

Mitigation of salt induced stress via arbuscular mycorrhizal fungi (*Rhizophagus irregularis*) inoculation in *Cucurbita maxima* Duch

Abstract

It has been projected that about 7% of the earth's agricultural land is exposed to extreme soil salinity levels. High presence of salts in soil reduces plant water content and nutrient uptake thereby disrupting the dissemination of ions at both the cellular and the whole-plant levels, ultimately inducing osmotic and ionic disparities. The current research was carried out to examine the role of arbuscular mycorrhizal fungi (*Rhizophagus irregularis*) in alleviating adverse effects of salt stress in *Cucurbita maxima*. Physicochemical properties of the experimental soils analysis (saline and garden soils) indicated significant ($p=0.05$) differences between the two soil types in; pH, total nitrogen, available phosphorus, Ex. Ca, Ex. Mg, Ex. K, OC, Ex. Na and EC. Saline soil treatment significantly ($p=0.05$) reduced photosynthetic pigments contents (chlorophyll a, b and carotenoids), minerals (N, P, K, Mg and Ca), leaf relative water content (LRWC), shoot length, dry weight as well as percentage arbuscular mycorrhizal fungi colonization (45.45 to 20.34%) and mycorrhizal dependency (100.00% to 13.87%). The effects of mycorrhizal symbiotic association on *Cucurbita maxima* showed improvements on the growth and physiology of *Cucurbita maxima*. Using different mechanisms *Cucurbita maxima* by itself or in association with arbuscular mycorrhizal fungi (*Rhizophagus irregularis*) can tolerate or survive soil salinity. However, symbiotic association of the fungus *R. irregularis* with roots of *Cucurbita maxima* gives the plant the ability to resist the stress as a result of morphological and physiological vicissitudes and improved vigour/water content, adjusted rate of K^+/Na^+ , extensive network of the mycorrhizal plant roots and enhanced nutrient uptake are all among the processes that made the mycorrhizal inoculated plant to survive under severe salt stress conditions.

Keywords: cucurbitaceae, glomeraceae, mycorrhiza, salinity salt tolerance

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Introduction

Soil solution is regarded as one of the best medium of plant growth and development because it contains various important soluble salts.¹ A key threat to worldwide crop production is the build-up of salts in the soil.^{2,3} According to estimates, 7% world's agricultural lands are exposed to very high soil salinity levels.³ Thus, Salinity can be defined as the soil content of water-soluble salts, predominantly sodium, chloride, potassium, magnesium and calcium. When the electrical conductivity of a soil saturated paste extract (ECE) measures up to 4dS/m or higher, which is comparable to 40mm NaCl, are distinctive of saline soils. This definition of salinity is driven from the ECE that significantly reduces the yield of most crops. Under natural conditions, the salts in soil contaminated by sea water are usually mixtures of several cations and anions.

These soluble salts (such as; sodium, chloride, potassium, magnesium and calcium) in the soils are readily absorbed by roots of plants and translocate to the aerial and different plant parts where these salts are essential to performing several metabolic activities. Nevertheless, when these water-soluble salts are present in the soil in abundance, they inhibits plant water and nutrient ascent and consequently upset the proper dissemination of ions in the cellular and the whole-plant levels, thus prompting osmotic stress and ionic disproportions.¹ This extreme and sudden change results in diminutive growth and development of plants mostly resulting in the death of the plant. The excessive accumulation of water-soluble salts such as Na^+ and Cl^- in the tissues of plants results in oxidative mutilation (mostly regarded as secondary stress), the plant membranes integrity is severely damaged (damage to proteins, lipids and nucleic acids),

marring the actions of biocatalysts and workings of chloroplast (photosynthetic apparatus), This can be attributed to the generation and lethal properties of reactive oxygen species (ROS) which are produced under soil salinity stress conditions.^{1,4,5}

