

Editorial





Photoluminescent gold nanomaterials as sensitive probes

Editorial

Photoluminescent gold nanomaterials have become interesting sensing materials because of their ease in preparation and conjugation, large Stokes shift, long lifetime, and biocompatibility. 1-3 They are often called gold nanodots (Au NDs) or gold nanoclusters (Ag NCs), with sizes usually smaller than 2nm; Au NCs are commonly reffered to the one having less than 30Au atoms per cluster. The molecularlike optical properties (photoluminescence) of Au NDs and NCs are highly dependent on the size of Au core and the number of Au atoms per templates, respectively. 4,5 The Au complexe on the surface of Au core and its surface density affect the stability and optical properties of Au NDs, while the nature and size of the template affect that of Au NCs. Thus, the optical properties of AuNDs/NCs can be easily tuned by selecting suitable templates or ligands and controlling their molar concentration ratio to Au3+. Their emission wavelengths are usually longer than 600nm when excited with a UV light (commonly at 365nm). With a support of having a large Stoke shift and long lifetime, it is believed that their photoluminescen is mainly through the ligand-metal charge transfer transition (LMCT) or ligand-tometal-metal charge transfer transition (LMMCT).

Au NDs can be prepared from etching of small Au nanoparticles (<3 nm in diameter) by thiol compounds such as 11-mercaptoundecanoic acid (11-MUA) under alkaline conditions. 6 On the other hand, proteins such as bovine serum albumin (BSA), lysozyme, and horseradish peroxidase as a template and reducing agent are commonly used to prepare stable Au NCs.7 The reducing ability of these proteins mainly comes from their tyrosine residues that can reduce Au³⁺ at pH values>10.0. Although Au NCs can also be prepared using thiol compounds such as glutathione as a reducing agent, they are usually less stable and have weaker photoluminescence intensity than that of BSA-Au NCs. Protein-Au NCs are more stable than 11-MUA -Au NDs against photo irradiation and salt induced quenching, mainly because large protein molecule protect Au core more efficiently, which minimizes the access of quenchers such as oxygen to the core. While, large amounts (usually>several mM) of expensive proteins are required to prepare stable and bright Au NCs.

Au NDs and NCs have been employed for sensing of various analytes such as metal ions, anions and proteins, as well as for cell imaging. 11-MUA-Au NDs were used for quantitation of Hg²+ using 2,6-pyridinedicarboxylic acid (PDCA) as a masking agent to prevent the interference from Pb²+ and Cd²+, with limit of detection (LOD) 5 nM.6 In the presence of Hg²+, photoluminescence quenching of 11-MUA-Au NDs occurs, due to the Au-Hg aurophilic interaction and aggragation of the Au NDs through the interaction of Hg²+ ions with the carboxylate groups present on the surfaces of the 11-MUA-Au NDs. To minimize salt quenching, poly(N-isopropylacrylamide) microgels (PNIPAM MGs) containing 11-MUA-Au NDs were prepared and used for quantitation of Hg²+ in the presence of 500mM NaCl, with LOD of 1.7 nM.8 Because H₂O₂ can oxidize the Au core of 11-MUA-Au NDs, leading to photoluminescence quenching, the Au NDs is selective for quantitation of glucose when using glucose oxidase to catalyze the

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Jinshun Cang, Huan-Tsung Chang^{2,3}

Department of Chemical Engineering, Yancheng Institute of Industry Technology, China

²Department of Chemistry, National Taiwan University, Taiwan ³Department of Chemistry, Chung Yuan Christian University, Taiwan

Correspondence: Huan Tsung Chang, Department of Chemistry, National Taiwan University, Taipei, 10617, Taiwan, Email changht@ntu.edu.tw

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reaction of glucose and oxygen. ⁹ 11-MUA-Au NDs conjugated with platelet-derived growth factors (PDGF) were used for quantitation of PDGF receptor, with great selectivity and sensitivity (LOD of 0.25nM). ¹⁰ Au NDs prepared from Au nanoparticles using different etching agents have been used for quantiation of various analytes, including S²⁻, NO₂-, concanavalin A (*Con A*), Phospholipase C, and so on. ^{11–14} Based on analyte-induced PL quenching, lysozyme-Au NCs were used for quantitation of Hg²⁺, with LODs of 10nM. ¹⁵

Through core-etching induced PL quenching, BSA-Au NCs were used for the detection of cyanide with a LOD of 200nM.¹⁶ BSA-Au NCs were used for determination of the activity of trypsin, with LOD of 86pM.¹⁷ Through digestion of BSA by trypsin, the BSA shell was destroyed, leading to PL quenching as a result of oxidation of Au NCs by oxygen. To specifically target cancer cells, BSA-Au NCs can be conjugated with recognition elements such as small ligands and antibodies. For example, folic acid-BSA-Au NCs are specific toward cancer cells over-expressed folate receptors. 18 A ratiometric probe using BSA-Au/Ce NCs was developed for monitoring the pH values of HeLa cells. 19 The BSA-Au/Ce NCs possess dual emission bands at 410 and 650nm when excited at 325 nm, which separately correspond to the BSA-Ce complexes and Au NCs. The PL at 410 nm is pHdependent while that at 650 nm is pH-insensitive. Because the red fluorescence of BSA-Au/Ce NCs is quenched by H₂O₂, the NCs with glucose oxidase were used for quantitation of glucose.²

Although biocompatible Au NCs and NDs have shown their potential for sensing of various analytes and for cell imaging, their quantum yields are generally lower than that of quantum dots²¹ and carbon dots.²² To provide specifiity toward tumors, strategies for improved stability and affinity through functionaliztion of Au NDs and NCs with biopolymers, synthetic poymers, small ligands are still highly demanded. It is also interesting to prepare theranostic nanomaterials (therapeutic drugs and diagnostic agents into one single platform) using other reporters and drugs to conjugate AuNCs/NDs.²³ With advances in nanotechnology, it is our belief that more stable and brighter Au NCs/NDs will soon be realized. They may have great potential for cell tracking and for single molecule detection.



