

An updated review of single herbal drugs in the management of osteoporosis

Abstract

Osteoporosis is a bone disease characterized by a decrease in bone mass and micro-architectural alterations. This would lead to a bone with less tensile strength and significantly more susceptibility to fracture with less force. This syndrome is clinically silent but progressive, usually only noted when a fracture occurs. Though there is no direct reference of this condition in classical texts of Ayurveda, it can be correlated with the *Astikshaya*, on the basis of pathophysiology and symptoms. For the management of this condition, various therapeutic measures are recommended in the classical texts. Among them, single herbal remedies for the management of osteoporosis are in routine clinical practice. The present review has been carried out to compile different pre-clinical and clinical research works reported on single herbal drugs for the management of osteoporosis. Analysis of results shows that, about 11 different drugs mentioned in classical texts of Ayurveda are reported for their anti-osteoporotic properties in different clinical and experimental studies. Clinically *Nigella sativa* is reported for its effect on the bone markers of postmenopausal women. Plants like *Withaniasomnifera*, *Cissusquadrangularis*, *Punicagranatum*, *Tinosporacordifolia*, are studied experimentally and found effective in the management of osteoporosis. The findings of present review highlight the use of these single and simple herbal remedies for the treatment of patients suffering from osteoporosis and can give a lead to further extensive research on these drugs.

Keywords: *astikshaya*, ayurveda, herbal drugs, medicinal plants, osteoporosis

Volume 11 Issue 2 - 2018

Shubhashree MN,¹ Raghavendra Naik,²
Doddamani SH,¹ Sulochana Bhat³

¹Research Officer (S-2), Regional Ayurveda Research Institute for Metabolic disorders, India

²Research Officer (Ay), Regional Ayurveda Research Institute for Metabolic disorders, India

³Research Officer (S-3), Regional Ayurveda Research Institute for Metabolic disorders, India

Correspondence: Shubhashree MN, Research Officer (S-2), Regional Ayurveda Research Institute for Metabolic disorders (RARIMD), Govt Central Pharmacy Annexe, Near Ashoka Pillar, Jayanagar, Bangalore, Karnataka, India, Tel 9448 0169 68, Email shubhathejas@gmail.com

Received: August 18, 2017 | **Published:** March 27, 2018

Introduction

Osteoporosis is the most common metabolic bone disorder characterized by decreased bone strength. It is a major problem of health care delivery services, both in the developed and developing countries. According to WHO, osteoporosis is a “progressive systemic skeletal disease characterized by low bone mass and micro architectural deterioration of bone tissue, with a consequent increase in bone fragility and susceptibility to fracture.”¹ This is a heterogeneous cluster of abnormal processes characterized by the net loss of bone. It results in a decrease in total mineralized bone without a decrease in the ratio of bone mineral to the organic matrix.^{2,3} As a result, there is a decrease in the overall amount of bone.

In Ayurvedic classical literature, there is no direct reference regarding osteoporosis and its management. Acharya Sushruta⁴ has explained about *Astikshaya* where the causative factors, signs and symptoms are similar to osteoporosis. Different Acharya like Charaka⁵ and Vagbhata⁶ have opined the same.

Prevalence of this disease is increasing as the population of elderly is on a rise. It is estimated that as many as 1 in 2 women and 1 in 5 men are at risk for an osteoporosis-related fracture during their lifetime.⁷ Because of increase in ageing population, it is estimated that by the year 2020, the number of women affected will be double.⁸ Thus, this condition has assumed major public health importance in recent years. It has been estimated that spinal compression fractures are associated with osteoporosis and 15 to 34% of all patients with hip fractures will die from complications within 6 months.⁹

Though different treatment modalities like HRT, Calcium & Vit-D supplement are commonly used, there is no treatment which has satisfactory improvement without side effects. Due to some adverse effects and lacunae of synthetic drugs, there is a need of finding out

better remedy for the management of osteoporosis. Therefore the present review has been made to compile the single herbal drugs reported for their beneficial effect on osteoporosis and to present them in a single place.

