

Alterations in transcript levels of hypothalamic endosomal recycling genes in high-fat diet-fed mice

Abstract

It has been well established that body weight gain observed in the high-fat diet (HFD) murine models is associated with inflammation-induced disturbances in protein degradation, leading to neuronal damage. We hypothesized that HFD dysregulates the protein recycling machinery in the mice hypothalamus. Therefore, we determined transcript levels of endosomal sorting proteins in HFD C57BL/6J fed mice. A qPCR-array of retromer-wash and cargo protein genes was performed in the arcuate nucleus (ARC) isolated from mice that had received high-fat or standard diets (StD). We found that transcript levels of *Ube2o*, *Tbc1d5*, *Igfr2*, and *Cdm6pr* were negatively associated with weight gain and hyperglycemia. We concluded that HFD-induced weight gain altered transcript levels of endosome retrieval system genes, suggesting that it could be dysregulating the trafficking of steady-state surface receptors in the hypothalamus.

Keywords: obese mice, high-fat diet, hypothalamus, retromer-wash complex, satiety

Volume 6 Issue 6 - 2018

Fernando Victor Martins Rubatino, Luana Assis Ferreira, Mariana Lacerda de Freitas, Cristina Moraes Junta, Celio Jose de Castro Junior, Adriana Aparecida Bosco, Fernanda Sarquis Jehee, Karla Simone Fernandes
Instituto de Ensino e Pesquisa, Santa Casa de Misericórdia de Belo Horizonte, Brazil

Correspondence: Karla Simone Fernandes, Instituto de Ensino e Pesquisa, Santa Casa de Belo Horizonte, P.O. Box: 30250-140, Belo Horizonte, Brazil, Tel +55 31 981047557, ++55-31 3238 8398, Fax +55 31 981047557, ++55-31-3238-8398, Email fernandaskss@yahoo.com.br

Received: October 23, 2018 | **Published:** November 23, 2018

Introduction

The central nervous system (CNS) plays an important role in food intake and energy expenditure regulation.¹ In the CNS, the ARC is suggested to be a key hub for energy balance, reducing food intake and improving energy expenditure through leptin and insulin interactions with their receptors, ObR and IR, respectively (Coll et al., 2007). An important step for physiological ligand-mediated cell signaling is the maintenance of a steady state of functional receptors at the plasma membrane, which involves dynamic intracellular transport mechanisms.² Leptin and insulin receptors require internalization and retrograde routes upon the binding of their ligands,³ then sorted and transported by vesicles, for direct recycling to the plasma membrane or via the Trans-Golgi Network (TGN) or lysosomal degradation.⁴ Retromer, a protein trimer, is essential for the recruitment of nucleation-promoting factors to endosomes, including the WASH complex, which has a carboxy-terminal VCA motif that binds to actin, allowing subsequent actin filament nucleation by the Arp2/3 complex.^{5,6} On the other hand, non-recycled cargoes are directed to lysosomal degradation pathways. Newly synthesized hydrolases acquire a mannose-6-phosphate marker during their maturation and are then transported by the mannose-6-phosphate receptors (CDM6PR, CIM6PR also named IGFR2) in a step for lysosomal formation.^{7,8} In this study, we evaluated the transcript levels of genes involved in endosomal recycling in a murine model of HFD induced obesity and showed that HFD fed mice presented altered transcript levels for *Ube2o*, *Igfr2*, *Cdm6pr* and *Tbc1d5*. And showed significant body weight gain and hyperglycemia.

Material and methods

Experimental murine model

All animal procedures were approved by the ethics committee at IEPSCBH, 0001-15. StD (3.7kcal/g, 10% from saturated fat) and HFD (5.3 kcal/g, 60% from saturated fat), were purchased from PragSoluções Biociências, Brazil. Male, 4-week-old C57BL/6J^{Unib} had housing occupation up to four animals per cage. Temperature (22 ± 2°C) and light cycle (12-h light/dark) were controlled and animals had food and water *ad libitum*. Animals were randomized into two

groups of six animals: one received 8 weeks of the HFD and the other receiving the StD for 8 weeks.

