

Research Article





Estimation of phytochemical, antioxidant screening profile and thrombolytic activities of methanolic extract of *Antidesma bunius* L. leaf

Abstract

The crude methanolic extract of Antidesma bunius L. leaves belongs to the family of Euphorbiaceae. The purpose of this study was to evaluate total antioxidant capacity, free radical scavenging activity, thrombolytic activity and screening of phytochemical constituents. Total antioxidant capacity and free radical scavenging activity were determined by standard methods. In thrombolytic test clot lysis method was used. Phytochemical screening was done by characteristic color changes or color precipitate using standard phytochemical reaction methods. Results of the preliminary phytochemical screening of the methanolic extract of A. bunius L. leaves revealed the presence of various bioactive components which include saponins, steroids, alkaloids, flavonoids and proteins in the extract. The total antioxidant capacity of crude methanolic extract was also found very good compared to standard catechin. In DPPH assay the scavenging activity of the methanolic extract (IC₅₀ 3.10μg/ml) showed very good activity compared to standard BHT (IC₅₀ 2.30μg/ ml). The methanolic extract of leaves of A. bunius exhibited thrombolytic activity 21.1 %, thrombolytic activity respectively compared to standard streptokinase (61.31%) lysis of clot. Observing the in-vitro studies, it can be concluded that the methanolic extract of A. bunius leaves could be used in drug formulation or pharmaceutical application because of its effective pharmacological properties. Hence, the plant may further be explored for its various pharmacological activities.

Keywords: phytochemical, antioxidant, *antidesma bunius* leaves, methanolic extract, thrombolytic activity

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Introduction

Native flora has remedial potency for different diseases and it has been stated by traditional herbal medicinal practitioners.1 Medicinal plants was considered as one of the major medicament used in traditional system for various medical purposes. People also give priorities to natural products and that's why natural medicine was widely used than allopathic medicine. Furthermore natural medicine has minimum side effects.² A. bunius L. (Family-Euphorbiaceae) is traditionally used as sudorific and also in the treatment of snakebite. Additionally, it is used to increase perspiration in febrile condition and juice of this plant is beneficiary in the treatment of insomnia. Fresh juice of the fruits is used to manufacture wine as an antioxidant. Roots and leaves has antihelminthic property and used in indigestion cough and stomachache. The seeds can be used against round worms and threadworms, coughs, flatulence, intestinal colic and it can acts as pesticide.3 The function of antioxidant in human body is to give protection against free radicals by scavenging them.4 Free radicals are responsible for desolation of protein as well as lipid and DNA by oxidative process during aerobic metabolism in the biological system⁵ which lead to many degenerative diseases and chronic diseases along with atherosclerosis, carcinogenesis, diabetes mellitus, ischemic cardiac disease, immunocompromised and neurodegenerative diseases.6 Thromboembolus malady plays a major role behind morbidity and mortality in Bangladesh.⁷ Nowadays

cardiovascular disease which is caused by blood clot formation is considered as one of the life-threatening disease and number of people affected by it is increasing day by day.⁸ Consequently, thrombosis is the process of clotting blood inside the blood vessel results in blocking the flow of blood through the circulatory system.⁹ By using thrombolysis medications it is possible to dissolve clots formed in blood vessels.^{10,11} Streptokinase Alteplase, Anistreplase, Urokinase and Tissue Plasminogen Activator (TPA) are some of the commonly used thrombolytic drugs all over the world for the treatment of thrombosis^{12–14} but those drugs has adverse effects like haemorrhage,¹⁵ anaphylactic reaction and lack of specificity.¹⁶ So, scientists are still trying to develop the medicine from various plants and animal sources which must have thrombolytic,^{17–19} antiplatelet,^{20,21} antithrombotic²² and anticoagulant activity^{23,24} along with less side effects.