Material and methods

Different herbal drugs reported for their effect on management of osteoporosis were compiled from published research articles. The available data is arranged according to their Sanskrit name, English name, botanical identity and part used. The mechanism of action of reported drugs is analyzed critically and presented systematically.

Results and discussion

Analysis of data reveals that, about 11 different drugs mentioned in classical texts of Ayurveda are reported for their anti-osteoporotic properties in different clinical and experimental studies (Table 1). Clinically *Nigella sativa* is reported for its effect on the bone markers of postmenopausal women. Different parts of the plants like *Withaniasomnifera*, *Cissusquadrangularis*, *Punicagranatum*, *Tinosporacordifolia*, *Curcuma longa*, *Nigella sativa*, *Meliaazedarach*, *Asparagus racemosus*, *Moringaoleifera*, *Zingiberofficinale* and *Sesamumindicum* are studied experimentally and found effective in the management of osteoporosis.

Ashwagandha-withaniasomnifera (L.) wunal

Effect of *Withaniasomnifera* root ethanolic extract has been evaluated for anti-osteoporotic activity in ovariectomized Sprague-Dawley rats. Extract at the dose of 65 mg/kg for 16 weeks has shown a significant increase in serum alkaline phosphatase levels and urinary calcium and phosphorus excretion. Histological findings has revealed narrowed, and disappearance of trabeculae with widened medullary spaces in the ovariectomized group.¹⁰

Table 1 Single herbal drugs reported for their effect on osteoporosis

S No	Sanskrit name	English name	Botanical identity	Part used
	Ashwagandha	Winter cherry, Indian Ginseng	<i>Withaniasomnifera</i> (L.) Dunal (Solanaceae)	Root
	Asthishrinkhala	Edible stemmed wine	<i>Cissusquadrangularis</i> L. (Vitaceae)	Aerial parts
	Dadima	Pomegranate	<i>Punicagranatum</i> L. (Lythraceae)	Fruit
	Guduchi	Gulbel, Indian Tinospora	<i>Tinosporacordifolia</i> (Thunb.) Miers (Menispermaceae)	Stem
	Haridra	Turmeric	<i>Curcuma longa</i> L. (Zingiberaceae)	Rhizome
	Krishna jiraka	Black cumin	<i>Nigella sativa</i> L. (Ranunculaceae)	Seeds
	Nimba	Neem	<i>Meliaazedarach</i> L. (Meliaceae)	Bark
	Shatavari	Wild Asparagus, Sparrow grass	<i>Asparagus racemosus</i> Willd. (Asparagaceae)	Tuberous root
	Shigru	Moringa, drumstick tree	<i>Moringaoleifera</i> Lam. (Moringaceae)	Aerial parts
	Shunthi	Ginger	<i>Zingiberofficinale</i> Roscoe (Zingiberaceae)	Rhizome
	Tila	Sesam	<i>Sesamumindicum</i> L. (Pedaliaceae)	Seeds

Asthishrunkala– *cissusquadrangularis* L.