Beneficial microorganisms such as arbuscular mycorrhizal fungi (AMF) are capable of colonizing plants in their regular environment and in stressed environments. Some of these microorganisms are beneficial and has been observed to play important roles in the improvements of plant tolerance and performance especially in environments that experience abiotic stress thereby giving rise to improved crop production and yield.⁶ It has been reported that AM fungi forms association with about 80% land-dwelling plant species.⁷ Many researches has reported the beneficial effects of inoculating plants with AM fungi under saline soil conditions and its role in salinity tolerance under such conditions. AM fungi ubiquitous in the different ecosystem types³ and have been reported in extreme environments such as sub-polar regions, arid regions, tropical regions and even in aquatic ecosystems.⁸ The symbiotic association between plant and AM fungi are extremely antique and dates back to about 400 million years.³ Every plant species has its dissimilar degree of mycorrhizal dependency, example; *Vicia faba* (faba bean) is highly dependent on mycorrhizas and depends highly on its association for subsequent growth and establishment.⁹

Cucurbita maxima belong to the family Cucurbitaceae and is a climbing, annual, herbaceous vine, reaching a length of 4m or more. Leaves are hispid, rounded, 15-30cm in diameter, heart-shaped at the base, shallowly 5-lobed, with finely toothed margins, and often mottled on the upper surface. Flowers are bell-shaped, erect, yellow

and about 12 centimetres long. Fruit is large, variable in shape, fleshy, with a yellow pulp. Seeds are ovoid or oblong, compressed, and about 1.3 centimetres long.¹⁰ *C. maxima* varieties are used for preparation of many dishes. In Cameroon, Nigeria and other Western African countries, seeds of *C. maxima* are widely used as a vegetable, roasted and salted, or ground into a thick paste that is mixed with vegetables in cooking.¹¹ The medicinal uses of *C. maxima* include the dried pulp, in the form of confection, used as remedy for hemoptysis and hemorrhages from the pulmonary tract.¹⁰

Rhizophagus irregularis belongs to the family *Glomeraceae*. Spores colour of *Rhizophagus irregularis* is white, cream, and yellow-brown. Shape is elliptical with irregularities. Size is generally between 40-140µm, Hyphae shape is Cylindrical or slightly flared.¹² Size-Width: 11-18µm. In numerous scientific studies *Rhizophagus irregularis* has been publicized to increase phosphorus uptake in multiple plants as well as improve soil aggregation due to hyphae; because of these qualities, *Rhizophagus irregularis* is commonly found in mycorrhizal based fertilizers. *Rhizophagus irregularis* colonization peaks earlier than many of the other fungi in the *Glomus* genus.¹² There tends to be extensive hyphal networking and intense intraradical spores associated with older roots of host plants. At times the spores

are densely clustered or patchily distributed, depending on the host species. When the spores are heavily clustered, mycorrhizologists and others will tend to mistake *Rhizophagus irregularis* for *Glomus fasciculatum*.¹²

Materials and methods

Study area

Saline soil and salt water were collected from the saline ecosystem of Iwuochang, Ibeno Local Government Area (Latitude 4.56°N and Longitude 7.57°E), Akwa Ibom State, Nigeria, with an annual rainfall of about 4021 mm and mean temperature variation of 22–31°C. The experiment was set up in a safe and secured environment at Mbioto 1, Etinan Local Government Area (Latitude 4.51°N and Longitude 7.50°E), Akwa Ibom State, Nigeria, with an annual rainfall of about 4000mm and mean temperature variation of 26–36°C (AKSG, 2008). Non-saline soil for the control and non-saline treatments was obtained from a farmland in Mbioto 1, Etinan Local Government Area; fresh water was used for watering the non-saline and control treatments. A map showing the saline water/soil collection and experimental set-up locations is presented in Figure 1 (Table 1).



Figure 1 Map showing saline water/soil collection and experimental set-up locations (Source: Field Data).