Transcript level analyses

Brains were removed from StD (n=6) and HFD (n=6) mice and immediately immersed in liquid nitrogen to preserve RNA. Hypothalamus were stored in RNAlater solution. RNA extraction was performed using the RNeasy Tissue Microarray (Qiagen®, Hilden, Germany), following the manufacturer's protocol. We performed transcript quantifications using a quantitative RT-PCR (qRT-PCR) assay with a two-step reaction process. cDNA synthesis was performed using the RT2 First Strand Kit (Qiagen®, Hilden, Germany). qPCR for the target genes was performed using a mouse customized RT2 Profiler PCR Array platform (Qiagen®, Hilden, Germany). Gene transcripts were determined according to Bustin et al.⁹

Statistical analysis

IBM SPSS Statistics 20 and R-3.3.0 software were used. A normal distribution was assumed for continuous variables followed by the Kolmogorov-Smirnov test. All data are represented as means ± standard deviations from a representative experiment of at least three independent experiments. Significance was accepted at p<0.05. For univariate analysis Student *t*-test was used. Bivariate comparisons were evaluated using the Pearson coefficient determination. For multivariate analysis, a Euclidean distance matrix was created. Ward's method for hierarchical clustering was applied and the number of clusters was obtained considering the D index values.¹⁰

Results

High-fat diet (HFD) induces weight gain and hyperglycemia in mice.

Mice receiving the HFD up to the 16th week were significantly heavier (42.3±4.3g; 45,8%) than mice that received the StD (29±1.9g; p<0.001; Figure 1A). They also presented significant hyperglycemia (192.3±27.3 mg/dL), when compared with StD mice (45.4±14.4 mg/dL; p<0.001; Figure 1B).

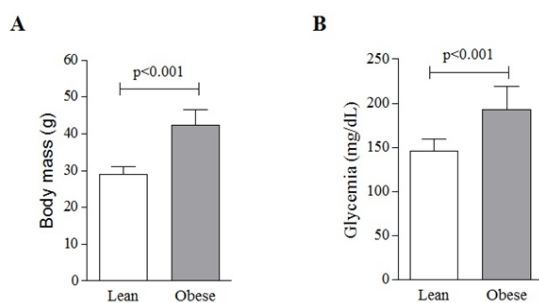


Figure 1 High-fat diet-induced obesity in mice. Twelve-week-old C57BL/6_{unib} male mice were randomly assigned to standard (St) (n=6) or high-fat diet (n=6). Body mass (A) and plasma glycemia (B) were determined on the 8th week. Values are means \pm SD. t Student test was applied to compare means between groups. A representative of three independent assays. 3.2. Transcript levels of endosomal protein sorting machinery are altered in HFD obese mice.

Mage-L2, *Trim27*, *Wash*, *Fam21*, *Vps35*, *Kiaa1033* and *Arcp2* genes presented unaltered transcript levels after HFD feeding ($p>0.05$; data not shown). Decreased transcript levels of *Ube2o* (62.3%; $p=0.027$), *Igfr2* (64.1%; $p=0.026$), *Cdm6pr* (54.8%; $p=0.002$) and *Tbc1d5* (67.1%; $p=0.04$) were observed in the hypothalamus of HFD mice, when compared to StD mice (Figure 2A). Total body weight negatively correlated to *Ube2o* ($p=0.027$), *Igfr2* ($p=0.038$), *Cdm6pr* ($p=0.003$) and *Tbc1d5* ($p=0.03$) levels, according to Pearson's coefficient. Plasma glucose levels also negatively correlated to *Ube2o* ($p=0.047$), *Igfr2* ($p=0.026$), *Cdm6pr* ($p=0.008$) and *Vps35* ($p=0.01$) levels.

