

Review Article





Fusarium head blight and crown rot on wheat & barley: losses and health risks

Abstract

Wheat and Barley is the most production and consumption grains in the world. The necrotrophic Fusarium spp is pathogen caused many diseases on plants, the major two disease caused by Fusarium on wheat is Fusarium Crown rot (FCR) and head blight (FHB), also known as scab. These both disease caused severe damage on yield quality and quantity, the produced seed seem to be small, shriveled pale white appearance with low contain of protein, that reflected on reduce yield up to 61%. Mycotoxins contamination in the grain, will lead to contamination the products that manufactured from these crops, as well as the contamination yield will be lower price than the healthy yield, which leads to a significant loss to the producer. The major FCR and FHB mycotoxins as DON, NIV, ZEN can caused many disease symptoms to consumers who consumes contaminate food, whether is human or animal, such as cancers, immunosuppression, fertilization problem, abortion and etc.

Keywords: triticum spp, hordeum vulgare, fusarium, mycotoxins

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Abbreviations: FCR, *fusarium* crown rot; FHB, *fusarium* head blight; DON, deoxynivalenol; ZON, zearalenone; AcDON, monoacetyl–deoxynivalenols (3–AcDON, 15–AcDON); BEA, beauvericin; DAS, diacetoxyscirpenol; ENS, enniatins; FUS, fusarenone–X (4–Acetyl–NIV); HT2, HT–2 toxin; MON, moniliformin; NEO, neosolaniol; NIV, nivalenol; T2, T–2 toxin; T2ol, T–2 tetraol; ZEN, zearalenone

Introduction

Wheat and barley is the major crops cereal produce in the word. The global statistics indicate that wheat production in the world for 2014–2015 was 707.2million ton and for barley 135.65million ton, which form the proportion of one–third of the world's total grain production.^{1,2}

Fusarium crown rot (FCR)

Crown rot (FCR) caused by many kinds of *Fusarium* spp as *F. pseudograminearum* and *F. culmorum* is a common pathogen to FCR while *F. grarnineurm* group I, *F. crookwellense*, *F. avenaceum* and *F. nivale*.³⁻⁶ FCR disease infect the stem base of wheat and barley causing necrosis and dry rot of the crown bases always brown, often extending up 2–4nodes, basal stem and root tissue commonly known as crown rot.⁷ Whitehead formation is most severe in seasons with a wet start and dry climates, pinkish fungal growth may form on lower nodes especially during moist weather (Figure 1).

Fusarium Head Blight (Scab) (FHB)

FHB is caused by mainly fungus as *F. graminearum* group II (also known as *Gibberella zeae* sexual stage), *F. culmorum*, *F. poae*, *F. avenaceum* and *F. culmorum*. ^{8,9} Other species, can caused the disease such as *F. langsethiae*, *F. poae*, *F. sporotrichioides*, and *Microdochium nivale* (formerly known as *F. nivale*). ^{10,11} Disease symptoms are confined to the head, grain, and sometimes the peduncle (neck). Typically, the first noticeable symptom is bleaching of some or all of the spikelets while healthy heads are still green. As the fungus moves into the rachis, spikelets located above or below the initial infection

point may also become bleached. In barley the symptom is little different, infection spikelest is located and don't move in to rachis of spike. During humid and wet condition, pink to orange masses of spores may be visible on infected spikelets (Figure 2). 12-14 Wet and warm weather during crop growing and maturation may favour to FHB, 15 and hot and dry conditions increase CR disease. 16

Disease cycle

The causal agent for FCR and FHB is interplay. Infection occurs when spores land on susceptible wheat and barley heads. Heads are most susceptible at early flowering and infection may occur up to the soft dough stage, although severity is greatly reduced. If the flowers are infected just after their emergence kernels will not develop. Florets that are infected later will produce tombstones. Kernels that are infected by the pathogen during late kernel development may appear healthy, but be contaminated with mycotoxins. *Fusarium* crown and root rot can develop in the following season if infected seed is planted. Also maize is one of the crop that infected by same *Fusarium* spp that infected wheat and barly, so cultivation wheat or barly near or after maize rotation will increase inoculation which leaded to hight disease incidence (Figure 3).¹⁷

