Fish preservation: a multi-dimensional approach

Abstract

Fish is one of the most nutritionally valuable and highly perishable foodstuffs. The delicacy of fish can be easily noticed from its rapid quality deterioration immediately after harvest, if held improperly and not processed promptly. The natural composition of fish and the contaminations they encounter during the processing are the main causes for the onset and subsequent undesirable quality changes in these commodities. The spoilage patterns of fish are broadly categorized as enzymatic, chemical and microbiological. The growth of pathogenic microorganisms as a result of contamination mostly does not produce detectable changes in the sensory qualities of fish, which adds to the challenge of fish processing. Thus, a number of preservation mechanisms have been and are being developed to tackle the quality and safety problems associated with fish and fishery products. Although every method can produce effective results, yet there is no single best method capable of ensuring the safety as well as halt the progress of spoilage at the same time. The various preservation methods used in the fish and fishery industry can be categorized into three as physical, chemical and bio–preservation methods. These methods have different modes of actions and are applied to extend the shelf life of fish and fishery products. The present review article provides an overview of some of the physical, chemical and biological preservation methods applied to fish and fishery products.

Keywords: fish preservation, fish quality and safety, fish spoilage, microbial safety and stability, shelf life extension, physical preservation, chemical preservation, hurdle technology

Introduction

The world population is growing drastically and thus, the demand for food has been increasing likewise. In the modern era, the awareness on the benefits of consuming nutritious food has made the demand for certain nutritious food stuffs, such as fish, to rank at the top of the highly demanded foodstuffs. According to FAO,1 global seafood consumption has increased dramatically from 9.9kg per capita in the 1960s to 20kg in 2016. In addition to this, the exports of fishery products in developing countries deposited an income of $80 billion and were estimated to be $148 billion on world scale exports in 2014.

The biochemical composition of live fish differs from species to species. Within the same species also their chemical composition differs depending on different factors such as, feed intake, migratory swimming and sexual changes in connection with spawning. Despite these variations, fish has proved to be one of the most nutritious food stuffs providing 20% of the protein intake of one third of the world’s population2 and is also the source of health–friendly oils, omega–3 (ω–3) polyunsaturated fatty acids (PUFAs). Moreover, they are a rich source of a number of micronutrients like, vitamin D and different minerals, which can be found in higher quantities in aquatic animals than in mammalian meats and plants.3

However, fish is also known for its high perishability nature. The rapid deterioration of quality of fish and other seafoods after their harvest is mainly due to the various mechanisms of spoilage that take place. The spoilage mechanisms associated with such deteriorations are grouped into microbial metabolic activities, endogenous enzymatic activities and chemical oxidation of lipids, all of which shorten the shelf life of seafoods. According to Gram and Huss,4 the high composition of non–protein nitrogen compounds and low acidity (pH>6) of the flesh of seafoods are the major cause of their spoilage, as these condition favor the growth of spoilage microorganisms. These microbes in turn produce metabolites that affect the organoleptic properties of the products and render them undesirable attributes.

Similarly, autolytic activities by endogenous enzymes of seafoods also results in products that initially cause loss of the characteristic fresh odor and taste of fish and then softens the flesh. These changes start short after the death of seafood animals and progresses to produce a number of volatile compounds which give the products their spoilage characteristics. The details of the mechanism of such changes have been described by several authors.5,7 These enzyme driven changes are often accompanied by microbial metabolic activities.

Chemical oxidation of lipids is a common spoilage pattern in fatty fish in particular. This spoilage pattern depends mainly on the presence of oxygen which is responsible for the oxidative rancidity of the polyunsaturated fatty acids of fish and fishery products.5

Even though the spoilage patterns of fish can be explained as three distinct groups, all the mechanisms progress simultaneously, speeding up the overall spoilage of the products. As already noted, the spoilage mechanisms can be driven by microbial growth, enzymatic activities or chemical reactions. Thus, the preservation methods intended to stop the various spoilage patterns are expected to target these three causes in different approaches.