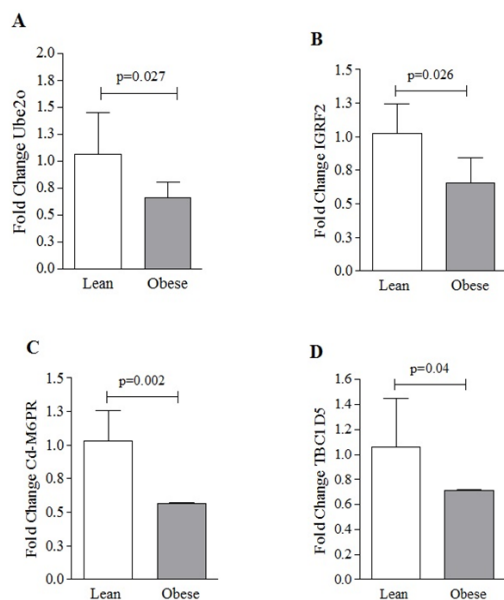


Figure 2 Determination of *Ube2o*, *Igfr2*, *Cdm6pr* and *Tbc1d5* mRNA levels in the arcuate nucleus by qPCR. Twelve-week-old C57BL/6_{unib} male mice were randomly assigned to a standard (St) (n=6) or high-fat diet (n=6). Gene expression data were determined by qPCR on the 8th week and represented as fold change relative quantitation. Values were expressed in means \pm SD. Comparisons between groups were evaluated by one-way analysis of variance, followed by the Student T-test. $p<0.05$ was considered significant. For both groups, n=6.

High-fat diet determines phenotypes by cluster definition based on *Ube2o*, *Tbc1d5*, *Igfr2* and *Cdm6pr* transcript levels in the hypothalamus

We performed a multivariate analysis to determine whether the HFD was able to induce alterations in gene expression profiles in the hypothalamus. This approach provides a graphical visualization of intragroup homogeneity and intergroup heterogeneity¹⁰ and this clustering method defined three distinct phenotypes for the *Ube2o*, *Tbc1d5*, *Igfr2* and *Cdm6pr* transcript levels, considering the D index (Figure 3A).

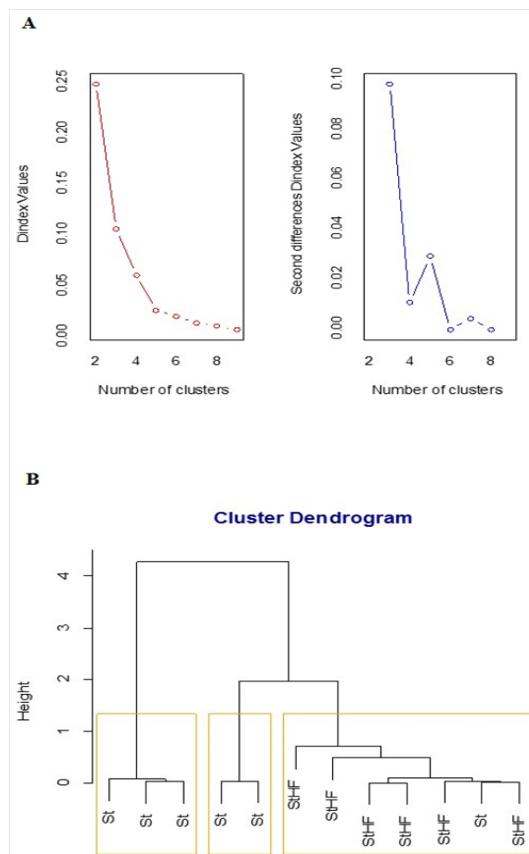


Figure 3 Hierarchical Clustering for St and StHF feed mice by Ward's method. Twelve-week-old C57BL/6_{unib} male mice were randomly assigned to a standard (St) (n=6) or high-fat diet (StHF; n=6). Fold change values were determined by qPCR on the 8th week. Euclidean distance was used as metric with *UBE2O*, *IGFR2*, *CDM6PR* and *TBC1D5* fold change values as dimensions (clustering dendrogram). The number of clusters was obtained by D Index.

Discussion

This is the first study reporting retromer-wash complex regulation in HFD fed mice. Our main finding is that body weight gain induced by HFD alters transcript levels of genes involved in the endosome-to-Golgi retrieval pathway. More specifically, transcript levels of the cargo proteins, *Cdm6pr* and *Igfr2*, Rab-GAP *Tbc1d5* and *Ube2o* E2-conjugase, were significantly decreased in HFD obese mice. The endosome-to-Golgi retrieval pathway is suggested to form part of the recycling machinery of a variety of proteins, from early to late endosomes and back to TGN, the plasma membrane and lysosomal degradation, and is essential for diverse cellular functions.^{5,11,12} Indeed, we specifically targeted some genes involved in endosomal protein sorting. They have been also associated with neuronal functions¹³

and neurodegenerative disorders.^{11,12} Quantitative PCR analyses showed that *Ube2o*, *Igfr2* and *Tbc1d5* Rab-GAP are decreased in the hypothalamus of obese mice and are inversely correlated with glycemia. *Ube2o* and *Mage-L2-Trim27* siRNA knockdown has been shown to impair retromer complex activation,⁵ where both *Mage-L2-Trim27* and *Ube2o* are essential for *Igfr2* (cargo) endosome-to-Golgi retrograde transport. We found no alteration in hypothalamic *Mage-L2* and *Trim27* levels in HFD fed mice, and no differences in WASH components.