Yield losses

There are many diseases accompanying with wheat and barley cultivation, mostly about 9% of yield is losses by diseases in Kansas State was estimated for the last 20 year, *Fusarium* CR and HB constitutes about 0.29% losses from the total yield. ¹⁸ *Fusarium* infection reduces quality and crop yield, both FHB and FCR become an endemic disease in wheat and barley grower's area in the world wide. ¹⁹ *F. pseudograminearum* is the predominant CR pathogen in the 11 million/ha wheat growing region in Australia [7] where, nearly \$80 million each year is lost from reduced grain yield and quality due to CR. ²⁰ Both *F. graminearum* and *F. pseudograminearum* improved by bioassays that can caused equally severe CR disease. ²¹





Figure 1 Different symptom and sing of Fusarium crown rot (FCR) on wheat.

- (A,B) Pink/red fungal mycelium and discoloration of stem bases, Photo by SIMMYT.
- (C) Whitehead symptom on spike right: non infected, left: infected (White head), Photo by Guihua Bai.

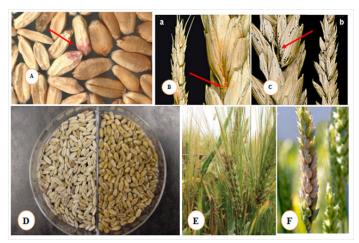


Figure 2 Head blight symptom on wheat and barley caused by Fusarium (FHB).

(A,D)small, shriveled pale white appearance and sometimes pink, infected (left)and healthy (right) Photographer by Department of Agriculture and Aquaculture Canada K. Lynch.

- (B,C) Orange sporodochia and Bluish black perithecia are formed at the base of the glumes, Photographer by Canadian Grain Commission.
- (E,F) Premature bleaching of Barley and wheat spikes.

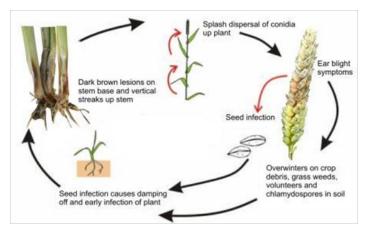


Figure 3 The typical life cycle of Fusarium in wheat Copyright: HGCA, UK.

In the Pacific Northwest of the United States, *F. pseudograminearum* can reduce winter wheat yield by up to 61%.²² While in during 1998–2000 FHB inflicted an estimated US \$ 2.7billion loss due to reduced yield and price discounts from lowered grain quality in the northern Great Plains and central USA.¹⁹ It has been reported that FCR could reduce wheat yield by up to 35% in the Pacific North– West of the United States.

Recent study found crown rot infestation causes an average of 25 percent yield loss in bread wheat and 58% loss in durum wheat across a wide range of environments in Australia, estimated annual yield loss of \$23million Australian dollars per year in barley and in wheat and barley combined in Australia. ^{20,23}

Since 1993, North Dakota, South Dakota, and Minnesota have lost 73% of their malting barley market with losses in Minnesota alone approaching 95%, In barley, losses have been equally devastating with estimated losses from 1993 to 1999 totaling in excess of \$400million. Yield losses in wheat since 1990 have exceeded 13Tg (500million bushels) with economic losses estimated at \$2.5billion.²⁴ FCR losses in yield arising on grain heads will be partially or half filled (Figure 2), or no grain in the head (preventing grain from forming in the heads of wheat) in severe infestation, as will reduction in grain quality and infected plants may also contaminated with mycotoxins.²⁵

FHB can caused significant yield losses from floret sterility and light weight kernel are produced as a result of infection, Quality reduction may occurrence if the pathogen produce mycotoxins in seed face^{26,27} (Table 1).