The phytoestrogen-rich fraction (IND–HE) from aerial parts of *C. quadrangularis* has shown presence of phytoestrogen-rich fraction. Treatment with IND–HE (75 and 100mg/kg) showed statistically significant increase in bone thickness, bone density and bone hardness in ovariectomised in rats. IND–HE and estrogen treatment significantly increased serum estradiol, serum vitamin D3 and serum calcium compared to control. Alkaline phosphatase was significantly reduced. Results of Histopathology studies indicated that IND–HE (75 and 100mg/kg) prevented bone loss.¹¹ In another experimental study, ethanol extract of *Cissusquadrangularis* at two different dose levels of 500 and 750mg/kg per day showed a definite antiosteoporotic effect.¹² The petroleum–ether extract at the dose of 500mg/kg, for 3 months reduced bone loss, as evidenced by the weight gain in femur, and also reduced the osteoclastic activity there by facilitating bone formation in experimental animals.¹³ Asthishrunkhala contains anabolic and phytoestrogenic steroids like Ketosteroids, sitosterol, alpha amayrin, alpha ampyrone and tetracyclic triterpenoids.¹⁴ These anabolic and steroidal components showed a marked influence on fracture–healing. Ketosteroid acts as antagonists to the glucocorticoid receptor and promotes good bone health. It mobilizes fibroblast and chondroblasts to an injured tissue and enhances regeneration. The anabolic steroidal component of Asthishrunkhala showed a marked influence in the rate of fracture healing by influencing early regeneration of all connective tissues of mesenchyme origin, namely the fibroblasts, the chondroblasts and osteoblasts involved in the healing and quicker mineralization of the callus.¹⁵

Dadima – *punicagranatum* L.

Anti–osteoporotic activity of ethanolic extract of *Punicagranatum* in ovariectomized rat model of osteoporosis at 100, 300 and 500 mg/kg is reported experimentally. There was significant increase in femur length, weight and density, increase in serum calcium, phosphorus and reduction in alkaline phosphatase, tartrate resistant acid phosphatase, osteocalcin whereas urine calcium, creatinine and phosphorous levels were significantly decreased. Histology of femur exhibits restorative progress with increased ossification, mineralization and increased osteoclastic activity.¹⁶ Polar fraction of *Punicagranatum* (L) peel extract at the doses of 50, 100, and 200 mg/kg significantly prevented bone loss in ovariectomized rats. These effects were described in increased mineral content of calcium. On histology data shown that fraction could increase osteoblast number.¹⁷ Consumption of pomegranate peel extract was able to significantly prevent the decrease in bone mineral density and bone micro–architecture impairment in ovariectomized mice.¹⁸ The alcoholic extract of fruit peel at the dose of 500mg/kg and 750mg/kg, daily for 90 days showed significant increase in uterine

weight, femur BMD and femur hardness. In addition, increased levels of calcium and phosphorus in serum and significant decrease in urine were observed.¹⁹ Exposure of different concentrations (10–100µg/ml) of the ethanolic extract on osteoblastic cells showed characteristic morphological changes and increment in cell number. A significant growth in cell proliferation, ALP activity, collagen contents and matrix mineralization of osteoblasts in a dose dependent manner suggested that extract has a stimulatory effect on osteoblastic bone formation or potential activity against osteoporosis.²⁰

Guduchi–*tinoporacordifolia* (thunb.) miers

Effects of alcoholic extract of *Tinosporacordifolia* on the proliferation, differentiation and mineralization of bone like matrix was studied on human osteoblast–like cells MG–63 and primary osteoblast cells isolated from femur of rats. The extract at a dosage of 25µg/ml stimulated the growth of osteoblasts, increased the differentiation of cells into osteoblastic lineage and increased the mineralization of bone like matrix on both the osteoblast model systems used in the study. Cell morphology studies clearly indicated the increase in cell numbers and absence of adverse change in the cell morphology on treatment with the extract.²¹ Aqueous and alcoholic extracts were evaluated for osteogenic effect using a widely employed in vitro model system for human osteoblasts (human osteoblast like cells SAOS–2). It was observed that ethanolic extract stimulated proliferation of osteoblasts at a dosage of 25µg/ml but, the aqueous extract showed no influence on cell proliferation. The extract also elicited pro–stimulatory effects on osteoblasts.²² Probably with this insight, the fermented form of medication is recommended for therapeutic purposes in Ayurveda.

Haridr –*curcuma longa* L.