Source of arbuscular mycorrhizal (am) fungi

The AM Fungi (*Rhizophagus irregularis*) inoculum used in this experiment (60-65 spores per 5g) was purchased from International Institute of Tropical Agriculture (IITA) Ibadan, Nigeria.

Physico chemical properties of experimental soils

The soil samples were taken and air-dried at room temperature and ground in a wooden mortar to pass through a 2mm mesh sieve and stored in labelled bags. Sub-samples were taken from each soil

sample and analyzed for physico-chemical properties of the soil. All soil analyses were carried out in the Soil Science Department of the University of Uyo, Uyo, Akwa Ibom State. Soil samples were analyzed following the standard procedures outlined by the Association of Official Analytical Chemist procedure for wet acid digestions.¹³

Table 1 Experimental design

Treatments	Meaning
S- M-	- Salinity, - Mycorrhiza
S+ M-	+ Salinity, - Mycorrhiza
S+ M+	+ Salinity, + Mycorrhiza (<i>Rhizophagus irregularis</i>)
S- M+	- Salinity, + Mycorrhiza

Estimation of photosynthetic pigments

The photosynthetic pigments (such as chlorophyll a, chlorophyll b and carotenoids) were extracted from leaves of *C. maxima* plants using dimethyl sulfoxide (DMSO) as recommended.¹⁴ Absorbance was determined spectrophotometrically at 480, 510, 645, and 663nm (T80 UV/VIS Spectrometer, PG Instruments Ltd, USA). DMSO was used as blank.

Determination of mineral content

The plant samples were sent to Ministry of Science and Technology, Akwa Ibom State for mineral analysis. Mineral contents: Nitrogen (N) was determined using the Macro-Kjeldahl method while calcium (Ca), magnesium (Mg), potassium (K), sodium (Na) and phosphorus (P) of plant samples were determined by atomic absorption spectrophotometer (AAS), flame photometry and spectrophotometry according to the methods of AOAC.¹⁵

Determination of shoot length

Measurement of shoot length was taken at nine (9) weeks after planting following seedling emergence using a measuring tape.

Determination of leaf area

Measurements were obtained using graph paper (grid method). The area (A) of the leaf was determined by tracing the outlines of the leaves on a standard graph paper. The area covered by the outline was then calculated (one small square on the graph represents 1cm²). The correlation factor (K) was determined by dividing the area (A) by product of length x breadth of the leaf. Thereafter, the leaf area for each plant was determined using the formula:

$$A = L \times B \times K$$

Where: A=the traced area, L=Leaf length, B=Leaf width, K=Correction factor

Determination of leaf dry weight

Leaf dry weight was determined after oven drying the leaves to constant weight at 70°C

Leaf relative water content

Two (2) young fully expanded leaves were collected from about two plants per replicate. The leaves were detached from the stem and weighed to determine the fresh weight (FW). To determine the turgid weight (TW), leaves were floated in distilled water inside a closed petri dish. Leaf samples were weighed periodically after gently

wiping the water from the leaf surface with tissue paper until a steady state was achieved. At the end of the imbibition period, leaf samples were placed in a preheated oven at 80°C for 48 hours to determine the dry weight (DW). Values of FW, TW, and DW were used to calculate LRWC using the formula (Kaya *et al.*, 2003):

$$LRWC(\%) = \frac{(FW-DW)}{TW-DW} \times 100$$

Quantification of arbuscular mycorrhizal colonization in plant roots

Feeder roots of about 2-4cm of *T. occidentalis* were separately collected, fixed in 50% ethanol and stored for colonization assessment. The fixed roots were rinsed in tap water before clearing in 10% KOHw/v and autoclaved for about 15 minutes at 121°C. Cleared roots were collected on a fine sieve and rinsed with water several times before being transferred into the staining solution. Staining of the plants roots was carried out using 5% ink diluted in vinegar (5% acetic acid). The roots segments were soaked in the ink and left in staining solution at room temperature for one day. Stained roots were later destained in 50% glycerol for 1 hour.¹⁶