 $\begin{tabular}{ll} \textbf{Table I} Some statistical estimates for diseases losses on wheat yield, prepared by Kansas State University. 18 \\ \end{tabular}$

Disease name	%Percentage 4.33	
Rust disease		
Septoria complex	0.84	
Scab and crown rot	0.29	
Powdery medley	0.09	
Talk all	0.03	
Bunt & loose smut	0.02	

Mycotoxin contamination and risk

The mycotoxin risks connected with the consumption of contaminated forage, grain and straw by livestock must not be ignored. ^{28,29} High humidity and frequent rainfall increase *Fusarium* head blight (FHB) and crown rot disease in wheat and barley by dispersal and production spores from infected residue, the optimum temperatures for infection are between 75°F and 85°F. ³⁰ Mycotoxins are chemically and thermally stable contaminants which have toxic effects in animals and humans. ³¹ They can accumulation in the grain and, when contaminated grain is consumed via feed or food products, cause a potential risk to human and animal health. ³² Matny et al., ³³ found that most samples of animal feed that collected from market and animal farm in Iraq was contaminated with DON toxin at 722μg/kg.

Toxigenic strains of *F. culmorum* have been divided into two types the main type B trichothecenes produced that is DON and NIV chemotypes, strains of DON-types also produced AcDON (3-

AcDON),³⁴⁻³⁶ while strains of NIV-type were also able to produce FUS. In field trials, DON and NIV chemotypes exhibited different aggressiveness toward winter rye.³⁵ FHB in wheat is widespread disease caused losses up to 25–50%, and high amounts of DON and its derivatives were frequently found in freshly harvested infected grains. In particular, a very high occurrence of DON incidence of 100%, range 7.25–36.25mg kg⁻¹ was founding on wheat affected by FHB predominantly caused by *F. graminearum*, associated with consistent levels of 4,7–dideoxy–NIV (0.16–1.25mg kg⁻¹).³⁷ Most frequently encountered mycotoxins in field surveys in Europe infected wheat with FHB is deoxynivalenol (DON) and zearalenone (ZON) produced by *F. graminearum* and *F. culmorum*.^{29,38–40}

In Poland, samples collected from wheat naturally infected with *F. graminearum* and *F. culmorum* were founded to be contaminated with DON and 3AcDON 100% and 80% respectively at very high concentrations up to 30.4 and 29.54mg kg⁻¹ respectively, the levels of DON and 3AcDON in the chaff were 5–50times higher than in kernels⁴¹ (Table 2).

 $\textbf{Table 2} \ \text{Some statistical estimates for diseases losses on wheat yield, prepared by Kansas State University. I 8 \\$

S. No	Fungus Species	Mycotoxins
1	F. graminearum	DON, NIV, ZEN, AcDON
2	F. avenaceum	MON, ENS, BEA
3	F. culmorum	DON, ZOH,NIV, ZEN
4	F. poae	NIV, FUS, BEA, DAS
5	F. equiseti	DAS, ZOH, ZEN
6	F. tricinctum	MON
7	F. cerealis	NIV, ZOH, FUS, ZEN
8	F. sporotrichioides	T2, NEO, HT2, T2ol
9	F. acuminatum	T2, NEO
10	F. subglutinans	MON

AcDON, monoacetyl-deoxynivalenols (3-AcDON, 15-AcDON); BEA, beauvericin; DAS, diacetoxyscirpenol; DON, deoxynivalenol (Vomitoxin); ENS, enniatins; FUS, fusarenone-X (4-Acetyl-NIV); HT2, HT-2 toxin; MON, moniliformin; NEO, neosolaniol; NIV, nivalenol; T2, T-2 toxin; T2ol, T-2 tetraol; ZEN, zearalenone; ZOH, zearalenols (α and β isomers).