Curcumin is considered as a potential treatment in numerous diseases, including osteoporosis. Curcumin has been reported to affect osteoclastogenesis and osteoblast proliferation and activity in vitro.²³ Extracts prepared from *Curcuma longa* L., containing bioactive phenolic curcuminoids was evaluated for bone–protective effects in a hypogonadal rat model of postmenopausal osteoporosis. The curcuminoid–enriched turmeric prevented up to 50% of ovariectomized induced loss of trabecular bone and also preserved the number and connectedness of the strut–like trabeculae.²⁴ Treatment with curcumin was able to reverse all the ovariectomy–induced deteriorations. The high dose of curcumin treatment was not only able to reduce the osteoclast number but also increase the osteoblast count.²⁵ Curcumin administration ameliorates oxidative stress–induced apoptosis in osteoblasts by preserving mitochondrial functions and activation of Akt–GSK3β signalling. These data provide experimental evidence supporting the clinical use of curcumin for prevention or treatment of osteoporosis.²⁶

Krishna jeeraka –nigella sativa linn.

In an experimental study, ovariectomized rats showed significant decrease in plasma Ca²⁺, accompanied by a significant increase in plasma ALP, amino terminal collagen type 1 telopeptide, MDA, nitrates, TNF- α and IL-6. These changes were reversed by supplementation of test drug.²⁷ *Nigella sativa* seed oil improved the micro architecture and biomechanical properties of the femur in male diabetic rats to a level equivalent to that achieved with parathyroid hormone treatment.^{28,29} In a clinical study, effects of *Nigella sativa* supplements was evaluated on the bone markers of postmenopausal women. The test drug failed to cause any significant changes in the bone markers levels, when supplemented for the duration of 3 months to these postmenopausal women.³⁰ The sample size of the study was only 15 and the duration of study was not longer to obtain the readings of bone markers at several time points and any changes in the bone mineral density. So, a long term study with larger sample size may give more convincing results.

Nimba –meliaazadirech linn.

In an in vitro study, it has been reported that, the root extracts of *M. azedarach*, could be used as medicines for osteoporosis. The extract s inhibited osteoclast proliferation and induced apoptosis by up-regulation of caspase activity and increase of mitochondrial pro-apoptotic proteins expression. Furthermore, the extracts enhanced differentiation, but did not affect proliferation of both osteoblasts and chondrocytes. The osteo-inducible effect was also observed in cultured primary bone marrow cells.³¹

Shatavari– asparagus racemosus willd.

Methanolic and aqueous extract obtained from *Asparagus racemosus* root has shown significant effect on mineralization, ossification and osteoclastic activity suppression were observed in histopathological examination. Significant increase in total ash weight, ash percent and ash calcium content were obtained. The extract significantly reduced serum alkaline phosphatase activity, serum calcium and also inhibited the ovariectomized induced excessive loss of calcium in urine. It also improved biomechanical parameters including hardness of 4th lumbar vertebra, femoral length and its weight. Phytosterols and other active constituents present in the root of *Asparagus racemosus* may effect on estrogen receptor similar to estrogen and produce antiosteoporotic effect.³²

Shigru– moringaoleifera lam.

In an in-vivo study, methanolic extracts of *Moringaoleifera* components showed a positive effect on osteoblast cell line SaOS2. Flower and fruit were found to have significant osteoblast stimulating property. Flower extract was found to be increasing the number of osteoblastic cells; while the fruit extract was having more elaborative effect as it increased ALP activity, induced bone formation, increased hydroxyproline content and bone mineral formation.³³ Regenerative effect of *Moringaoleifera* extract on haematological parameters and bone marrow of adult Wistar rat is reported.³⁴ Ethanolic extract at the dose of 600 mg/kg, significantly reduced urinary calcium excretion and significantly increased calcium content of bones in ovariectomised rats. The osteoprotective effect was comparable with estradiol.³⁵ Ethanolic extract of leaves at the dose of 100, 200, and 300ng/ml enhanced osteogenic differentiation capacity of porcine bone marrow derived mesenchymal stem cells as demonstrated by increased alkaline phosphatase staining and alkaline phosphatase activity.³⁶