Our findings concerning altered *Tbc1d5* levels suggest a dysfunction in retromer complex recruitment. It is known that retromer recruitment for the sorting of *Igfr2* to lysosomes is mediated by Rab7 and inhibited by the GTPase effect of *Tbc1d5*.¹⁴ In addition, the disruption of one step in retrograde transport is thought to be sufficient to impair all recycling of cargo proteins.⁵

It is well established that obesity induced by high-fat diet is accompanied by insulin and leptin resistance in mice^{15–19}. The limitations of our study were the lack of leptin and insulin resistance measurements or fat mass and food intake indexes. We chose body weight gain as a parameter for associating with endosome trafficking gene expression and found increases in mean body total weight gain and hyperglycemia that were consistent with those of previous reports.^{15,19,20} Thus, although our study did not investigate ObR expression in the hypothalamus of obese mice, reduced expression of *Igfr2* in HFD fed mice may suggest some alteration in ObR retrograde transport to TGN. ObR is reported to be endocytosed in a ligand-independent manner and colocalizes mainly with IGFR2 in the TGN compartment.¹ Furthermore, ObR is postulated to be retained intracellularly, suffering subsequent lysosomal degradation, which could maintain ObR surface expression at low levels in an in vitro and controlled milieu.²¹ The dynamics of ObR activity have also been elucidated in diet-induced obesity (DIO) mice, where both ObR cell surface density and STAT3 signaling activity were diminished in HFD mice.^{21,22,23} In further regard to degradation pathways, we detected diminished levels of *Cdm6pr* in HFD fed mice, which suggests an impairment of lysosomal degrading pathway.²⁴ These findings suggest a disruption of the lysosomal degradation pathway of proteins in HFD obese mice. Moreover, it has been reported that DIO mice present defects in the ubiquitin-proteasome system,²⁰ as HFD obese mice presenting inflammation-induced protein aggregates in the hypothalamus were also unable to regulate body energy homeostasis.

Hierarchical-cluster analyses have been used as a multivariate approach for behavior and phenotype group recognition. It is important to highlight that in this study, different groups were assumed after D index analysis, thus being recognized as non-arbitrarily clustered groups. Therefore, we found three heterogeneous phenotypes of transcript expression in the hypothalamus for all mice analyzed. Regarding animal distribution among the three groups, the StD fed mice group was represented by at least one individual in the three distinct phenotype clusters, while all HFD fed mice were clustered into just one phenotype cluster. This finding may suggest a dynamic variation of transcript levels in a normal physiological process, as observed for the StD fed group, which could be ablated during the weight gain process in HFD fed mice.

Conclusion

In conclusion, our findings suggest that protein-sorting machinery may be altered in HFD fed C57BL/6j mice and may be associated with weight gain and hyperglycemia.

Acknowledgments

The study was supported by FAPEMIG grant number APQ-01996-14.

Conflict of interest

The authors declare that they have no conflict of interest.

Author contributions

All authors contributed to the preparation of the manuscript. FVMR, KSF, FSJ and AAB accomplished experimental design and literature analysis. FVMR, LAF, MLF, CMJ, CJCJ and KSF performed experimental assays and data analysis. FVMR, KSF, CJCJ and AAB wrote the manuscript.