Stained roots were randomly dispersed in a 9cm diameter Petri plate with grid lines. Vertical and horizontal gridlines were scanned at x40 magnification with a dissecting microscope. The proportion of root length that is mycorrhizal and total root length can then be calculated from a conversion factor derived from the total length of grid lines and the area of the dish. A minimum of 100 intersections was used to assess the stained root samples; the samples were re-randomized and counted several times. Mycorrhizal root colonization was thus determined by estimation of percentage of root segments containing hyphae, arbuscules and vesicles.¹⁷

$$MC = \frac{\text{Total number of roots infected intersecting gridlines}}{\text{Total number of roots intersecting gridlines}} \times 100$$

Determination of mycorrhizal dependency (md)

Mycorrhizal dependency (MD) was calculated according to the following formula:

$$MD = \frac{DW \text{ inoculated Plants} - DW \text{ non-inoculated Plants}}{DW \text{ inoculated Plants}} \times 100$$

Statistical analysis

The study was conducted using complete randomized design with three (3) replicates for each plant. All data in the present study were subjected to analysis of variance (ANOVA) using Statistical package for Social Sciences and data are presented as standard error of mean (\pm S.E.M.) of triplicate experiments. The differences between the means were separated and compared using the Duncan's multiple range tests. However, a probability level of p=0.05 was considered statistically significant.

Results

Physicochemical properties of the experimental soils (saline and non-saline soil)

The t-test analysis carried out on the physicochemical properties of the experimental soils (saline and garden soils) indicated significant (p=0.05) differences between the two soil types in; pH, total nitrogen, available phosphorus, silt, clay, sand, Ex. Ca, Ex. Mg, Ex. K, OC, Ex. Na and EC. However, there was no significant (p=0.05) difference in Ex. Acidity, ECEC and base saturation (Table 2).

Table 2 Physicochemical properties of the experimental soils

S/No	Parameters	Garden soil	Saline soil	t-values
1	pH	6.78	7.75	-56.655*
2	Total Nitrogen (%)	2.27	0.49	6.928*
3	Available P. (mg/kg)	36.31	24.66	663.929*
4	Silt (%)	4	5.6	-51.995*
5	Clay (%)	4.2	12	-193.742*
6	Sand (%)	92.04	82.4	261.909*
7	Ex. Ca (cmol./kg)	5.25	2.97	-148.956*
8	Ex. Mg (cmol./kg)	4.36	3.8	23.714*
9	Ex. Na. (cmol./kg)	0.41	8.81	1.000*
10	Ex. K. (cmol./kg)	6.98	1.48	43.301*
11	Organic Carbon (%)	5.61	1.61	-599.00*
12	Exchangeable acidity (meq/100g)	3.56	3.2	20.785
13	ECEC (cmol./kg)	20.56	20.26	-228.669
14	Base saturation (%)	82.68	84.2	-64.341
15	EC(dS/m)	0.32	7.8	-93.260*

*Significant at t = 0.05, Ex, exchange; ECEC, effective cation exchange capacity; EC, electrical conductivity

Effect of arbuscular mycorrhizal fungi (amf) inoculation on the photosynthetic pigments of *c. maxima* grown in saline soil

The content of photosynthetic pigments of *Cucurbita maxima* (such as chlorophyll a, chlorophyll b and carotenoids) grown in saline soils taken at 9 WAP were significantly (p=0.05) reduced when compared to the control while inoculation with AMF (*R. irregularis*) significantly (p=0.05) increased these pigments in the test plant (Figure 2).