The could place accompanied epidemics infection with *F. sporotrichioides* and *F. poae* may be lead to occurrence of T–2 derivatives (T2, HT2, T2ol), and DAS and MAS, respectively in infected grain, NIV and FUS toxin has also been founded to correlative to the activity of *F. poae* and *F. cerealisin* Sweden and other northern countries in grains from central to northeast Europe.^{42,43}

MON was reported to be founded in freshly harvested durum wheat in Austria up to 0.88mg kg⁻¹.⁴⁴ In all these surveys, the MON contamination in kernels was well correlated with the *F. avenaceum* infection.⁴⁵ Zearalenone is secondary metabolite produced by many *Fusarium* species, especially by *F. graminearum* and *F. culmorum*, this toxin has known as estrogenic properties, which means it can cause infertility, abortion, or other breeding problems. As little as 1 to 5ppm zearalenone in a feed ration may produce an estrogenic effect in swine.⁴⁶ Study done by Matny⁴⁷ about companion *Fusarium*

pathogens to FCR and FHB to wheat, it's found that *F. verticillioides* and *F. proliferatum* can produce with high amount of several kind of mycotoxin like DON, Fumonosin, ZEN and T2 toxin.

Economic impact of mycotoxins

Mycotoxins have significant economic impacts in many crops, especially in grain like wheat, maize, peanuts and other nut crops, cottonseed, and coffee. The Food and Agriculture Organization has estimated that 25% of the world's crops are contaminated by mycotoxins each year, with annual losses of around 1billion metric tons of foods and food products. Economic losses occur because of:

- i. Yield loss due to diseases induced by toxigenic fungi.
- ii. Reduced crop value resulting from mycotoxin contamination.
- Losses in animal productivity from mycotoxin-related health problems.
- iv. Human health costs.

Additional costs associated with mycotoxins include the cost of management at all levels prevention, sampling, and research costs. These economic impacts are felt all along the food and feed supply chains: crop producers, animal producers, grain handlers and distributors, processors, consumers, and society as a whole (due to health care impacts and productivity losses). Reduced crop value is a significant component of the losses caused by mycotoxins. This affects crops entered into local trade as well as crops intended for export.⁴⁸

Fusarium head blight losses to barley producers in the upper Midwest of the USA over the years (1992–1997) have been estimated to as high as \$364million. Deoxynivalenol reduces barley quality, lowering producer prices, and limiting the amount of uninfected barley available for the malting and brewing industries, which are not willing to accept barley with detectable levels of deoxynivalenol because conditions during malting promote Fusarium growth, leading to malted samples containing higher level of the toxin than in the original seeds. The limited amount of uninfected barley in the U.S. has had effects on the malting and brewing industry, which paid and extra \$200million in 1994–1995 to get non–infected grain from Canada.⁴⁹

Estimates of mycotoxins losses costs in the United States vary, one report estimated \$0.5 to \$1.5billion/year and another estimated \$5billion/year for the U.S. and Canada. Neither of these estimates included human health impacts or crop yield losses. Losses associated with deoxynivalenol in the United States were estimated at \$655 million/year, with the majority of the losses in wheat. In developing countries, few estimates are available. Human health impacts of mycotoxins are the most difficult to quantify. These effects are due to acute (single exposure) toxicoses and immunosuppression by mycotoxins, as well as chronic (repeated exposure) effects. Diseases modulated by mycotoxins accounted for 40% of lost disability—adjusted life years in a 1993 World Bank report on human health. Of the reported \$900million impact of mycotoxin in Southeast Asia, \$500million of the costs were related to human health effects.

