

Research Article





Quality assessment of waterside river, ogbor hill, aba 2: effect of three-point samples on some functional parameters of protein metabolism of wistar rats

Abstract

This study assessed total solids, TS, total dissolved solids, TDS and total suspended solids, TSS, of three-point samples of waterside River Ogbor hill Aba (WRO) and the effect on rats serum protein metabolism parameters. The TS, DS, TDS and TSS (mg/L solids) respectively, of the downstream-sample (300.08±0.01, 16.25±0.35 and 60.03±12.62) was higher (p<0.05) than that for the upstream and midstream. Aside TS and TSS, the TDS of the samples were lower (p<0.05) than the corresponding EPA standard though the TDS value of the midstream sample was the least (p<0.05). The protein metabolism parameters as determined by total protein, albumin, total bilirubin and direct bilirubin concentration (mg/100ml) revealed lower corresponding value compared to the control. As compared among the groups, the midstream sample elicited the least total protein but highest direct bilirubin concentration while the upstream sample had the highest albumin concentration whereas the downstream sample had the highest total bilirubin concentration. The observation was not marked but statistically significant (p<0.05). Thus, the apparently clear WRO samples significantly altered the protein metabolism of the rats with possible adverse effects, warranting follow up and interim intervention measures, including provision of alternative, perhaps treated, water source for the residents.

Keywords: albumin, total dissolved solids, total suspended solids, total protein, total bilirubin, direct bilirubin

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Introduction

Generally, water is a universal, including physiological, solvent. Thus, water quality is critical to overall animal, including human, physiology. Potential health hazards usually result from animal, including aquatic, exposure to poor quality water¹ hence the need for quality assessment of water sources especially such that are utilized by inhabitants for their overall water needs, including for drinking. The need for quality assessment of water sources is heightened in the face of evident activities that could contaminate and pollute hence compromise the quality of such water sources as waterside River, Ogbor hill Aba in Aba North Local Government Area, Abia State, eastern Nigeria. Water source is usually polluted by prevailing environmental factors and human activities. The Environmental Protection Agency, EPA² listed the major causes of water pollution to include discharges from municipal wastewater or silage and effluent discharges from farms and factories. Either banks of waterside River Ogbor hill Aba traverse the rear of many factories, including breweries (soft drinks and beer) as well as soaps and detergents factories. Other prevailing activities that could predispose WRO to contamination and pollution include that as from abattoirs (hide and skin burning), auto mechanic workshops as well as human wastes (faeces, urine) and general refuse disposal.

Raji et al.,³ concurred that rivers are contaminated by the activities of the residents and industries and that high levels of pollutants in river water systems (as may be the case with WRO) could increase the total dissolved solids (TDS), total suspended solids (TSS) which are among the basic indicators of water quality status.⁴ In particular, little seems to have been done on to study and document the quality status *vis a vis* highlighting the possible health implications following animal exposure to WRO. These warranted this study aimed at determining the total solids, TS, total dissolved solids, TDS and total suspended solids, TSS) of three-point samples of waterside

River Ogbor hill Aba (WRO) and assessing the effect of the water samples on rats serum protein metabolism parameters. Protein metabolism parameters as studied herein are known bioindicators of health status hence useful in assessing physiological functions and/or dysfunctions in animals, including humans. Products of protein metabolism (total protein, albumin, total bilirubin and direct bilirubin concentration) are important biochemical indicators of physiological functions and dysfunctions.⁵⁻⁷ While TS, TDS, TSS do not have any health implication² the values nevertheless are important indicators of water quality status. For instance, high suspended solids can lower water quality by absorbing instead of reflecting heat thereby and increasing the water temperature and lowering the ability of the water to hold oxygen.⁸

Materials and methods

Sample collection

Three-point water samples were collected from the waterside River Ogbor hill Aba as indicated in the map thus: The first point, Upstream, the second point, Downstream and the third point, Midstream. The water samples were collected using a properly labeled plastic containers and used without any treatment shown in Figure 1.

Animal study

Animal procurement and exposure groups

Twenty adult male albino rats (*Rattus novergicus*) used in this study were procured from the animal house of the Faculty of Biological Sciences, University of Nigeria, Nsukka. The animals were acclimatized for 2 weeks and then randomized (based on weight) to four experimentation groups with sample size of five rats. Rats in the Group A (Control) were sham-exposed to 0.5 ml of clean sachet water while rats in Groups B, C and D were respectively exposed to





0.5 ml of waterside river sample collected Upstream, Midstream and Downstream. The exposure was *per* oral, using a gavage, and daily for 4 weeks (twenty-eight days). Rats in all the groups were allowed free access to water and rat chow (commercial growers mesh feeds (Vital Feeds, Jos Nigeria).