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Conflict of interest

The author declares no conflict of interest.

References

- Chen LY, Wang CW, Yuan Z, et al. Fluorescent gold nanoclusters: recent advances in sensing and imaging. Anal Chem. 2015;87(1):216–229.
- Chen PC, Periasamy AP, Harroun SG, et al. Photoluminescence sensing systems based on copper, gold and silver nanomaterials. *Coord Chem Rev.* 2016;320:129–138.
- Ravindranath R, Prathik R, Chang HT. Synthesis, optical properties, and sensing applications of gold nanodots. *Chem Rec.* 2016;16(3):1664– 1675
- Forward JM, Bohmann D, Fackler JP, et al. Luminescence studies of gold(I) thiolate complexes. *Inorg Chem.* 1995;34(25):6330–6336.
- Luo Z, Yuan X, Yu Y, et al. From aggregation-induced emission of Au(I)-thiolate complexes to ultrabright Au(0)@Au(I)-Thiolate Core-Shell Nanoclusters. J Am Chem Soc. 2012;134(40):16662–16670.
- Huang CC, Yang Z, Lee KH, et al. Synthesis of highly fluorescent gold nanoparticles for sensing mercury(II). Angew Chem Int Ed. 2007;46(36):6824–6828.
- Kawasaki H, Hamaguchi K, Osaka I, et al. pH-dependent synthesis of pepsin-mediated gold nanoclusters with blue green and red fluorescent emission. Adv Funct Mater. 2011;21(18):3508–3515.
- Chen LY, Ou CM, Chen WY, et al. Synthesis of photoluminescent Au ND-PNIPAM hybrid microgel for the detection of Hg²⁺. ACS Appl Mater Interfaces. 2013;5(10):4383–4388.
- Shiang YC, Huang CC, Chang HT. Gold nanodot-based luminescent sensor for the detection of hydrogen peroxide and glucose. *Chem Commun.* 2009;23:3437–3439.
- Huang CC, Chiang CK, Lin ZH, et al. Bioconjugated gold nanodots and nanoparticles for protein assays based on photoluminescence quenching. *Anal Chem.* 2008;80(5):1497–1504.

- Yuan Z, Peng M, Shi L, et al. Disassembly mediated fluorescence recovery of gold nanodots for selective sulfide sensing. *Nanoscale*. 2013;5(11):4683–4686.
- Chen WY, Huang CC, Chen LY, et al. Self-assembly of hybridized ligands on gold nanodots: tunable photoluminescence and sensing of nitrite. Nanoscale. 2014;(19):11078–11083.
- Huang CC, Chen CT, Shiang YC, et al. Synthesis of fluorescent carbohydrate-protected au nanodots for detection of concanavalin A and Escherichia coli. Anal Chem. 2009;81(3):875–882.
- Chen WY, Chen LY, Ou CM, et al. Synthesis of fluorescent gold nanodotliposome hybrids for detection of phospholipase C and its inhibitor. *Anal Chem.* 2013;85(18):8834–8840.
- Wei H, Wang Z, Yang L, et al. Lysozyme-stabilized gold fluorescent cluster: synthesis and application as Hg²⁺ sensor. *Analyst*. 2010;135(6):1406–1410.
- Liu Y, Ai K, Cheng X, et al. Gold-nanocluster-based fluorescent sensors for highly sensitive and selective detection of cyanide in water. Adv Funct Mater. 2010;20(6):951–965.
- Hu L, Han S, Parveen S, et al. Highly sensitive fluorescent detection of trypsin based on BSA-stabilized gold nanoclusters. *Biosens Bioelectron*. 2012;32(1):297–299.
- Wang YL, Chen JJ, Irudayaraj J. Nuclear targeting dynamics of gold nanoclusters for enhanced therapy of HER²⁺ breast cancer. ACS Nano. 2011;5(12):9718–9725.
- Chen YN, Chen PC, Wang CW, et al. One-Pot synthesis of fluorescent BSA-Ce/Au nanoclusters as ratiometric pH probes. *Chem Commun* (Camb). 2014;50(62):8571–8574.
- Wang CW, Chen YN, Wu BY, et al. Sensitive detection of cyanide using bovine serum albumin-stabilized cerium/gold nanoclusters. *Anal Bioanal Chem.* 2016;408(1):287–294.
- Nolf KD, Cosseddu SM, Jasieniak JJ, et al. Binding and packing in twocomponent colloidal quantum dot ligand shells: Linear versus branched carboxylates. J Am Chem Soc. 2017;139(9):3456–3464
- Roy P, Chen PC, Periasamy AP, et al. Photoluminescent carbon nanodots: Synthesis, physicochemical properties and analytical applications. *Materials Today*. 2015;18(8):447–458.
- Chen WY, Chang HY, Lu JK, et al. Self-assembly of antimicrobial peptides on gold nanodots: against multidrug-resistant bacteria and wound healing application. Adv Funct Mater. 2015;25(46):7189–7199.