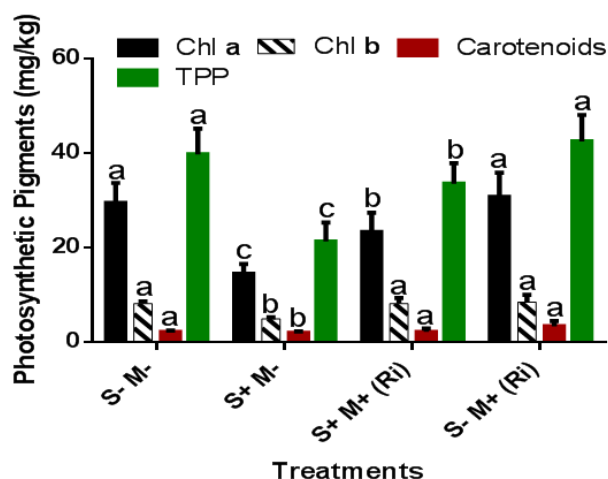


Figure 2 Effects of AMF inoculation on the photosynthetic pigments content of *C. maxima* grown in saline soil. *Means within of each coloured bar followed by different letters are significantly different at p=0.05 according to Duncan's Multiple Range Test. S- (No salinity), M- (No mycorrhiza), S+ (Plus salinity), M+ (Plus mycorrhiza), (Ri) – *Rhizophagus irregularis*.

Effect of arbuscular mycorrhizal fungi (amf) inoculation on the mineral nutrient contents and biomass yield of *c. maxima* grown in saline soil

Mineral nutrient contents of *C. maxima* P, K, Mg and Ca were significantly (p=0.05) reduced in saline soil treatments when compared to the control while, N showed slight decrease in saline soil treatments, while Na increased in saline soil treatments (Table 3). Inoculation with AMF (*Rhizophagus irregularis*) significantly (p=0.05) increased the mineral nutrient contents, leaf relative water content and biomass yield in the test plant in saline and non-saline soil treatments (Table 3).

Table 3 Effects of arbuscular mycorrhizal fungi (AMF) inoculation on the mineral nutrient contents of *C. maxima* grown in saline soil

Parameters	S- M-	S+ M-	S+ M+ (Ri)	S- M+ (Ri)
N (%)	*5.03 ^a	0	3.44 ^b	5.28 ^a
P (mg/kg)	754.10 ^b	0	510.10 ^c	894.40 ^a
K (mg/kg)	4610.10 ^b	0	2215.10 ^c	4811.10 ^a
Mg (mg/kg)	560.14 ^b	0	418.00 ^c	601.10 ^a
Ca (mg/kg)	3008.10 ^b	0	1620.20 ^c	3240.00 ^a
Na (mg/kg)	387.70 ^a	0	1320.20 ^c	309.10 ^b
Na+/K+ ratio	0.08 ^b	0	0.59 ^a	0.06 ^b
Leaf Area (cm ²)	56.21 ± 0.72 ^b	0	25.44 ± 1.32 ^c	60.31 ± 1.12 ^a
Shoot Length (cm)	54.61 ± 3.99 ^a	0	17.73 ± 0.45 ^b	59.76 ± 2.13 ^a
Leaf Relative Water Content (LRWC) (%)	38.00 ± 2.51 ^a	0	36.36 ± 1.66 ^b	42.65 ± 1.57 ^a
Total Dry Weight (gplant-1)	4.06 ^a	0	1.18 ^b	4.43 ^a

*Mean of three replicates. ^aMeans within of each column followed by different letters are significantly different at p=0.05 according to Duncan's Multiple Range Test. S- (No salinity), M- (No mycorrhiza), S+ (Plus salinity), M+ (Plus mycorrhiza), (Ri)-*Rhizophagus irregularis*, 0.00 (Plants died off).

Percentage AMF root colonization of *C. maxima* was significantly (p=0.05) reduced in saline soil treatments of the test plants (Table 4). However, inoculated saline soil treatment showed significant (p=0.05) increase in the %AMF root colonization.