Trichothecenes toxins are the mean secondary metabolite produced by *Fusarium* spp, its very toxic to mammalian. Mode of action of these toxins is though inhibition of protein and DNA synthesis, impairment of ribosome function, inhibition of mitochondrial protein synthesis, induction of reparable single strand breaks in DNA and immunosuppression.⁵⁰

Swine are very sensitive to *Fusarium* mycotoxins in their diet. Deoxynivalenol can cause reduced feed intake, and vomiting, reduced body weight gain or body loss. Deoxynivalenol has also been associated with abortions, stillbirths, and weak piglets. Immunosuppression with its associated increases in infection and disease incidence also increases production costs.⁵¹ Immune cells are more sensitive to DON than other cell types due to the induction of a T–cell activation response by increased intracellular calcium levels.⁵²

Zearalenone affects reproduction in swine and can have major economic effects in number of pigs produced per sow per year determine profit margins. Based on 1991 prices, it was calculated that a 10 or 20% reduction in farrowing rate combined with a 10 or 20% reduction in growth (as may occur if deoxynivalenol and zearalenone contaminated feed was consumed) would result in a 17 to 44% reduction in profit margins, due to increased feeding and veterinary costs per head, and a decline in the number of pigs marketed.⁵³

Conclusion

This study highlights on the economic loses consequent form disease damage to plant and reflected on yield, and health risk on consumers whether is human or animal, The fact that the pathogen *Fusarium* spp is responsible for producing mycotoxins in the infected cereal and the straw that produced from this plants. Therefore, preventive measures should be taken to reduce pathogen infection and spread, by using Integrate pest management for the disease through using resistance cultivars, biological control, rotation, crop management etc.

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Conflict of interest

The author declares no conflict of interest.

References

- 1. FAO. World Food Situation; 2014.
- United States Department of Agriculture (USDA). Global Barley Production. USA: United States Department of Agriculture; 2014.
- Murray G, Brennan J. Economic importance of wheat disease in Australia. Orange, NSW: NSW Agriculture; 1998.
- Dodman RL, Wildermuth GB. Inoculation Methods for Assessing Resistance in Wheat to Crown Rot Caused by Fsariurn grannineurn Group 1. Australian Journal of Agricultural Research. 1987;38(3):473–486.
- Backhouse D, Burgess LW. Climatic analysis of the distribution of Fusarium graminearum, F. pseudograminearum and F.culmorum on cereals in Australia. Australasian Plant Pathology. 2002;31(4):321–327.
- Scott J, Akinsanmi O, Mitter V, et al. Prevalence of *Fusarium* crown rot pathogens of wheat in southern Queensland and northern New South Wales. 12th AAC, proceedings of the 4th International Crop Science Congress, Brisbane, Australia; 2004.
- Backhouse D, Abubakar A, Burgess L, et al. Survey of Fusarium species associated with crown rot of wheat and barley in eastern Australia. Australasian Plant Pathology. 2004;33(2):255–261.
- Snijders CHA. Breeding for resistance to Fusarium in wheat and maize. In: Miller JD, et al. editors. Mycotoxins in Grain – Compounds other than Aflatoxin. USA: Eagan Press; 1994. p. 37–58.