Shunti– zingiberofficinale roscoe

The osteo-protective effects of structurally-related polyphenols (gingerols) isolated from the rhizomes of *Z. officiale* was reported in experimental studies. All the extracts were found bone protective in streptococcal cell wall induced arthritis, preventing bone mineral density loss as determined by dual energy absorptiometry.³⁷ Treatment with 6-gingerol stimulated osteoblast differentiation in normal and inflammatory settings. This compound induced the differentiation of osteoblast like cells with increased transcription levels of osteogenic markers, upregulated ALP enzyme activity, and enhanced mineralized nodule formation. It also reduced the degree of inflammation in TNF- α -treated MG-63 cells.³⁸ Oil extract of garlic possibly has a positive role in suppressing ovariectomy induced bone resorption. Garlic oil extract supplementation prevented ovariectomy-induced significant alteration of serum alkaline phosphatase activity, serum tartrate resistant acid phosphatase activity, urinary excretion of calcium, phosphate, hydroxyproline and urinary calcium to creatinine ratio.³⁹

Tila– sesamumindicum linn.

In an experimental study, feeding of 10% sesame oil reduced the significantly altered alkaline phosphatase activity and tartrate resistant acid phosphatase activity in ovariectomized rats. The test drug also reduced disruptive, lytic bone trabeculae and improved bone microarchitecture.⁴⁰

Conclusion

Osteoporosis is the most common metabolic bone disorder characterized by reduced bone mass and osteoporotic fracture. This condition seriously hampers the quality life of individual and needs an effective treatment measure without any adverse effect. Different single herbal drugs mentioned in Ayurveda showed significant results in improvement of osteoporotic changes. All these drugs are easily available, simple for administration and devoid of any adverse reactions. Further clinical studies can be planned to establish their role in the effective management of osteoporosis in clinical practice.

Acknowledgments

Authors are thankful to Director General, CCRAS for his constant support and encouragement.

Conflicts of interest

The authors declared that there are no conflicts of interest.

References

1. World Health Organisation (WHO). Assessment of fracture risk and its application to screening for postmenopausal osteoporosis. World Health Organisation (WHO): Geneva; 1994.
2. Simon LS. Osteoporosis. *Clinics in Geriatric Medicine*. 2005;21(3):603–629.
3. Avioli LV, Lindsay R. The female osteoporotic syndrome(s). In: Avioli LV, Krane SM, editors. *Metabolic Bone Disease and Clinical Related Disorders*. Philadelphia, USA: WB Saunders; 1990.
4. Samhita S. The Nibandhasangraha commentary of Sri Dalhanacharya. In: Vaidya Yadavji, Trikrampi Acharya, editors. *Chaukhambha Sanskrit Sansthan*. 4th ed. Sutra Sthana 15/13. Varanasi: 1980.
5. Charaka Samhita revised by Charaka & Dridhbala, with Ayurveda Dipika commentary by Chakrapanidatta, edited by Yadavaji V, Acharya T, editors. *Chaukhambha Sanskrit Sansthan*. 5th ed. Sharira Sthana 3/7: Varanasi; 2001.