Improper handling and processing of seafoods, including fish, can lead to its contamination and subsequent growth of pathogenic microorganisms. In addition, the natural occurrence of aquatic biotoxins and natural pathogenic flora of the aquatic environment also contribute to seafood borne diseases. Huss et al.,8 have classified seafoods into seven groups based on their risk of microbial contamination and the processing method undergone. Generally, processing mainly controls microbiological hazards but leaves chemical hazards or biotoxins virtually unaffected. Effective control of chemical hazards and biotoxins has to be applied mostly during primary production and the pre–harvest stages.9

The present article provides a brief overview of different preservation methods applied to fish and fishery products by...
categorizing them into three broad groups; physical, chemical, and microbiological methods.

**Physical methods of fish preservation**

The physical methods of fish preservation can be described as three distinct spoilage control measures, namely: moisture control, thermal control and non-thermal control technologies.

**Moisture control measures**

**Drying:** The availability of water or moisture is a critical need for microbial metabolism and growth. The free water available in the matrix of foods that support the growth of microorganisms is termed as water activity ($a_w$). High $a_w$ level is required for the growth of most microorganisms. The $a_w$ level of different microorganisms is different, majority of the bacterial species being the top based on their requirements for the highest levels (Table 1) and thus, are the main contributors of fish quality deteriorations. On the other hand, fish and fishery products are known for their high moisture content in their fresh status (Table 2) which makes them conducive environments for microbial proliferation. Thus, drying and curing are the oldest, simplest and widely used preservation methods intended to control the moisture content of food stuffs by lowering their $a_w$.

**Table 1** Minimum values of water activity for microbial growth.

<table>
<thead>
<tr>
<th>Microorganisms</th>
<th>Water activity ($a_w$)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Bacteria</td>
<td>0.91</td>
</tr>
<tr>
<td>Yeasts</td>
<td>0.85</td>
</tr>
<tr>
<td>Moulds</td>
<td>0.8</td>
</tr>
<tr>
<td>Halophilic bacteria</td>
<td>0.75</td>
</tr>
<tr>
<td>Xerophilic bacteria</td>
<td>0.65</td>
</tr>
<tr>
<td>Osmophilic bacteria</td>
<td>0.6</td>
</tr>
</tbody>
</table>

**Table 2** Chemical composition of the fillets of various fish species.

<table>
<thead>
<tr>
<th>Species</th>
<th>Scientific name</th>
<th>Water%</th>
<th>Lipid%</th>
</tr>
</thead>
<tbody>
<tr>
<td>Blue whiting</td>
<td>Micromesistius poutassou</td>
<td>79.0–80.0</td>
<td>1.9–3.0</td>
</tr>
<tr>
<td>Cod</td>
<td>Gadus morhua</td>
<td>78.0–83.0</td>
<td>0.1–0.9</td>
</tr>
<tr>
<td>Eel</td>
<td>Anguilla anguilla</td>
<td>60.0–71.0</td>
<td>8.0–31.0</td>
</tr>
<tr>
<td>Herring</td>
<td>Clupea harengus</td>
<td>60.0–80.0</td>
<td>0.4–22.0</td>
</tr>
<tr>
<td>Plaice</td>
<td>Pleuronectes platessa</td>
<td>81.0</td>
<td>1.1–3.6</td>
</tr>
<tr>
<td>Salmon</td>
<td>Salmo solar</td>
<td>67.0–77.0</td>
<td>0.3–14.0</td>
</tr>
<tr>
<td>Trout</td>
<td>Salmo trutta</td>
<td>70.0–79.0</td>
<td>1.2–10.8</td>
</tr>
<tr>
<td>Tuna</td>
<td>Thunnus spp.</td>
<td>71.0</td>
<td>4.1</td>
</tr>
<tr>
<td>Norway lobster</td>
<td>Nephrops norvegicus</td>
<td>77.0</td>
<td>0.6–2.0</td>
</tr>
<tr>
<td>Pejerrey</td>
<td>Basilichthys borniensis</td>
<td>80.0</td>
<td>0.7–3.6</td>
</tr>
<tr>
<td>Carp</td>
<td>Cyprinus carpio</td>
<td>81.6</td>
<td>2.1</td>
</tr>
<tr>
<td>Sabalu</td>
<td>Prochilodus platensis</td>
<td>67.0</td>
<td>4.3</td>
</tr>
<tr>
<td>Pacu</td>
<td>Colossoma macropomum</td>
<td>67.1</td>
<td>18.0</td>
</tr>
<tr>
<td>Tambaqui</td>
<td>Colossoma brachypomum</td>
<td>69.3</td>
<td>15.6</td>
</tr>
<tr>
<td>Chincuina</td>
<td>Pseudoplatsostoma tigrinum</td>
<td>70.8</td>
<td>8.9</td>
</tr>
<tr>
<td>Corvina</td>
<td>Plagioscion squamosissimus</td>
<td>67.9</td>
<td>5.9</td>
</tr>
<tr>
<td>Bagre</td>
<td>Ageneiosus spp.</td>
<td>79.0</td>
<td>3.7</td>
</tr>
</tbody>
</table>