- 9. Boutigny AL, Ward TJ, Van Coller GJ, et al. Analysis of the *Fusarium graminearum* species complex from wheat, barley and maize in South Africa provides evidence of species—specific differences in host preference. *Fungal Genet Biol*. 2011;48(9):914–920.
- Parry DW, Jenkinson P, McLeod L. Fusarium ear blight (scab) in small grain cereals – a review. Plant Pathology. 1995;44(2):207–238.
- Xu X-M, Parry DW, Nicholson P, et al. Predominance and association of pathogenic species causing *Fusarium* ear blight in wheat. *European Journal of Plant Pathology*. 2005;112(2):143–154.
- Wise K, Woloshuk C. Fusarium Head Blight (Head Scab). Purdue Extension: 2004
- Fernandez MR, Pearse PG, Holzgang G, et al. Fusarium head blight in common and durum wheat in Saskatchewan in 1999. Can Plant Dis Surv. 2000:80:57–59.
- 14. Fernandez MR, Holzgang G, Celetti MJ, et al. The incidence of Fusarium head blight in barley, common wheat and durum wheat grown in Saskatchewan during 1998. *Can Plant Dis Surv.* 1999;79:79–82.
- McMullen M, Jones R, Gallenberg D. Scab of wheat and barley:a reemerging disease of devastating impact. *Plant Disease*. 1997;81(12):1340– 1348.
- Burgess LW, Backhouse D, Summerell BA, et al. Crown Rot of Wheat. In: Summerell BA, et al. editors.USA: APS Press; 2001. p. 271–294.
- 17. Bushnell WR, Hazen BE, Pritsch C. Histology and physiology of *Fusarium* head blight. In: Leonard KJ, et al. editors. *Fusarium Head Blight of Wheat and Barley*. USA: APS Press; 2003. p. 44–83.
- Jon AA, DeWolf E, Todd T, et al. Kansas cooperative plant disease survey report preliminary 2014 Kansas wheat disease loss estimates; 2014.
- Goswami RS, Kistler HC. Heading for disaster: Fusarium graminearumon cereal crops. Mol Plant Pathol. 2004;5(6):515–525.
- Murray G, Brennan J. Estimating disease losses to the Australian wheat industry. Australas Plant Pathol. 2009;38(6):558–570.
- Akinsanmi OA, Mitter V, Simpfendorfer S, et al. Identity and pathogenicity of *Fusarium* spp. isolated from wheat fields in Queensland and northern New South Wales. *Aust J Agr Res*. 2004;55(1):97–107.
- Smiley R, Gourlie J, Easley S, et al. Crop damage estimates for crown rot of wheat and barley in the Pacific Northwest. *Plant Disease*. 2005;89(6):595– 604.
- 23. Nicol JM, Rivoal R, Trethowan RM, et al. IMMYT's approach to identify and use resistance to nematodes and soil fungi developing superior wheat germplasm. In: Bedo Z, et al. editors. Wheat in a Global Environment, USA: Kluwer Academic Publishers; 2001. p. 381–389.
- Windels CE. Economic and social impacts of *Fusarium* head blight: Changing farms and rural communities in the northern Great Plains. *Phytopathology*. 2000;90(1):17–21.
- Liu C, Jill G. Finding the jewels in crown rot research. CSIRO Plant Industry No. 215; 2009.
- McMullen M, Zhong S, Neate S. Fusarium head blight (Scab) of small seed. Plant Disease Management. NDSU, ND Agriculture Experiment Station; 2008.
- Matny ON, Tawfeeq JA, Alorchan SH, et al. Investigation on the deoxynivalenol in rations and some imported and local ingredients in Iraq and its effect on the in vitro degradation. *J Food Industries & Nutr Sci.* 2012;2(1):77–85
- LBP Bavarian State Research Center for Agricultural Sciences and plant construction (2000) Risks from Ahren parasites. Fusarium graminearum results LBP research network.