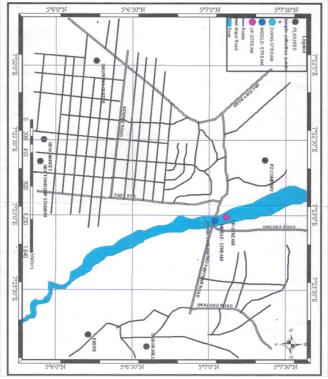


Figure I Map of the study area.

Sacrifice, blood sample collection and preparation

After 4 weeks (28 days) exposure, the rats were sacrificed the next day after overnight fast by cardiac puncture technique⁹ after cervical dislocation and the blood sample of the respective rats was collected individually into clean non-heparin tubes. The respective blood sample thus collected was allowed to clot and then centrifuged at 3000 g for 10 min. The resultant serum sample was separated and stored in a refrigerator at 0 °C and used the next day to determine the studied serum protein metabolism parameters *viz:* total protein, albumin, total bilirubin and direct bilirubin concentration.

Ethical consideration

This study considered and adhered to the standard ethical use of experimental animals. Throughout out the experimentation (acclimatization and exposure periods), all rats were housed at 25 °C in stainless steel cages under normal daylight/dark cycle and humid tropical conditions. The rats were allowed free access to rat feed (Vital Feeds, Jos Nigeria) and tap water, and generally received humane care in accordance with the guidelines of the National institute of Health, USA for ethical treatment of laboratory animals as approved by the various (departmental and college) ethical committees of Michael Okpara University of Agriculture Umudike, Nigeria.

Determination of serum protein parameters

The serum total protein concentration of the rats was determined by the Biuret method as described by Henry and Winkelman¹⁰ based on the principle that the carboxyl and the amino ends of peptide bonds of proteins react with cupric ions in moderately alkaline medium to form violet colour whose intensity is proportional to the concentration of protein present in the sample. Two and a half milliliters of Biuret reagent was respectively added to 0.05 ml of serum sample, protein standard and blank (distilled) in respective clean test tubes and incubated at room temperature for 20min. The respective absorbance thereafter was read using in a spectrophotometer at 540nm. The serum albumin concentration of the rats was determined with commercial diagnostic kit (Fortress, United Kingdom) according to the method that used Bromocresol Green (BCG) as described by Douman et al.11 This was based on the principle that serum albumin binds with Bromocresol green indicator in an acid medium to form a green Bromo Cresol compound that is directly proportional in concentration with the albumin concentration present in the sample. To 0.1ml each of distilled water or serum sample pipetteed into clean test bottles respectively labelled blank and sample respectively was added 3.0ml of bromocresol green reagent and allowed to stand for 5 min at 29 °C and the absorbance read with a spectrophotometer at 630nm. The serum total and conjugated/direct bilirubin concentration determination was by the colorimetric method as described by Jendrassik & Grof¹² based on the principle that conjugated (direct) bilirubin reacts with diazotized sulphanilic acid in an alkaline medium to form a blue coloured complex which is measured with a spectrophotometer and calculated to give the concentration of conjugated/direct bilirubin in the serum sample.

Total bilirubin is determined on the principle that in the presence of caffeine total or albumin-bound bilirubin are released on reacting with diazotized sulphanilic acid. However, conjugated/direct bilirubin is determined on the principle that in the presence of Nacl, conjugated/ direct bilirubin are released on reacting with diazotized sulphanilic acid forming a coloured compound with the intensity as measured using a spectrophotometer proportional to the concentration. To four labelled clean test tubes labelled serum sample 1, serum sample 2, distilled water blank 1 and distilled water blank 2, 0.2ml of 29 mmol/L sulphanilic acid was pipetted followed by 1.0 drop of 25mmol/L nitrate. Then, 1ml of caffeine was added to sample 1and blank 1 (for total serum bilirubin determination) while 2.0 ml 0.9% NaCl was added to sample 2 and blank 2 (for conjugated/direct serum bilirubin determination). A volume of 0.2ml of serum and 0.2 ml of distilled water was added to the sample and sample blank respectively. All the solutions were mixed and incubated at room temperature for 10 minutes. The absorbance of the conjugated bilirubin was read against the sample blank at a wavelength of 546nm in a spectrophotometer. One milliliter of 0.93mmo1/1 tartrate was deposited into the set up for total bilirubin and further incubated for 10min before the absorbance was read spectrophotometrically at a wavelength of 578nm.