**Figure 1** Solar tent drier (Adapted from Doe et al.16)

**Thermal control**

Low temperature treatments are the methods used immediately on board following the harvest of aquatic products. Chilling in various forms including, wet ice, refrigerated seawater (RSW), chilled seawater (CSW), solid and liquefied forms of carbon dioxide (CO$_2$), liquid nitrogen, and chill air have been reported by several authors. It has been recommended by FAO10 that fish should be cooled to chilling temperature (0°C) immediately after capture. According to Olley & Ratkowsky,19 the preservative effect of chilling is due to its ability to reduce the growth rate of spoilage microorganisms and their metabolic activities and thus maintains the post-harvest quality changes to the minimum. Such treatments can extend the shelf life of fish and fishery products only for days, as certain endogenous autolytic enzymes and proteolytic enzymes produced by contaminant microbes can deteriorate the quality in a short-run.
In RSW, the product is cooled to a temperature much lower than the melting ice treatment. This is due to the reduced freezing point of seawater attributed to the presence of salts. Thus, RSW has been used as a better option for chilling operations onboard of fishing vessels for its greater cooling rate, reduced pressure and physical damage on fish, less exposure to oxygen as well as more rapid handling of large quantities of fish with little delay or labor involvement, and in many cases, an extended shelf life. However, the high chance of spread of spoilage microbes in the circulating water, excessive uptake of salt, uptake of water by low fat species, loss of protein and problems with anaerobic bacteria are non–negligible drawbacks of RSW systems.

Freezing has been used as a long term preservation technique for decades. Fish and other meat foods were reported to keep for months under proper frozen storage. The treatment retards microbial growth and enzymatic activity in fish to a greater extent when compared to chilling and thus, preserves the flavor and nutritional properties better. However, the formation of ice crystals during freezing has been observed to affect the quality of frozen products. With the formation of large crystals, textural damage and membrane disruption with subsequent oxidation has been reported by Alizadeh et al.

**Non–thermal control:** Non–thermal physical preservation approaches employed in the seafood industry include: high pressure processing, packaging technologies, irradiation, ozonation, as well as pulsed electric fields and oscillatory magnetic fields treatments. Of these, there are insufficient reports on the preservative potential and suitability of the latter two in their use on fish and fishery products. The term advanced oxidation process is often used to describe treatments such as, ozonation and ultraviolet (UV) irradiations, to refer to the processes that involve oxidation processes used to eliminate inorganic and organic contaminants and/or pollutants with the help of reactive free radicals produced by ozone ($O_3$), hydrogen peroxide ($H_2O_2$), titanium oxide ($TiO_2$), zinc oxide ($ZnO$), and UV radiations.

**High pressure treatments:** High pressure treatments have almost same effect on microorganisms and enzymes as high temperature treatments. These technologies are based on the observation that microorganisms are inactivated due to lethal structural and biochemical alterations caused to the cells as a result of residence under such high pressures. Moreover, the cell deformation and damage associated with high pressure treatments were reported to be reversible with treatments in the range of 100–300MPa. However, these treatments fail to inactivate resistant bacterial spores. Such spores require pressure treatments as high as 1200MPa for their inactivation.