Discussion

Analysis of the saline and garden soils used in this study revealed significant (p=0.05) variations in their soil physico-chemical parameters. Significant (p=0.05) increase in parameters such as pH, EC and Ex Na⁺ was observed in the saline soil while there was a decrease in organic carbon, total nitrogen and available phosphorus in saline soil. This observation is in line with the work of Miller¹⁸ who reported an increase in pH and EC in saline soils in New Jersey due to salt stress. Deleke¹⁹ also made similar findings as they observed an increase in pH, EC and Ex Na⁺ in saline soils and a decrease in organic carbon, organic matter, total nitrogen and phosphorus in salinity influenced soils in Nigeria. Soil organic carbon content is influenced by two opposing factors: reduced plant inputs and reduced rates of decomposition.²⁰

Table 4 Arbuscular mycorrhizal fungi (AMF) colonization of *C. maxima* grown in saline soil

Non-inoculated treatment	Root colonization (%)	Mycorrhizal dependency (%)	Inoculated treatments	Root colonization (%)	Mycorrhizal dependency (%)
S-M-	0	0	S+M+ (Ri)	*20.34 ^d	100.00 ^a
S+M-	0	0	S-M+ (Ri)	45.45 ^b	13.87 ^d

*Mean of three replicates. ^aMeans within of each column followed by different letters are significantly different at p=0.05 according to Duncan's Multiple Range Test. S- (No salinity), M- (No mycorrhiza), S+ (Plus salinity), M+ (Plus mycorrhiza), (Ri)-*Rhizophagus irregularis*.

Salt stress significantly (p=0.05) reduced the total photosynthetic pigments contents in *C. maxima* in both mycorrhizal and non-mycorrhizal saline treated plants. Non-mycorrhizal plants in saline soil treatments were more severely affected than *C. maxima* inoculated with *R. irregularis*. Amira²¹ also recorded decrease in chlorophyll 'a', 'b', and total chlorophyll in *V. faba* exposed to saline stress. Also, Tort²² reported that exposure of *Hordeum vulgare* L. (barley) to different salinity levels 0, 120, and 240mm of NaCl results in the reduction in chlorophyll 'a', chlorophyll 'b' and total chlorophyll content. Under stress conditions, the photosynthetic activity of the plant is often affected due to the alteration in the level and activity of various photosynthetic enzymes such as rubisc,²³ sucrose phosphate synthase and fructose-1,6-bisphosphatase.²⁴ and PEP carboxylase.²⁵ All these could have led to reduction in photosynthesis and subsequent lower biomass accumulation resulting in growth reduction in saline soil grown plants. The influence of AMF on photosynthetic pigments of AMF inoculated plants growing under salinity stress conditions has been reported by several researchers.²⁶⁻²⁹ The increased rate of photosynthesis in AMF-colonized plants under salinity stress has been correlated with lower intercellular CO₂ concentration in mycorrhizal plants, since the higher photosynthetic capacity increases water use efficiency for the assimilation of more carbon per unit water transpiration.²⁶

The mineral (N, P, K, Mg and Ca), leaf relative water content and biomass yield of *C. maxima* were significantly (p=0.05) reduced in saline soil treatments in this study, while foliar uptake and accumulation of Na⁺ was significantly (p=0.05) increased in saline soil treatments than in non-saline treatments. Comparing the influence of Na⁺ foliar uptake on minerals, leaf relative water content and biomass yield, it was observed that Na⁺ accumulation had negative effects on the minerals, leaf relative water content and biomass yield of *C. maxima* (Figure 3) (Figure 4). This observation agrees with the work of Robert