Plant description

A. bunius tree is shrubby, generally height in between 10 to 26 ft. (3-8 m) but it can extend up to 50 or even 100 ft. (15-30 m). Leaves are shaped like oblong which is 4 to 9 inch (10-22.5 cm) long, 2 to 3 inch (5-7.5 cm) wide. The tree is tiny, odorous, reddish and axillary racemes are 3 to 8 inch (7.5-20cm) long. Fruits of this plant are shaped like round or ovoid and taste acidic, slightly sweet. For some people it taste like bitter, straw-colored stone, an irregular, flattened oval, ridged, very hard, 3/8 in (1 cm) long, 1/4 in (6 mm) wide.³





Methods and materials

Preparation of extract

Fresh leaves of *Antidesma bunius* were collected from Chittagong hills in the month of August, 2017 and the plant was identified by an expert taxonomist. Based on the identification we submitted a voucher specimen to the national herbarium which was located in Mirpur, Dhaka, Bangladesh. After that an accession number: DACB 43490 was given. Firstly, 400gm dried and powdered materials were soaked in 1.7 liter of methanol in an amber glass container for about 14 days at room temperature with occasional shaking. The solution was primarily filtered by using cotton and followed by Whitman's filter paper number 1 after 14 days. Finally, we collected the filtrates which was concentrated to solid mass by using a rotary evaporator. ^{25,26}

Phytochemical analysis of methanolic crude extract

Experiment for the detection of tannin

We used the following method to find out the presence of tannin.²⁷ At first, we took the powdered sample (1gm) and then boiled the sample with 50 ml of filtered water. Then ferric chloride test is done by using the collected filtrate. In a test tube of 3ml filtrated solution, few drops of ferric chloride was added. We observed a greenish black precipitate which indicates the presence of tannin.

Experiment for the detection of flavonoid

By using the below²⁸ method, we determined the presence of flavonoid in the sample. To carry out the method we took about 10 ml ethyl acetate and mixed with 0.2g of the powdered sample. Finally the solution was heated for 5 minute in a hot water bath. When the mixture were cooled, we collected the filtrates which was used for the test.

Experiment for the detection of terpenoids

In this terpenoid test, 2ml of chloroform ($CH_3CO)_2O$ is mixed with 2ml of the extract. After that, the solution was evaporated to dryness and followed by 2-3 drops of concentrated sulphuric acid (H_2SO_4) into the solution and finally the solution was heated for about 2 min.²⁹ Observation: Deep red coloration which indicates the presence of terpenoids.

Experiment for the detection of saponin

The presence of saponin was determined by Foam and emulsion test as described by.³⁰ In 100 mL beaker, 0.25 gm of the powdered sample is mixed with 20 ml of water and the solution was boiled and finally collected filtrates were used for the test.

Experiment for the detection of steroids (salkowski test)

In this steroid test, 2ml of acetic anhydride is mixed with 2ml of the extract followed by boiling and coloring. Then 2ml of conc. sulphuric acid was added gradually in the test tube.³⁰ Observation: Reddish brown ring was formed in the intersection of two solutions which indicates the presence of steroid.

Experiment for the detection of phlobatannins (precipitate experiment)

In this phlobatannin test, the 2ml of the extract was added to 2ml of HCl (1%) and the extract was heated.³⁰ Observation: Red precipitate was taken as an indication of presence of phlobatannins.

Experiment for the detection of carbohydrates (molisch's test)

During performing carbohydrate test, In a test tube 2ml of the extract was added with 10ml of H_2O , treated with 2 drops of ethanolic anaphthol (20%) solution and concentrated sulphuric acid was added.³⁰ Observation: Reddish violet ring was formed in the intersection of two solutions results in determining the existence of carbohydrate in the sample.

Experiment for the detection of coumarins

In this coumarin test, the 2ml of the extract was added to 3ml of NaOH (10%). 30,31 Observation: Yellow coloration formation indicates the existence of coumarins.

Experiment for the detection of alkaloids (hager's test)

In this alkaloid test, few drops of Hager's reagent (saturated picric acid solution) is mixed 2ml solution of the extract solution.^{30,31} Observation: Yellow color precipitate formed and it indicates the existence of alkaloids in the sample.

Experiment for the detection of (xanthoproteic test)

In this protein test, the 1ml of the extract solution and 1ml of H₂SO₄ (conc.). 2ml extract and treated with 2ml NH₄OH and 3ml benzene.³⁰ Observation: Red coloration which indicates the presence of proteins.

Experiment for the detection of anthraquinones (borntrager's test)

In this anthraquinone test, the 3ml of the extract was treated with 3ml of benzene and then 5ml aqueous NH₃ (10%) was added in a test tube.³⁰ Observation: Pink, violet or red coloration in aqueous layer indicated the presence of anthraquinones.