Determination of total solids, total dissolved solids and total suspended solids

The total suspended solids (TSS) was determined by gravimetric method described in UNEP¹³ The respective sample aliquot was passed through a pre-ignited and pre-weighed Whatman filter. The Whatman filter containing the residue was placed in a porcelain dish, oven-dried at 105 °C for two hours thirty minutes, cooled for 15 minutes and dessicated using a desiccator to a constant weight. The total dissolved solid (TDS) of the samples was determined using a TDS meter (Model C0150, manufactured by Hach) according to the method described by UNEP¹³ The power key and the IDS key of the meter were switched on, and the meter was also temperature adjusted.

The probe dipped below the surface of the water samples. Time was allowed for the reading to be stabilized and reading was recorded. Total solids, residue after evaporation to dryness, was determined measuring the initial weight (W1) of an evaporating dish using a laboratory digital balance (Adams, USA) before heating on a hot air oven at 105°C for 1 hour and cooling in a desiccator. Then, 100 ml of water sample was added into the dish and boiled over a steam bath to dryness. The residue in the dish was further heated for one hour in an oven at 105°C, cooled and weighed to constant weight (W2). Total solid in the sample was calculated using the relation

Total solid
$$(mg/L \text{ solids}) = (W2 - W1) \times 100$$

To determine total suspended solid, Whatman filter paper was rinsed in distilled water, dried in an oven at 105°C for one hour, cooled in a desiccators and its initial weight (W1) determined using a weighing digital balance. Then, 100 ml of water sample was filtered through the paper and thereafter dried at 105°C for one hour. The final weight (W2) of filter paper containing the residue was recorded and the total suspended solids calculated using

Total suspended solids
$$(mg/L \text{ solids}) = (W2 - W1) \times 100$$

Statistical analysis

All numeric data collected were analyzed by one way analysis of variance (ANOVA) using the statistical package for Social Science (SPSS version 17; SPSS Inc., Chicago.IL.,USA). Results were presented as means±standard deviation (Mean±SD) at 95 % significance level (p<0.05).

Results and discussion

This study determined total solids, TS, total dissolved solids, TDS and total suspended solids, TSS of three-point samples of waterside River Ogbor hill Aba (WRO) and assessed the effect on rats serum protein metabolism parameters. The TS, TDS and TSS (mg/L solids) respectively, of the downstream-sample (300.08±0.01, 16.25±0.35 and 60.03±12.62) was higher (p<0.05) than that for the upstream and midstream. Aside TS of the three-point samples that was within the standard range provided by the Environmental Protection Agency, EPA¹⁴ and the TSS which was not stated, the TDS of the samples were lower (p<0.05) than the corresponding EPA standard value (Table 1). The results suggest that the samples may be clear with apparent acceptable quality. Also, the samples may have low inorganic salt content hence low electrical conductivity, low mineralization, low salinity and low corrosiveness.14 Generally, total suspended solids, TSS, solid materials, including organic and inorganic, plankton and industrial waste⁸ that are in the water, provides clue to water clarity and also water quality while high TDS indicated high total inorganic satls hence high salinity.14 High Concentration of TSS could lower water quality by absorbing heat and increasing the water temperature thereby lowering the ability of the water to hold oxygen necessary