et al.,³⁰ who reported that exposure to NaCl injures plants in part through lowered soil water potential and the ensuing osmotic stress, but ultimately it may be more injurious via direct toxicity of Na⁺ ions. Ullah et al.,³¹ reported that irrigation of tomato plants with seawater increased the uptake of sodium chloride and decreased the uptake of P and Fe. Increased concentration of Na⁺ and Cl⁻ in soil solution competes with the uptake of vital ions such as Ca⁺², P, K⁺, Mg⁺², and N and alters the ideal salt ratios in the soil solution, therefore affecting plant nutrient acquisition and restricting plant growth and biomass. In this study, it was also observed that the minerals composition of *C. maxima* under saline and non-saline treatments was significantly increased with AMF *R. irregularis* inoculation. A higher N, P, K, Mg and Ca concentration in mycorrhizal than non-mycorrhizal plants can favourably alleviate the toxic effects of NaCl by inducing a higher K⁺/Na⁺ rate leading to salt adaptation.³² Cantrell³³ reported increased N, P, K, Mg and Ca uptake in mycorrhizal lettuce.

AMF root colonization (MC) of *C. maxima* was significantly (p=0.05) reduced in saline soil treatments when compared to non-saline treatments. This is in agreement with earlier studies reporting that addition of salt to soil inhibits hyphal growth, which subsequently reduces the spread of mycorrhizal colonization.^{32,34} It was also observed in this study that mycorrhizal dependency (MD) varied with saline treatments. Root colonization of *C. maxima* by *R. irregularis* showed great dependency of these plants on the AMF when compared to the purely saline non-mycorrhizal treatment. This corroborates the work of Beltrano et al.,³⁵ who demonstrated the favourable relationship between pepper and *G. intaradices* (*R. irregularis*), and shows that when roots were associated with AM fungi, the detrimental effect of the salinity stress decreased significantly. Taken together, although salinity reduced mycorrhizal colonization, the dependency of pepper plants on mycorrhizal fungi was high.³⁵⁻⁴¹

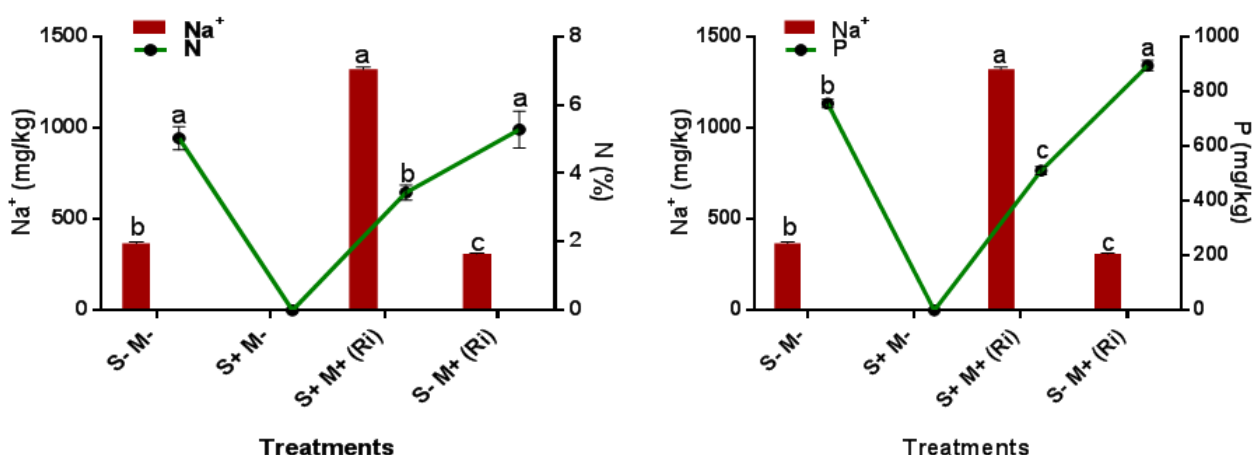


Figure 3 Influence of AMF inoculation on foliar Na⁺ uptake in *C. maxima* as it affects its N and P content.