Experiment for the detection of anthocyanins

In this anthocyanin test, the 2ml of the extract was treated with 2ml of HCl (2N) and then added NH₃ in a test tube.³¹ Observation: The formation of pinkish red to bluish violet coloration which indicates the presence of anthocyanins.

Antioxidant evaluation

Total Antioxidant capacity determination

Antidesma bunius plant has antioxidant properties which was determined by the following method suggested by Prieto et al. 32 with slight modifications. Detection of antioxidants such as catechin in the plant is carried out by the phosphomolybdenum method which was performed by observing the reduction of Mo (VI) to Mo (V) done by antioxidant compound. At first, in a test tube 3 ml of reaction mixture containing 0.6 M sulphuric acid, 28 mM sodium phosphate and 1 % ammonium molybdate was added to 0.5ml of plant extract/ standard at different concentrations (7.825-250 µg/mL). To complete the reaction, test tubes was incubated at 95 °C for 10 minutes. By using a spectrophotometer, the absorbance of the solution was measured at 695 nm against a blank solution at room temperature. A reference sample, Catechin was used as standard in this test. 3ml of reaction mixture contained in a typical blank solution and appropriate volume (500µl) of the same solvent used for the samples/standard was incubated as like the rest of the samples solution under the same conditions. Increased total antioxidant capacity is observed due to increased absorbance of the reaction mixture. Standard/samples were used at six different concentrations ranges from (7.825 to 250µg/ml)

to perform each antioxidant assay. Repetition of the experiment was done three times at each concentration.

DPPH radical scavenging assay

Free radical scavenging activity of various compounds and medicinal plants was evaluated by widely used 1, 1-diphenyl-2picrylhydrazyl (DPPH) method according to.^{23,24} When it reacts with hydrogen donors, free radical of DPHH is reduced to the corresponding hydrazine results in making stable free radicals in both aqueous or methanol solution and produces purple/violet color in methanol solution and in the presence of antioxidants, it fades to shades of yellow color. Then, in the test tube 3 ml of methanol solution of DPPH was mixed with 2 ml of methanol solution of plant extract or standard sample at different concentration (3.906-150 µg/mL). To complete the reaction, the test tube was incubated at room temperature for 30 minutes in dark place. By using a spectrophotometer, the absorbance of the solution was measured at 517 nm against a blank solution. All reagents except plant extract or standard solution contained in a typical blank solution. Finally, we calculated the percentage (%) inhibition activity according to the following equation mentioned below:

% DPPH radical scavenging activity =
$$\{(A_0 - A1)/A_0\} \times 100$$

In the above equation, absorbance of the control is A_0 and absorbance of the extract/standard is A1. Then we plotted the % of inhibition against concentration. Finally IC $_{50}$ was calculated from the graph. Repetition of the experiment was done three times at each concentration.

Thrombolytic activity

By following the method,³⁵ with slight modifications In vitro clot lysis activity of the leaves of A. bunius was determined. At first, 5 ml of venous blood were drawn from healthy volunteers followed by distribution in ten different pre weighed sterile vials (1ml/vial) and for 45 minutes it was incubated at 37°C. After formation of the clot, the serum was fully removed without disturbing the clot and each vial having clot again weighed to determine the actual clot weight (clot weight=weight of clot containing vial - weight of vial alone). 100µl solution of the crude extract was added separately to each vial containing pre-weighed clot. As a positive control and as a negative non thrombolytic control 100µl of streptokinase (SK) used and then separately 100µl of distilled water were added to the control vials. Before observing for clot lysis, all the vials were incubated for 90 minutes at 37 °C and removal of fluid were done after incubation and to observe the difference in weight after clot disruption, vials were again weighed. Finally, calculated the weight difference which was taken before and resulted clot lysis was expressed as percentage of clot lysis as shown below:

% clot lysis = (Weight of the lysis clot / Weight of the before lysis) \times 100

Results and discussion

Phytochemical screening of the extract was done to determine the existence or absence of bioactive compounds. Tannins, saponins, steroids, phlobatannins, flavonoids, coumarins, alkaloids, proteins, emodins, and alkaloids were presence (Table1) whereas, anthraquinones, anthocyanins, carbohydrates and terpenoids were absent.

