Isolation and Quantification of three Genes involved in the Expression of Anthocyanins in *Rubus glaucus* Benth by RT-qPCR

**Abstract**

The violet and purple colored fruits contain phenolic compounds specifically, anthocyanins. These have a high nutritional quality and anthocyanins are believed to have anti-aging, anti-carcinogenic and antioxidant properties in the fruit. Gene expression studies have demonstrated the importance of flavonoids and their biosynthetic pathways, further increasing the interests of research and sources. In the present study, for the isolation and quantification of the β-Actin, RuANS and RuMYB10 genes involved gene pathway induces anthocyanin in *Rubus glaucus* Benth, the Real-Time Polymerase Chain Reaction Reverse transcription (RT-qPCR). According to the values obtained with the statistical analyzes DCA and the Tukey test at 95% confidence, observed a relationship between genes as no significant differences existed in the expression of the three, generating the following results: β Actin presented higher concentration (273.42 ng/mL), followed by the RuMYB10 gene (220.33 ng/mL) and finally the RuANS gene (212.75 ng/mL), concluding that the species studied had similar levels of expression three genes analyzed.

**Keywords:** Anthocyanins; β-Actin gene; RuANS gene; RuMYB10 gene; RT-qPCR; *Rubus glaucus* Benth

**Abbreviations:** RT-qPCR: Real-Time Polymerase Chain Reaction Reverse Transcription; ANOVA: Analysis of Variance; ANS: Anthocyanidin Synthase

**Introduction**

Anthocyanins are secondary metabolites involved mainly in the ripening of fruits. These phenolic compounds are found in leaves, flowers and above all in the fruits of plants. They are incorporated into the human body in daily intake, so the greatest interest lies in the fruits of violet colors [1]. According to Wu X et al. [2], the estimated daily intake of anthocyanins is 12.5 mg / day / person in the United States, Germany in 2002, daily intake of 2.7 mg / person, ranging from 0 to 72 mg / person [3].

At present, interest in the properties of anthocyanins has increased; Miyazawa et al. [4] state that anthocyanins do not undergo any changes during their passage through the digestive tract into the bloodstream, thus generating their therapeutic effect in reducing coronary disease, improving visual acuity, anticancer effects, antiaging, antitumor and anti-inflammatory [5], thanks to its antioxidant activity [6].

The *Rubus glaucus* Benth also called mora of Quito [7], is a plant of wild origin of the Andes. It is the most produced variety in Ecuador; due to its national and international demand, it is cultivated in diverse countries like: Colombia, Chile, Panama, Guatemala, Mexico and Canada. The fruit of blackberry is consumed both fresh and processed, currently the main markets for this fruit Europe and the United States, have increased the consumption of fruits and vegetables, taking into account the nutritional factor in their diet [8].

The main objective of the present investigation is to carry out molecular analyzes of gene expression in the biosynthetic pathway of anthocyanins. For this, the RT-qPCR allows us to make qualitative and quantitative measurements of specific genes in a given sample; due to its sensitivity, speed, accuracy specificity can be used as a more accurate method than conventional techniques [9]. For the purposes of this analysis, the β-actin gene has been used as a reference gene, as it is one of the most widely used because of its high concentrations [10]. This allows normalization of a standard curve and generation of reliable data in the analysis [11].

In this work, we compare the genes most directly associated with anthocyanin biosynthesis in the level of transcription of the gene that specifically controls this compound is anthocyanidin synthase for Rubus (RuANS) and RuMYB10 that encodes a transcription factor and is used for specific regulation in the route [12].

The knowledge of genes as well as their functioning and expression in the various species, allows the opening to a new era of Transcriptomics and biotechnology in Ecuador [13], because it reaches the genomic knowledge not only to understand but also their applications in the field of health, production, quality and improvement of food.