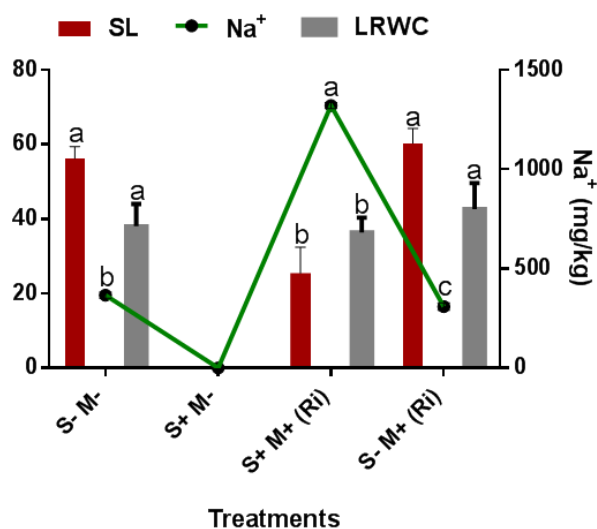


Figure 4 Influence of AMF inoculation on foliar Na⁺ uptake in *C. maxima* as it affects its Shoot length (SL) and Leaf relative water content (LRWC) content.

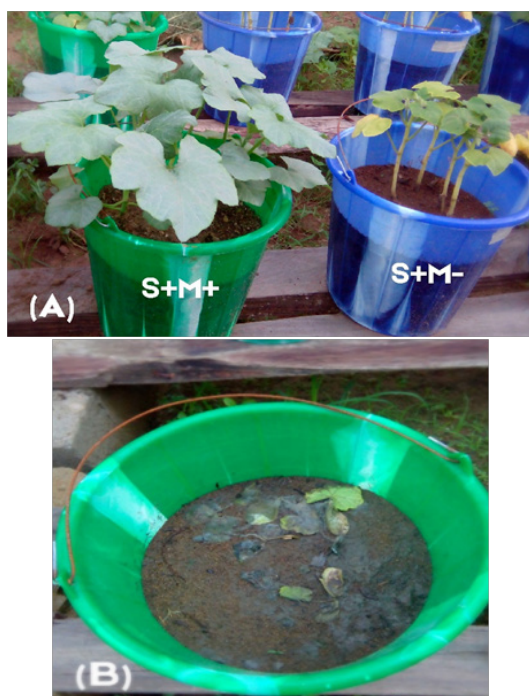


Figure 5 (A) Shows growth response of *Rhizophagus irregularis* inoculated saline treated (S+M+) and uninoculated saline treated (S+M-) *C. maxima* six (6) weeks after planting (B) uninoculated saline treated (S+M-) *C. maxima* dies off twelve (12) weeks after planting; soil shows hard surface crusting which results in poor soil aeration, porosity and low microbial activities.

Conclusion

The accumulation of salts in the soil is one of the major threats facing crop yield and production worldwide. It has been estimated that about 7% of the earth's land is exposed to high soil salinity levels. Results of this research revealed that salt stress negatively affected the physicochemical properties of the saline soil when compared to the garden soil, thus resulting in excessive uptake of Na⁺ and subsequently; negative effects on photosynthetic pigments,

minerals contents, relative leaf water content (RLWC), shoot length, dry weight, mycorrhizal colonization and dependency of *C. maxima*. The effects of mycorrhizal symbiotic association on *C. maxima* showed improvements on the growth and physiology of *C. maxima*. Using different mechanisms *C. maxima* by itself or in association with arbuscular mycorrhizal fungi (*R. irregularis*) can tolerate or survive soil salinity. However, symbiotic association of the fungus *R. irregularis* with roots of *C. maxima* gives the plant the ability to resist the stress as a result of morphological and physiological vicissitudes and improved vigour/water content, adjusted rate of K⁺/Na⁺, extensive network of the mycorrhizal plant roots and enhanced nutrient uptake are all among the processes that made the mycorrhizal inoculated plant to survive under severe salt stress conditions.

Acknowledgments

None.

Conflicts of interest

The author declares there is no conflicts of interest.

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