Table I Qualitative analysis of the phytochemical screening of the A. bunius L

Phytochemical tests	Crude methanol extract
Saponins	+
Tannins	+
Flavonoids	+
Terpenoids	-
Steroids	+
Phlobatannins	+
Carbohydrates	-
Coumarins	+
Alkaloids	+
Proteins	+
Emodins	+
Anthraquinones	-
Anthocyanins	-

Here, +, Present,-, Not present.

Total antioxidant capacity

Total antioxidant activity of the plant extracts and (+) catechin (standard) were depicted in Table 2 in Figure 1. As shown in Figure 1 considerable antioxidant activity was shown by the crude methanolic extract compared to (+)-catechin (Standard). The activity was less than that of catechin. The absorbance of crude methanol extract and (+)-catechin at the concentration 62.50µg/ml were 0.665 and 0.775 respectively. With the increased concentration of the extract the total antioxidant activity also increased. The absorbance of crude methanol extract and (+)-catechin at 250 µg/ml were 1.537 and 3.457 respectively.

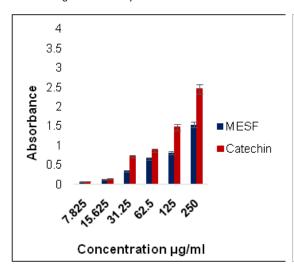
Table 2 Total antioxidant activity of the crude methanol extract of A. bunius and (+)-catechin (standard) at different concentrations

Cons (us/mil)	Absorb	ance	Absorbance		
Conc.(µg/ml)	a	a b		Mean ±STD	
7.825	0.072	0.073	0.075	0.073 ± 0.0012	
15.625	0.145	0.148	0.141	0.145 ± 0.0029	
31.25	0.723	0.720	0.718	0.720 ± 0.0020	
62.50	0.876	0.874	0.877	0.875 ± 0.0012	
125	1.461	1.463	1.469	1.469 ± 0.0033	
250	2.457	2.459	2.456	2.457 ± 0.0012	
	7.825 15.625 31.25 62.50	a 7.825 0.072 15.625 0.145 31.25 0.723 62.50 0.876 125 1.461	a b 7.825 0.072 0.073 15.625 0.145 0.148 31.25 0.723 0.720 62.50 0.876 0.874 125 1.461 1.463	a b c 7.825 0.072 0.073 0.075 15.625 0.145 0.148 0.141 31.25 0.723 0.720 0.718 62.50 0.876 0.874 0.877 125 1.461 1.463 1.469	

Table Continued....

Name of sample	Conc.(µg/ml)	Absorbance			Absorbance	
Name of Sample	Conc.(µg/mi)	a	b	c	Mean ±STD	
	7.825	0.061	0.065	0.068	0.064± 0.0028	
	15.625	0.133	0.136	0.131	0.133± 0.0020	
Crude methanol extract	31.25	0.334	0.336	0.339	0.336± 0.0020	
Crude methanol extract	62.5	0.663	0.665	0.668	0.665± 0.0020	
	125	0.812	0.815	0.818	0.815 ± 0.0024	
	250	1.506	1.504	1.601	1.537 ± 0.0452	

Each value is the average of three analyses±standard deviation



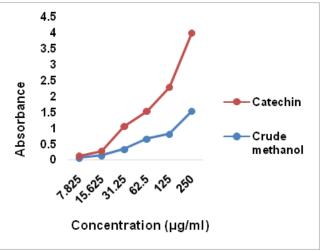


Figure I Total antioxidant capacity.

DPPH radical scavenging activity

By DPPH radical scavenging assay the DPPH radical scavenging activity of the extract of *A. bunius* was evaluated. The results of DPPH radical scavenging assays of plant extracts and BHT

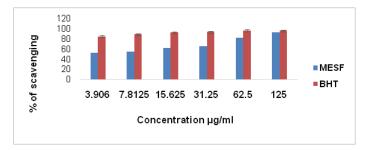
(butylatedhydroxytoluene) are given in Table 3 and in Figure 2.The scavenging activity shown by the crude methanol extract was very close to that of BHT (standard). IC $_{\rm 50}$ of standard and crude methanol extract were 2.30µg/ml and 3.75µg/ml respectively (Figure 3).