References

1. Kwon O, Kim KW, Kim MS. Leptin signalling pathways in hypothalamic neurons. *Cell Mol Life Sci.* 2016;73(7):1457–1477.
2. Belouard S, Delcroix D, Rouillé Y. Low levels of expression of leptin receptor at the cell surface result from constitutive endocytosis and intracellular retention in the biosynthetic pathway. *J Biol Chem.* 2004;279(27):28499–28508.
3. Fruhbeck G. Intracellular signaling pathways activated by leptin. *Biochem J.* 2006;393(Pt 1):7–20.
4. Seaman MN. Endosome protein sorting: motifs and machinery. *Cell Mol Life Sci.* 2008;65(18):2842–2858.
5. Hao YH, Doyle JM, Ramanathan S, et al. Regulation of WASH-dependent actin polymerization and protein trafficking by ubiquitination. *Cell.* 2013;152(5):1051–1064.
6. Seaman MN, Gautreau A, Billadeau DD. Retromer-mediated endosomal protein sorting: All WASHed up! *Trends Cell Biol.* 2013;23:522–528.
7. Hille-Rehfeld A. Mannose 6-phosphate receptors in sorting and transport of lysosomal enzymes. *Biochim Biophys Acta.* 1995;1241(2):177–194.
8. Gary-Bobo M, Nirdé P, Jeanjean A, et al. Mannose 6-phosphate receptor targeting and its applications in human diseases. *Curr Med Chem.* 2009;14(28):2945–2953.
9. Bustin SA, Beaulieu JF, Huggett J, et al. MIQE precis: Practical implementation of minimum standard guidelines for fluorescence-based quantitative real-time PCR experiments. *BMC Mol Biol.* 2010;11:74.
10. Aparecida MS. Análise de Dados Através de Métodos de Estatística Multivariada. 2013.
11. Seaman MNJ, Freeman CL. Analysis of the Retromer complex-WASH complex interaction illuminates new avenues to explore in Parkinson disease. *Commun Integr Biol.* 2014;7:e29483.
12. Fecto F, Esengul YT, Siddique T. Protein recycling pathways in neurodegenerative diseases. *Alzheimers Res Ther.* 2014;6(2):13.
13. Yi JJ, Ehlers MD. Emerging roles for ubiquitin and protein degradation in neuronal function. *Pharmacol Rev.* 2007;59(1):14–39.
14. Seaman MN, Harbour ME, Tattersall D, et al. Membrane recruitment of the cargo-selective retromer subcomplex is catalysed by the small GTPase Rab7 and inhibited by the Rab-GAP TBC1D5. *J Cell Sci.* 2009;122(Pt 14):2371–2382.
15. Hariri N, Thibault L. High-fat diet-induced obesity in animal models. *Nutr Res Rev.* 2010;23(2):270–299.
16. Coll AP, Farooqi IS, O'Rahilly S. The Hormonal Control of Food Intake. *Cell.* 2007;129(2):251–262.

17. Gautreau A, Oguievetskaia K, Ungermann C. Function and regulation of the endosomal fusion and fission machineries. *Cold Spring Harb Perspect Biol.* 2014;6(3):1–16.
18. Münzberg H, Flier JS, Bjorbaek C. Region-specific leptin resistance within the hypothalamus of diet-induced obese mice. *Endocrinology.* 2014;145(11):4880–4889.
19. Guo J, Jou W, Gavrilova O, et al. Hall Persistent diet-induced obesity in male C57BL/6 mice resulting from temporary obesigenic diets. *PLoS One.* 2009;4(4):e5370.
20. Ignacio-Souza LM, Bombassaro B, Pascoal LB, et al. Defective regulation of the ubiquitin/proteasome system in the hypothalamus of obese male mice. *Endocrinology.* 2014;155(8):2831–2844.
21. Uotani S, Bjørbaek C, Tornøe J, et al. Functional properties of leptin receptor isoforms: Internalization and degradation of leptin and ligand-induced receptor downregulation. *Diabetes.* 1999;48:279–286.
22. Seron K, Couturier C, Belouzard S, et al. Endospanins regulate a postinternalization step of the leptin receptor endocytic pathway. *J Biol Chem.* 2011;286(20):17968–17981.
23. Vauthier V, Roujeau C, Chen P, Endospanin, et al. Endospanin1 affects oppositely body weight regulation and glucose homeostasis by differentially regulating central leptin signaling. *Mol Metab.* 2017;6(1):159–172.
24. Kornfeld S. Structure and function of the mannose 6-phosphate/insulinlike growth factor II receptors. *Annu Rev Biochem.* 1992;61: 307–330.