for aquatic life. In particular, Total solid, TS, measures all suspended colloidal and dissolved solids in a water sample and high. Total solids, TS, residue after evaporation to dryness,15 indicates reduced water clarity and reduced amount of sunlight entering the water. The TS of the samples was lower than 380 mg/ $L_{\rm solids}$ for river water sample reported by Fajobi & Shittu¹⁶ and within the acceptable limit (20 -500 mg/L_{solids}) as recommended by EPA14 The results are surprising considering the enormity of ongoing activities in the area that could compromise the quality status of WRO. However, similar apparently acceptable quality status of the samples was noted in our earlier report and attributable to inherent self purification potential of flowing rivers.17 The protein metabolism parameters as determined by total protein, albumin, total bilirubin and direct bilirubin concentration (mg/100ml) revealed lower total protein concentration but higher albumin, total bilirubin and direct bilirubin concentration of the samples compared to the control. The midstream sample had the least total protein but highest direct bilirubin concentration compared to the control and the other samples. The upstream sample had the highest albumin concentration while the downstream sample had the highest total bilirubin concentration compared to the control and the other samples. The observation was not marked but statistically significant (p<0.05), Table 2 and Figure 2. This may be a significant indication of apparent adverse effect of the samples on the protein metabolism functional capacity of the rats. Bilirubin results from the enzymatic breakdown of hemoglobin and other haem-containing proteins including myoglobin and cytochromes⁵ and elevated serum bilirubin levels indicates impaired excretory capacity of the liver while lowered serum bilirubin indicates severe biliary obstruction.⁶ Albumin maintains osmotic pressure required for proper distribution of the body fluids among other physiologic functions and serum albumin concentration provides a clue on the synthetic capacity of liver, general liver dysfunction, onset of renal diseases and even malnutrition.^{6,7} Thus, higher serum albumin as noted in this study probably resulted from up-regulation in protein synthesis and then catabolism hence an indication of physiological dysfunction possibly following adverse influence of the water samples on the protein metabolism of the rats. The breakdown of proteins occurs mostly in the liver and to an extent in the intestine and kidney.¹⁸ Protein in excess of body need is not stored but broken down for excretion. However, in a normal or healthy state the apparently futile protein metabolism (simultaneous catabolism and anabolism) is less likely. Hence, the noted increase in total protein while indicative of increased protein synthesis19 could also be an indication of enhanced protein catabolism. The lower total protein concentration in the rats exposed to midstream sample compared to that in the rats exposed to upstream and downstream samples while indicating neither inhibited protein synthesis nor enhanced protein catabolism²⁰ could also be an indication of malnutrition in the rats. In a malnourished state, protein could serves as energy source resulting to its diminished concentration in the serum as noted in this study.

Table 1 TS,TSS and TDS compositions of three-point (Upstream, Midstream and Downstream) water samples from waterside River, Ogbor hill Aba, Abia State, Nigeria

| Parameters | EPA standard# | Upstream | Midstream | Downstream |
|--|---------------|-------------|-------------|-------------|
| Total Solid,TS (mg/L solids) | 20-500 | 200.14±0.01 | 200.07±0.01 | 300.08±0.01 |
| Total dissolved solid, TDS (mg/L solids) | 500 | 14.53±0.04 | 12.35±0.49 | 16.25±0.35 |
| Total suspended solid, TSS (mg/L solids) | NS | 40.04±0.01 | 40.10±0.14 | 60.03±12.62 |

NS=Not stated. Value presented as mean ± SD of triplicate determinations. Significant difference at p<0.05. #EPA²

Table 2 Effect of three-point (Upstream, Midstream and Downstream) water samples from waterside river, Ogbor hill Aba, Abia State on serum protein metabolism parameter of normal male albino rats

| Parameters | Control | Upstream | Midstream | Downstream |
|-----------------------------|-----------|-----------|-----------|------------|
| Total protein(mg/100ml) | 6.74±0.25 | 6.26±0.24 | 5.72±0.13 | 6.22±0.19 |
| Albumin(mg/100ml) | 2.07±0.25 | 2.75±0.11 | 2.16±0.02 | 2.20±0.02 |
| Direct bilirubin (mg/100ml) | 0.43±0.05 | 1.46±0.08 | 1.69±0.13 | 1.66±0.14 |

Value presented as mean ± SD of sample size, n=5 rats. Significant difference at p<0.05.

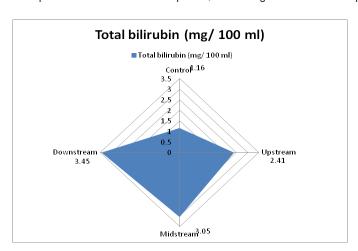


Figure 2 Effect of three-point (Upstream, Midstream and Downstream) water samples from waterside river, Ogbor hill Aba, Abia State on serum total bilirubin concentration (mg/100ml) of normal male albino rats.

Conclusion

Thus, the apparently clear WRO samples significantly altered the protein metabolism of the rats with possible adverse effects, warranting follow up and interim intervention measures, including provision of alternative, perhaps treated, water source for the residents.

Acknowledgements

None

Conflict of interest

The author declares there is no conflict of interest.

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