Table 3 DPPH radical scavenging activity of the crude methanol extract of A. bunius and BHT (Standard) at different concentrations

Name of sample	Conc. (µg/ml)	% of scavenging			% of scavenging	IC (=/==1	
		a	b	С	Mean ± STD	IC ₅₀ (µg/ml)	
	3.906	84.7	84.9	85.12	84.91±0.171		
	7.8125	88.74	89.32	88.72	88.93±0.278		
	15.626	93.02	91.89	92.78	92.56±0.486		
ВНТ		94.37	93.67	94.10	94.05± 0.288	2.30±0.002	
62	62.5	94.60	94.68	94.62	94.63±0.033		
	125	96.13	96.23	95.78	96.05±0.192		
Crude methanol extract	3.909	51.65	51.25	53.37	52.09±0.919		
	7.8125	54.06	54.98	55.01	54.68±0.440		
	15.625	60.6	62.12	61.55	61.42±0.626		
	31.25	64.67	66.42	65.33	65.47±0.883	3.75±0.079	
	62.5	80.96	81.77	83.66	82.13±1.131	3.73±0.077	
	125	92.82	91.88	91.68	92.12±0.497		

Each value is the average of three analyses±standard deviation.

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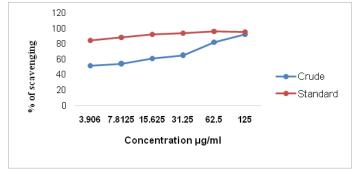


Figure 2 DPPH radical scavenging activity.

% of thrombolysis of different extract of leaf
Antidesma bunius

70 61.31

80 50

40 40

80 21.1

90 6.26

Bandle Santile

Figure 3 Thrombolytic activity of the extractives of the A. bunius leaves.

Thrombolytic activity

In present study, as a part of discovery cardio protective agents from natural sources, the extractives of *A. bunius* leaves were assessed for thrombolytic activity and the results are presented in the following Table 4.

Table 4 Thrombolytic activity (in terms of % of clot lysis) of the extractives of A. bunius leaves

Fractions	WImg	W2 (mg)	W3 (mg)	W4 (mg)	W5 (mg)	(W5/W4)X 100%
Sample	813	1160	1087	347	73	21.10%
Blank	815	1151	1130	336	21	6.26%
SK	835	1520	1100	685	420	61.31%

Discussion

The preliminary phytochemical screening tests may be beneficial in the detection of the bioactive components and may lead to the drug formulation and great importance in the field of drug research or drug discovery field. These tests facilitate their qualitative separation and quantitative estimation of pharmacologically active chemical compounds.³⁶ The results of the phytochemical screening of methanolic extract of A. bunius leaves has revealed the presence of saponins, phenols, tannins, flavonoids, alkaloids which could be responsible for the versatile medicinal properties or pharmacological actions of this plant part, like anti-diabetic, antioxidant, antiradical, cytotoxicity activity and pesticide agent.^{37,44} The antioxidant capacity of the methanolic extract was assessed by phosphomolybdenum method. This study confirmed that the methanolic extract showed a good total antioxidant activity that increasing concentration (Table 2). The antioxidant activity of the plant extract based on free radical scavenging is measured by DPPH test. This test measured the hydrogen donating ability of the plant extract to reduce DPPH to DPPH-H. In the DPPH assay, the conversion of purple color of DPPH solution to yellow to different grade depending on the existence and amount of antioxidant compounds within the extract. Our present study inspected that the methanolic extract of A. bunius had a similar free radical scavenging activity when compared with standard BHT (Table 3). In this study, the methanolic extract of leaves of A. bunius exhibited thrombolytic activity 21.1% compared to standard streptokinase 61.31%.

Conclusion

From this study, it was concluded that the phytochemical

screening of the methanolic extract of the leaves of *A. bunius* contain some bioactive components and these pharmacological properties are attributed to their phytochemical constituents. So, further studies are in progress to isolate the active constituents responsible for the observed effect.

Acknowledgments

None.

Conflicts of interest

Author declares that there is no conflicts of interest

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