6. Astanga Hridayam with commentaries Sarvangasundara of Arunadatta and Ayurvedarasayana of Hemadri. In: Harishastri B, Vaidya P, editors. Sutra Sthana 11/29. Krishnadas Academy: Varanasi; 2000.
7. US Department of Health and Human Services. Bone Health and Osteoporosis: A Report of the Surgeon General. MD: US Department of Health and Human Services Office of the Surgeon General: USA; 2004.
8. Jayanand, Behari J, Lochan R. Effects of low level pulsed radio frequency fields on induced osteoporosis in rat bone. *Ind J Exp Biol.* 2003;41(6):581–586.
9. Leonore C, Huppert. Hormone Replacement Therapy: Benefits, Risks, Doses, The Postmenopausal Woman. *Modern Clin N Am.* 1987;71(1):23–36.
10. Nagareddy PR, Lakshmana M. Withania somnifera improves bone calcification in calcium-deficient ovariectomized rats. *J Pharm Pharmacol.* 2006;58(4):513–519.
11. Aswar UM, Mohan V, Bodhankar SL. Antiosteoporotic activity of phytoestrogen-rich fraction separated from ethanol extract of aerial parts of *Cissusquadrangularis* in ovariectomized rats. *Indian J Pharmacol.* 2012;44(3):345–350.
12. Shirwaikar A, Khan S, Malini S. Antiosteoporotic effect of ethanol extract of *Cissusquadrangularis* Linn. on ovariectomized rat. *J Ethnopharmacol.* 2003;89(2-3):245–250.
13. Potu BK, Rao MS, Nampurath GK, et al. Evidence-based assessment of antiosteoporotic activity of petroleum-ether extract of *Cissusquadrangularis* Linn. on ovariectomy-induced osteoporosis. *Ups J Med Sci.* 2009;114(3):140–148.
14. Singh MP, Hemadri P. Medicinal Herbs with their formulations. *Daya publishing house.* 2005;21:1–21.
15. Resnick D. Diagnosis of bone and joint disorders. 3rd ed, 68th chapter. Pennsylvania: WB Saunders Company; 1995. p. 2735–2738.
16. Halekunche Y, Burdipad G, Kuppusamy S, et al. Anti-osteoporotic activity of ethanol extract of leave on *Punica granatum* leaves on ovariectomized rats. *Asian Journal of Pharmacy and Pharmacology.* 2016;2(4):85–92.
17. Bahtiar A, Arifin S, Razalifha A, et al. Polar Fraction of *Punicagranatum* L. peel extract increased osteoblast number on ovariectomized rat bone. *International Journal of Herbal Medicine.* 2014;2(1):65–70.
18. Spilmont M, Léotoing L, Davicco MJ, et al. Pomegranate Peel Extract Prevents Bone Loss in a Preclinical Model of Osteoporosis and Stimulates Osteoblastic Differentiation in Vitro. *Nutrients.* 2015;7(11):9265–9284.
19. Satpathy S, Patra A, Purohit AP. Estrogenic activity of *Punicagranatum* L. peel extract. *Asian Pacific Journal of Reproduction.* 2(1):19–24.
20. Siddiqui S, Arshad M. Osteogenic potential of *punicagranatum* through matrix mineralization, cell cycle progression and runx2 gene expression in primary rat osteoblasts. *DARU.* 2014;22:72.
21. Abiramasundari G, Sumalatha KR, Sreepriya M. Effects of *Tinosporacordifolia* (Menispermaceae) on the proliferation, osteogenic differentiation and mineralization of osteoblast model systems *in vitro*. *J Ethnopharmacol.* 2012;41(1):474–480.
22. Abiramasundari G, Sreepriya M. Pro-Stimulatory Effects of *TinosporaCordifolia* (Menispermaceae) on SAOS-2 Osteoblast Cells – Implications on Bone Remodeling and Therapy of Osteoporosis. *Research Journal of Pharmaceutical, Biological and Chemical Sciences.* 2014;5(3):354.
23. Folwarczna J. Curcumin and Its Potential Effects on the Development of Postmenopausal Osteoporosis. In: Hollins Martin C, Watson R, Preedy V, editors. *Nutrition and Diet in Menopause.* Nutrition and Health. Humana Press: Totowa; 2013. p. 165–180.