- Matny ON. Detection of Deoxynivalenol, Zearalenone and T2–Toxin produced by *Fusarium* species in different cultures. *Intl J Agri Crop Sci*. 2013;5(20):2385–2389.
- 30. Burrows M, Grey W, Dyer A. Fusarium Head Blight (scab) of Wheat and Barley. Montana State University Extension; 2012.
- Champeil A, Dore T, Fourbet JF. Fusarium head blight: Epidemiological origin of the effects of cultural practices on head blight attacks and the production of mycotoxins by Fusarium in wheat grains. Plant Science. 2004;166(6):1389–1415.
- Pestka JJ. Toxicological mechanisms and potential health effects of deoxynivalenol and nivalenol. World Mycotoxin J. 2010;3(4):323–347.
- Matny ON, Chakraborty S, Obanar F, et al. Molecular identification of Fusarium spp causing crown rot and head blight on winter wheat in Iraq. J Agri Technol. 2012;8(5):1677–1690.
- 34. Miller JD, Greenhalgh R, Wang YZ, Lu M (1991) Trichothecene chemotype of three *Fusarium* species. Mycologia 83(2):121–130.
- 35. Gang G, Miedaner T, Schuhmacher U, et al. Deoxynivalenol and nivalenol production by *Fusarium culmorum* isolates differing in aggressiveness toward winter rye. *Phytopathol*. 1998;88(9):879–884.
- D'Mello JPF, MacDonald AMC, Postel D, et al. 3–Acetyl deoxynivalenol production in a strain of *Fusarium culmorum* insensitive to the fungicide difenoconazole. *Mycotoxin Res.* 1997;13(2):73–80.
- Leonov AN, Kononenko GP, Soboleva NA. Production of DON-related trichothecenes by *Fusarium graminearum* Schw from Krasnodarski krai of the USSR. *Mycotoxin Res.* 1990;6(2):54–60.
- 38. Bottalico A, G Perrone. Toxigenic *Fusarium* species and mycotoxins associated with head blight in small–grain cereals in Europe. *European J Plant Pathol*. 2002;108(7):611–624.
- Miller JD. Mycotoxins in small grains and maize:old problems, new challenges. Food Addit Contam Part A Chem Anal Control Expo Risk Assess. 2008;25(2):219–230.
- 40. Matny ON. Mycotoxin production by *Fusarium* spp isolates on wheat straw in laboratory condition. *Res J Biotechnol*. 2013;8(7):35–41.
- Visconti A, Chelkowski J, Bottalico A. Deoxynivalenol and 3– acetyldeoxynivalenol–mycotoxins associated with wheat head fusariosis in Poland. Mycotoxin Res. 1986;2(2):59–64.
- 42. Bottalico A, Logrieco A, Visconti A. Mycotoxins produced by Fusarium crookwellense. Phytopathologia Mediterranea. 1990;29(2):124–127.
- 43. Eriksen GS, Alexander G. *Fusarium toxins in cereals –a risk assessment.* TemaNord Food 1998:502, Nordic Council of Ministers. Expressen Tryk & Kopicenter, Copenaghen, Denmark; 1998. 146 p.
- Adler A, Lew H, Brodacz W, et al. Occurrence of moniliformin, deoxynivalenol, and zearalenone in durum wheat (*Triticum durum Desf.*). *Mycotoxin Res.* 1995;11(1):9–15.
- 45. Kostechi M, Szczesna J, Chelkowski J, et al. Beauvericin and moniliformin production by Polish isolates of *Fusarium subglutinans* and natural cooccurrence of both mycotoxins in cereal grain samples. *Microbiologie, Aliments, Nutrition.* 1995;13:67–70.
- Minervini F, Dell'Aquila ME. Zearalenone and Reproductive Function in Farm Animals. *Int J Mol Sci.* 2008;9(12):2570–2584.
- 47. Matny ON. Screening of Mycotoxin Produced by Fusarium verticillioides and F. proliferatum in culture media. Asian J Agriculture and Rural Development. 2014;4(1):36–41.
- 48. Schmale DG, Munkvold GP. Mycotoxins in crops: a threat to human and domestic animal health. *American Phytopathological Society*. 2009;DOI:10.1094/PHI–I–2009–0715–01.

- Dahleen LS. ARS Fusarium Workshop. Athens, Georgia, USA: Richard Russell Research Center; 1997. p. 19–20.
- Ueno Y, Nakajima M, Sakai K, et al. Comparative toxicology of trichothec mycotoxins: inhibition of protein synthesis in animal cells. *J Biochem*. 1973;74(2):285–296.
- 51. Pestka JJ. Deoxynivalenol:Toxicity, mechanisms and animal health risks. *Animal Feed Science and Technology*. 2007;137(3–4):283–298.
- 52. Katika MR, Hendriksen PJ, van Loveren H, et al. Characterization of the modes of action of deoxynivalenol (DON) in the human Jurkat T–cell line. *J Immunotoxicol*. 2014;2:1–11.
- 53. Charmley LL, Trenholm HL, Prelusky DB, et al. Economic losses and decontamination. *Nat Toxins*. 1995;3(4):199–203.