Several authors have reported the pros and cons of such treatment and supported their statements with findings. Riz et al. have reported on the synergistic effect of high pressure treatment at 200MPa and sub–zero temperature treatment to have inhibitory effect on *Listeria monocytogenes* in smoked salmon. They have also observed lightening of color and toughening of texture of the product as undesirable outcomes. Similarly, Matejková et al. reported on the possibility of shelf life extension of vacuum–packed trout exposed to high pressure treatment stored at 3.5°C, from 5–6days to 21–28days, in reference to the reduced content of biogenic amines observed on the treated product. On the contrary, pressure treatments resulted in bright and good organoleptic characteristics at the same time.

The study used a standard laboratory scale unit with a pressurization chamber of 2.5liters volume capacity. The treatment was done at a pressure range of 150–600MPa. The results obtained indicated that treatment of 300MPa for rainbow trout and 450MPa for mahi were the optimum levels to control microbial load, lipid oxidation and color changes.

**Ozonation:** Ozone has been studied for its potential in disinfecting and improving the sensory quality of seafoods. Gonçalves stated that antimicrobial effect of ozone depends on its concentration, contact time, temperature and the production system used. According to Gonçalves, low concentrations of ozone in solutions can decrease the half–life of oxygen and thus weakening its bactericidal effect.

In a study conducted by Mendonça et al., treatment of whole fresh water tilapia with 1.5ppm ozonated water for 15min was sufficient to result in 88.25% reduction of initial microbial load. The same study also reported a reduction of 77.2% and 79.49% of microbial load in fillets of the same fish species treated with ozonated water at concentrations of 1 and 1.5ppm respectively. The results were obtained with no pH and color alterations of the fillets. However, initiation of lipid oxidation process was detected in reference to increased thiobarbituric acid reactive substances assay (TBARS) values.

**Irradiation:** Irradiation in foods is described in three different ways that do not involve heat treatments. Radurization (cold pasteurization) involves the use of low doses (<1kGy) which inactivates non–spore forming bacteria. Radicidation reduce the number of viable and non–spore forming pathogenic bacteria and parasites using intermediate doses (2–8kGy). Radapertization (cold sterilization) is used to eliminate highly pathogenic bacteria such as, *Clostridium botulinum* at high doses (>10kGy).

Irradiation doses of 1.0 to 2.0kGy has been suggested by Matches & Liston for the effective elimination of contaminant micro–organism in raw fish and seafood dishes. Low dose irradiation has also proved to be effective for fish preservation when applied in combination with chilling or heating processes. However, the destruction of large populations of bacterial spores requires high doses of radiation. Sikorski & Sun Pan stated that even medium doses of 4–10kGy can induce undesirable sensor changes, mainly off flavors in fish and fishery products. Thus, irradiation processes fail to achieve both, complete elimination of all types of microorganisms and maintain sound organoleptic characteristics at the same time.

**Packaging technologies:** Packaging technologies involving either the replacement of air by certain gases (such as, $CO_2$, $O_2$, $N_2$ or combination of all) or removal of air and thus reducing the headspace to possible minimum has been used in the preservation of muscle and plant foods. Such packages are often termed as modified atmosphere packaging (MAP) and vacuum packaging (VP). The use of packaging material with low gas and water permeability is very crucial in such technologies in order to prevent the exchange of gases between the interior and exterior of the packaged product and thus maintain the right atmospheric conditions around the product.

The preservative effect of such technology is based on the atmospheric condition they create around the product. This condition establishes a significant hurdle for most strictly aerobic and facultative spoilage microorganisms. Moreover, lipid oxidation can also be prevented to some extent. Anon recommended that the retail pack for lean fish and shell fish to be composed of a mixture of $CO_2$,$N_2$,$O_2$.
Fish preservation: a multi-dimensional approach

Curing

The term curing is referred to the addition of salt, sugar, nitrates, nitrates, seasonings or spices and phosphates to preserve foodstuff. In addition to preservation, this particular processing can result in a characteristic color and flavor of the cured products. Salt has been the most widely used ingredient for curing of meat foods. Its main antimicrobial mode of action is creating a low a₁₀₀ environment in the food which forces microorganisms to experience prolonged lag phase and eventually enter the death phase.

Although this preservation method is one of the oldest and simplest fish preservation methods, the growth of halophilic and halotolerant microorganisms cannot be arrested and often their growth produce white patches on the fish fillets. Moreover, salting has also been reported to be a pro-oxidant agent in several meat and seafood products. Yet, few reports support the neutral effect of salt on lipid oxidation and as antioxidant.

