conflicts of interest

The author declares there is no conflicts of interest.

Funding details

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References

1. Adeyemi OI, Ige OO, Olowu AA, et al. Combination with Tadalafil reduces the aphrodisiac activity of *Cola acuminata* and *Garcinia cola* in rats. *Nigerian Journal of Natural Products and Medicine*. 2016;20:167–173.
2. Aiyegoro OA, Okoh AI. Use of bioactive plant products in combination with standard antibiotics: implications in antimicrobial chemotherapy. *Journal of Medicinal Plants Research*. 2000;3(13):1147–1152.
3. Kambizi L, Afolayan AJ. An ethnobotanical study of plants used for the treatment of sexually transmitted diseases (njovhera) in Guruve District, Zimbabwe. *J Ethnopharmacol*. 2001;77(1):5–9.
4. Idowu TO, Onawumi GO, Ogundaini AO, et al. Antimicrobial constituents of *Chrysophyllum albidum* seed cotyledons. *Nigerian Journal of Natural Products and Medicine*. 2003;7(1):33–36.
5. Idowu TO, Ogundaini AO, Salau AO, et al. Doubly linked, A-type proanthocyanidin trimers and other constituents of *Ixoracoccinea* leaves and their antioxidant and antibacterial properties. *Phytochemistry*. 2010;71(17–18):2092–2098.
6. Ajao AO, Shonukan O, B Femi onadeko. Anti bacterial effect of aqueous and alcohol extracts of *Spondias mombin* and *Alchomea cordifolia*: 2 local antimicrobial remedies. *International Journal of crude drug Research*. 1985;23(2):67–72.
7. Abio KA, Ogunleye VO, Ashidi JS. Antimicrobial potential of *Spondias mombin*, *Croton zambesicus* and *Zygottitonia crocea*. *Phytother Res*. 1999;13(6):494–497.
8. Corthout J, Pieters L, Claeys M, et al. Antibacterial and molluscicidal phenolic acid from *Spondias mombin*. *Planta Med*. 1991;60(5):460–463.
9. Rodrigues K, Hasse M. Antimicrobial activities of secondary metabolites produced by endophytic fungi from *Spondias mombin*. *J Basic Microbiol*. 2000;40(4):261–267.
10. Kramer AE, Mosquera E, Ruiz J, et al. Ethanobotany and biological activity of plants utilized during pregnancy and child birth in the Peruvian Amazon. *Emanation*. 2006;4:31–35.
11. Njoku PC, Akumefula MI. Phytochemical and nutrient evaluation of *Spondias mombin* leaves. *Pak J Nutr*. 2007;6(6):613–615.

12. Ayoola PB, Adeyeye A, Onawumi OO. Trace elements and major mineral evaluation of *Spondias mombin*, *Vernoniaamygdalina* and *Momordicacharantia* leaves. *Pakistan Journal of Nutrition*. 2010;9(8):755–758.
13. Offiah VN, Anyanwu II. Abortifacient activity of an aqueous extract of *Spondiasmombin* leaves. *J Ethnopharmacol*. 1989;26(3):317–320.
14. Osuntokun OT, Olajubu FA. Comparative study of phytochemical and proximate analysis of seven Nigerian medicinal plants. *App Sci Res J*. 2014;2(1):10–26.
15. Nworu CS, Akah PA, Okoye FB, et al. The leaf extract of *Spondiasmombin* L. displays an anti-inflammatory effect and suppresses inducible formation of tumor necrosis factor- α and nitric oxide (NO). *J Immunotoxicol*. 2011;8(1):10–16.
16. Merken HM, Beecher GR. Liquid chromatographic method for the separation and quantification of prominent flavonoid aglycones. *J Chromatogr*. 2000;897(1–2):177–184.
17. Santos Buelga C, Scalbert A. Proanthocyanidins and tannin-like compounds—nature, occurrence, dietary intake and effects on nutrition and health. *Journal of the Science of Food and Agriculture*. 2000;80(7):1094–1117.
18. López Lázaro M. Distribution and biological activities of the flavonoid luteolin. *Mini Rev Med Chem*. 2009;9(1):31–59.
19. Sikkema J, De Bont JA, Poolman B. Mechanisms of membrane toxicity of hydrocarbons. *Microbiol Rev*. 1995;59(2):201–222.
20. Bajpai VK, Sharma A, Baek KH. Antibacterial mode of action of *Ginkgo biloba* leaf essential oil: Effect on morphology and membrane permeability. *Bangladesh Journal of Pharmacology*. 2015;1(2):337–350.
21. Redo MC, Rios JL, Villar A. A review of some antimicrobial compounds isolated from medicinal plants reported in the literature 1978–1988. *Phytotherapy Research*. 1989;3(4):117–125.
22. Roller S. *Natural Antimicrobials for the Minimal Processing of Foods*. Cambridge: Woodhead Publishing, Limited; 2003.
23. Delaquis PJ, Stanich K, Girard B, et al. Antimicrobial activity of individual and mixed fractions of dill, cilantro, coriander and eucalyptus essential oils. *Int J Food Microbiol*. 2002;74(1–2):101–109.
24. Djenane D, Yanguela J, Montanes L, et al. Antimicrobial activity of *Pistacia lentiscus* and *Saturejamontana* essential oils against *Listeria monocytogenes* CECT 935 using laboratory media: efficacy and synergistic potential in minced beef. *Food Control*. 2011;22(7):1046–1053.
25. Espina L, Somolinos M, Lorán S, et al. Chemical composition of commercial citrus fruit essential oils and evaluation of their antimicrobial activity acting alone or in combined processes. *Food Control*. 2011;22(6):896–902.
26. Nannapaneni R, Chalova VI, Crandall PG, et al. *Campylobacter* and *Arcobacter* species sensitivity to commercial orange oil fractions. *Int J Food Microbiol*. 2009;129(1):43–49.
27. Trombetta D, Castelli F, Sarpietro MG, et al. Mechanisms of antibacterial action of three monoterpenes. *Antimicrob Agents Chemother*. 2005;49(6):2474–2478.
28. Nikaido H. Molecular basis of bacterial outer membrane permeability revisited. *Microbiol Mol Biol Rev*. 2003;67(4):593–656.
29. Cotter PD, Hill C, Ross RP. Food microbiology: bacteriocins: developing innate immunity for food. *Nat Rev Microbiol*. 2005;3(10):777–788.
30. Fantner GE, Barbero RJ, Gray DS, et al. Kinetics of antimicrobial peptide activity measured on individual bacterial cells using high-speed atomic force microscopy. *Nat Nanotechnol*. 2010;5:280–285.
31. Osuntokun OT, Ogunleye AJ. Prospects of essential oils in drug discovery. *Adv Cytol Pathol*. 2017;2(1):17–19.
32. Oludare temitope Osuntokun, AO Oluduro, TO Idowu, et al. Assessment of Nephrotoxicity, Anti-inflammatory and Antioxidant properties of Epigallocatechin, Epicatechin and Stigmasterol phytosterol (synergy) Derived from ethyl acetate stem bark extract of *Spondiasmombin* on Wister Rats Using Molecular method of analysis. *Journal of Molecular Microbiology*. 2017;1(1):1–11.