24. Wright LE, Frye JB, Timmermann BN, et al. Protection of trabecular bone in ovariectomized rats by turmeric (*Curcuma longa* L.) is dependent on extract composition. *Journal of Agricultural and Food Chemistry.* 2010;58(17):9498–9504.
25. Hussan F, Ibraheem NG, Kamarudin TA, et al. Curcumin Protects against Ovariectomy-Induced Bone Changes in Rat Model. *Evid Based Complement Alternat Med.* 2012;2012:174916.
26. Dai P, Mao Y, Sun X, et al. Attenuation of Oxidative Stress-Induced Osteoblast Apoptosis by Curcumin is Associated with Preservation of Mitochondrial Functions and Increased Akt-GSK3 β Signaling. *Cell Physiol Biochem.* 2017;41(2):661–677.
27. Seif AA. Nigella Sativa reverses osteoporosis in ovariectomized rats. *BMC Complement Altern Med.* 2014;14(1):22.
28. Altan MF, Kanter M, Donmez S, et al. Combination therapy of Nigella sativa and human parathyroid hormone on bone mass, biomechanical behavior and structure in streptozotocin-induced diabetic rats. *Acta Histochem.* 2007;109(4):304–314.
29. Altan MF. Effects of Nigella sativa and human parathyroid hormone on bone mass and strength in diabetic rats. *Biol Trace Elem Res.* 116(3):321–328.
30. Shuid AN, Ping LL, Muhammad N, et al. The effects of Labisiapumila var. alata on bone markers and bone calcium in a rat model of postmenopausal osteoporosis. *J Ethnopharmacol.* 2011;133(2):538–542.
31. Mukudai Y, Kondo S, Koyama T, et al. Potential anti-osteoporotic effects of herbal extracts on osteoclasts, osteoblasts and chondrocytes *in vitro*. *BMC Complement Altern Med.* 2014;14:29.
32. Chitme HR, Muchandi IS, Burli SC, et al. Effect of Asparagus Racemosus Willd Root Extract on Ovariectomized Rats. *The Open Natural Products Journal.* 2009;2:16–23.
33. Patel C, Rangrez A, Parikh P. The anti-osteoporotic effect of Moringaoleifera on osteoblastic cells: SaOS 2. *IOSR Journal of Pharmacy and Biological Sciences.* 2013;5(2):10–17.
34. Owolabi JO, Opoola E, Caxton-Martins EA. Healing and Prophylactic Effects of Moringaoleifera Leaf Extract on Lead Induced Damage to Haematological and Bone Marrow Elements in Adult Wistar Rat Models. *Open Access Scientific Reports.* 2012;1(4):1–8.
35. Sanganna C, Burali SC. The beneficial effect of ethanolic extract of *Moringaoleifera* on osteoporosis. *International Journal of Pharmaceutical Applications.* 2010;1(1):50–58.
36. Marupanthorn K, Kedpanyapong W. The Effects of MoringaOleifera Lam. Leaves Extract on Osteogenic Differentiation of Porcine Bone Marrow Derived Mesenchymal Stem Cells. 4th International Conference on Advances in Agricultural, Biological & Ecological Sciences (AABES-16): UK; 2012. p. 1–4.
37. Janet L Funk, Fry JB, Wright LE, et al. Effects of Ginger (*Zingiber officinalis* L) on Inflammation-Induced Bone Loss. *The FASEB Journal.* 26(Supplement 1):263–265.
38. Fan JZ, Yang X, Bi ZG. The effects of 6-gingerol on proliferation, differentiation, and maturation of osteoblast-like MG-63 cells. *Braz J Med Biol Res.* 2015;48(7):637–643.
39. Mukherjee M, Das AS, Mitra S, et al. Prevention of bone loss by oil extract of garlic (*Allium sativum* Linn.) in an ovariectomized rat model of osteoporosis. *Phytother Res.* 2004;8(5):389–394.
40. Boulbaroud S, Mesfioui A, Arfaoui A, et al. Preventive effects of flaxseed and sesame oil on bone loss in ovariectomized rats. *Pak J Biol Sci.* 2008;11(13):1696–1701.