Smoking: Smoking is a means of chemical preservation applied to meat foods. Although the process involves heat treatment to some extent, the preservative weight is attributed to its chemical products. It has been reported that smoking has both antimicrobial and antioxidant effects. Varlet et al., has reported that the combined effect of phenolic compounds produced during smoking process and the high temperature condition result in reduced microbial growth and oxidation. Moreover, carbonyl compounds produced during the process can contribute to the characteristic color, texture and fishy odor of the product. Thus, the preservation objectives of smoking include, surface drying which acts as a barrier for microbial invasions; reduces water activities resulting from the pretreatment process with salt; deposition of phenolic compounds which aid in delaying autoxidation of fish lipids and deposition of antimicrobial substances (such as, aldehydes, ketones, alcohols, acids, hydrocarbons, esters, phenols, ethers, etc.).

Two methods of smoking are practiced for fishery products, cold smoking and hot smoking, depending on the temperature used. Products treated with cold smoking is carried out at around 30°C following salting and drying treatment of fish muscle. Smoked salmon is a common product prepared by this treatment and eaten raw. Hot smoking on the other hand, involves treatment at a temperature range of 70–80°C and results in cooked product that do not require subsequent processing. Although smoked lean fish are preserved longer compared to fatty fish, fumigation, storage temperature and salt content, are determinant factors of the shelf life of smoked products. Several reports have been produced on the effect of smoking on various fish species (Table 3). Details of the smoking method and its mode of actions are described by Varlet et al., Varlet et al., Many authors have suggested the use of smoking in combination of one or more preservation method for highly effective results.

<table>
<thead>
<tr>
<th>Species</th>
<th>Smoking process</th>
<th>Product</th>
<th>References</th>
</tr>
</thead>
<tbody>
<tr>
<td>Trout (gutted, split)</td>
<td>Cold/hot</td>
<td>Smoked trout</td>
<td>54</td>
</tr>
<tr>
<td>Eel (gutted)</td>
<td>Cold/hot</td>
<td>Smoked eel</td>
<td>57</td>
</tr>
<tr>
<td>Salmon (gutted, split)</td>
<td>Cold</td>
<td>Smoked salmon</td>
<td>58</td>
</tr>
<tr>
<td>Mackerel (fillets)</td>
<td>Hot</td>
<td>Smoked mackerel</td>
<td>59</td>
</tr>
<tr>
<td>Herring (fillets)</td>
<td>Cold</td>
<td>Kipper fillets</td>
<td>60</td>
</tr>
<tr>
<td>Haddock</td>
<td>Cold</td>
<td>Haddock</td>
<td>41</td>
</tr>
</tbody>
</table>

The traditional smoking process is carried out by direct exposure of foods to smoke. However, such treatments can lead to the accumulation of harmful substances (such as, polycyclic aromatic hydrocarbons (PAHs)) which result from incomplete combustion of wood. These compounds have potential adverse effects on human health. Moreover, Munasinghe et al., have reported the high possibility of uneven distribution of volatile compounds on the fish flesh treated by direct exposure to smoke, which can affect the quality of the end product undesirably.

Natural antimicrobial preservatives

Naturally occurring antimicrobial substances such as essential oils (EOs) and extracts of various plants, spices, and herbs are reported to have effective activity against a number of pathogenic and spoilage microorganisms associated with foods. A comprehensive detail of major components of EOs including, carvacrol, thymol, g–terpinene and p–Cymene (oregano and thyme), a–pinene, bornyl–acetate, camphor and 1,8–cineole (rosemary), eugenol and eugenyl acetate (clove, etc.) has been reviewed by Burt. These natural extracts have been incorporated in the packaging materials as well as edible films and coatings to enhance the keeping quality of minimally processed and delicate foods such as meat, fish and fruits.

In a study carried out by Fernandez Saiz et al., the antimicrobial activity of chitosanamide acetate films on Salmonella spp., Staphylococcus aureus and Listeria monocytogenes showed a direct correlation with the film weight, i.e., cell viability was reduced completely when high amounts of film (40–80mg) were used. However, the antimicrobial effect of the film was found to be lower in the fish soup compared to the results obtained in TSB culture media, although the use of the film did neither affect the sensory properties nor the pH of the soup. Similarly, Tsai et al. investigated the effect of chitosan films on fish spoilage microbes and reported an improvement of shelf life of treated fishery product from 5 to 9days. Several other authors have studied the biocide mode of action of this polysaccharide in great details and reported satisfactory results.

According to the findings of Iturriaga et al., the antimicrobial activity of citrus extract (water soluble extract) and the EOs, oregano...
and thyme were evident (Table 4). Their findings were illustrated as the antimicrobial activities of these substances tested against *Listeria innocua* (surrogate of *Listeria monocytogenes*), *Pseudomonas fluorescens* (dominant spoilage bacterium in fishery products) and *Aeromonas hydrophila*. Though the three proved to be effective, the authors suggested that citrus extract to be the best option to incorporate into three biopolymers gelatin, methyl cellulose and their (50:50w/w) blend to produce edible film for its odorless and water solubility properties as well as highest antimicrobial activity.

### Table 4 Minimum inhibitory concentration (MIC) obtained by disc diffusion method for the three active extracts against the tested bacteria (Adapted from Iturriaga et al. 72)

<table>
<thead>
<tr>
<th>Extract</th>
<th>Temperature</th>
<th>MIC(µl/ml) a</th>
<th><em>L. innocua</em></th>
<th><em>P. fluorescens</em></th>
<th><em>A. hydrophila</em></th>
</tr>
</thead>
<tbody>
<tr>
<td>Citrus extract</td>
<td>T opt b</td>
<td>2.5±0.0</td>
<td>40.0±0.0</td>
<td>10.0±0.0</td>
<td></td>
</tr>
<tr>
<td></td>
<td>4°C</td>
<td>1.3±0.0</td>
<td>26.7±11.5</td>
<td>6.7±2.9</td>
<td></td>
</tr>
<tr>
<td>Oregano EO</td>
<td>T opt b</td>
<td>5.0±0.0</td>
<td>5.0±0.0</td>
<td>2.5±0.0</td>
<td></td>
</tr>
<tr>
<td></td>
<td>4°C</td>
<td>5.0±0.0</td>
<td>9.7±2.9</td>
<td>2.5±0.0</td>
<td></td>
</tr>
<tr>
<td>Thyme EO</td>
<td>T opt b</td>
<td>5.0±0.0</td>
<td>5.0±0.0</td>
<td>2.5±0.0</td>
<td></td>
</tr>
<tr>
<td></td>
<td>4°C</td>
<td>4.2±1.4</td>
<td>5.0±0.0</td>
<td>2.5±0.0</td>
<td></td>
</tr>
</tbody>
</table>

a Values given as Mean±SD (n=3)
bT opt, optimal growth temperature, 37°C for *L. innocua*, 30°C for *P. fluorescens* and *A. hydrophila*

Some finding have revealed that the effectiveness of such antimicrobial substances also depend greatly on temperature. Bagamboula et al. 73 suggested that microorganisms tend to be more susceptible to antimicrobials at exposure temperatures close to their optimal due to their increased metabolic activity. Their suggestion was based on their observations on higher reductions in the population of *Shigella* spp. When treated at 22°C than at 7°C. On the other hand, the combined effect of more than one natural extract was found to be more effective than their single use. Ilhak & Gurun 74 have concluded that a combination of 0.1% thymol and 2% sodium lactate showed a synergistic effect on *Salmonella* Typhimurium on days 3 and 5 of storage of fish patty, although the sole use of 2% sodium lactate showed significant inhibition against *Listeria monocytogenes* than the aforementioned treatment. A number of other natural antimicrobial compounds have been investigated by Cheng et al. 75; Rubén et al. 76; Speranza et al. 77; Ortiz–Viedma et al. 78; Raesi et al. 79

**Organic acids:** Organic acids and their salts have also been reported to have positive impact on the quality and safety of food products. According to Sallam, 79 the growth of spoilage flora and lipid oxidation of salmon fillets packed in polyvinylidene film were maintained at the lowest pace for 15 days at 1°C when treated with 2.5%(w/v) aqueous solution of sodium acetate, sodium lactate, or sodium citrate dip for 10minutes at 4°C. Similarly, cold smoked salmon brine injected with potassium lactate and sodium acetate were observed to inhibit the growth of *L. monocytogenes* at a storage temperature of 4°C for 32days. 80 The same results were also reported on the same product and treatment stored at 10°C in vacuum package for 42days.80

### Microbiological methods of fish preservation

This is a method of tackling the growth of undesirable microorganisms by favoring the growth of competitive and antagonist (to the undesirable microbes) in a food product. The phenomenon is often termed “biopreservation”. In this method, either the bacterium or the metabolite it produces, play the role preservation. Lactic acid bacteria (LAB) are the most widely studied and dealt with in the area. LAB are not considered as indigenous microbes of the aquatic environment nor aquatic animals. But, many species (such as *Lactobacillus spp.*, *Carnobacterium spp.*, *Aerococcus spp.*, *Enterococcus spp.*, *Lacticoccus spp.*, etc.) are also reported to be associated with different fish products. The preservative action of LAB is attributed to either their competition with the undesirable microbes for nutrients or the antimicrobial metabolites they produce or both.81 A list of biopreservation studies made on fish by different authors is given in a review article authored by Ghanbari et al. 82 Table 5 illustrates some of the bacteriocins produced by LAB species and their target organisms.83

<table>
<thead>
<tr>
<th>Bacteriocin</th>
<th>Produced by</th>
<th>Active against</th>
</tr>
</thead>
<tbody>
<tr>
<td>Nicin(s)</td>
<td><em>Lc lactis subsp lactis</em></td>
<td><em>Bacillus cereus</em>; <em>Clostridium botulinum</em>; <em>Staphylococcus aureus</em></td>
</tr>
<tr>
<td>Pediocin(s)</td>
<td><em>P acidilactis</em></td>
<td><em>Lactobacillus</em>; <em>Clostridium sporogenes</em></td>
</tr>
<tr>
<td>Reuterins</td>
<td><em>Lb reuteri</em></td>
<td><em>Listeria monocytogenes</em>; <em>Salmonella typhimurium</em>; <em>Shigella spp</em></td>
</tr>
</tbody>
</table>

The use of live microbial antagonists (biopreservatives) is suggested to follow certain criteria and requirement for safety reasons.84 In addition to their desired antagonistic effect against a wide spectrum of spoilage and pathogenic microorganisms as well as their capability to withstand varying processing technologies and storage conditions, these protective cultures should not pose any adverse health effect (i.e., they should be safe) and also neither them nor their metabolites should affect the sensory, chemical and physical properties of the end product in a way that confer the product with unattractive characteristics.85-86

Hurdle technology of fish preservation

Almost all preservation methods reveal poor effect when used individually. It might also require their use at the highest concentration or intensity, which in turn can result in sensorial and nutritional deterioration of the product. Leistner⁸ has suggested the use of more than one preservative factor (hurdle) to keep undesirable microbes in a hostile environment, which inhibits their growth and leads them to death. The principle of the technology involves the use of more than one hurdle at mild levels and subjects the microorganisms to multi-stress reactions by disturbing their homeostasis. This unstable homeostasis condition eventually results in their metabolic exhaustion and death.

Conclusion

The causes of quality deterioration in fish and fishery products are numerous. The preservation methods suggested to combat every cause are also different. The challenge of the preservation of such delicate and nutritious product is also noted to be multi-dimensional with no absolute or ideal preservation method capable of tackling the varying deterioration causes and relieving the tension of the challenge. It is a dynamic area in the science of food, which will keep introducing innovative technologies with the progress of technological developments.

Acknowledgements

We would like to acknowledge our respective institutes for providing us with access to various databases from their archives and from the internet. We would also like to express our appreciation to the FAO for granting us to access their database (AGORA).

Conflict of interest

The authors hereby, declare that there is no conflict of interest in publishing this article.

References

1. Food and Agriculture Organization (FAO). Global per capita fish consumption rises above 20 kilograms a year. Italy; 2016.
Fish preservation: a multi-dimensional approach


Fish preservation: a multi-dimensional approach