**Materials and Methods**

The research was carried out in the Laboratories of Life Sciences at the Salesian Polytechnic University, Laboratory of Molecular Biology, located in Ecuador, Pichincha Province, Canton Quito, El Girón Campus.
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Collection of samples for analysis
The plant material was collected in the province of Tungurahua, Canton Patate, at an altitude of 2220 masl, was collected according to INEN regulations in force in the country. Two field visits were carried out and 1 kg of fruit were collected aseptically each time, taking into account the organoleptic characteristics. Cryopreserved samples were taken to the Laboratories at the Salesian Polytechnic University for the relevant treatments and analyze.

RNA extraction with pure link® RNA mini kit (ambion, life)
The fruits of *Rubus glaucus Benth* were macerated in an Eppendorf tube with 1mL of the Lysis Buffer solution, homogenized and centrifuged for 2min at 12000 x g. In one Eppendorf tube was placed 1mL of the supernatant plus 500mL of ethanol (70%). Then, 700μL were taken, added in a column tube and centrifuged for 15s at 12000 x g. 700μL of Wash Buffer I was added and centrifuged for 15s at 12000 x g. The liquid discarded and placed in a new collection tube, 500μL of Wash Buffer II was added and centrifuged for 15 sec at 12,000 x g. The liquid discarded and centrifuged for 2min at 12000x g. Finally, 50 μL of free Water of ARNases was added and the centrifuge was taken for 2min to 12,000 x g.

Transverse transcription with first strand transcriber cDNA synthesis kit (Roche)
In a microtube, 9 μl of extracted RNA, 1 μl of vial 5 (Anchored-digo (dT) Primer), 2 μl of vial 6 (Random Hexamer Primer), 1 μl of vial 7 (H2O PCR grade) were placed. 13 μL of solution were obtained, which were taken to the thermocycler for 10 min at 65°C and cooled to 4°C. Subsequently, 7 μL of the Master Mix was placed in the microtube together with the solution. The thermocycler was taken for 10min at 25ºC for 10min, 55ºC for 30min, 85ºC for 5min and the cooling at ∞ at 4°C.

Standard curve
For the elaboration of the standard curve we used *Rubus glaucus* with the β-Actin gene, Lin-Wang and others [14] mention that β-Actin is selected as a control gene because of its constant level of transcription along leaves and fruits. To obtain the reference values, logarithmic serial solutions of the complementary DNA obtained previously were prepared, the samples were quantified in the Qubit® 2.0 Fluorometer, with Qubit® dsDNA HS Assay Kit (Life) [13].

Real-time PCR in light cycler 2.0 (Roche)
Samples were prepared in each glass capillary with 15 μL of the master mix plus 5 μL of the *Rubus glaucus* cDNA sample. For the negative controls 5 μL of water (PCR grade) and the programming of the RT-qPCR: initial denaturation, target temperature 95°C, 1 cycle; cycled 60 times with three different target temperatures: 95, 60 and 72°C. Melting, 1 cycle with target temperatures of 95, 65 and 95°C, cooling, 1 cycle at a temperature of 40°C.

Results and Discussion
The standard curve points were obtained from the serial concentrations of cDNA on a logarithmic scale in the Qubit® fluorometer for the β-Actin gene, whereby the quantification of the unknown samples could be performed. The standard curve in acceptable parameters presents an efficiency of 1,922 and an error close to 0, providing certainty in the tests [13].

Analysis of variance (ANOVA) with a 95% confidence level [15] classifies genes as three homogeneous groups that present with each other, means with statistically non-significant differences, with F=2.20 and p=0.1273. To corroborate the data obtained, a Tukey test was used at 5%, concluding that the 3 genes make up the same group A, so that they are involved in the same way in the biosynthesis of anthocyanins, as can be observed in Figure 1.

Figure 1: Tukey test for the β-Actin, RuANS and RuMYB10 genes in *Rubus glaucus* Benth.

Figure 2 shows a run in the Real-Time PCR with three of the curves obtained for the analysis of the expression of the β-Actin, RuANS and RuMYB10 genes in *Rubus glaucus* Benth, thus confirming that all three have similarity, since that the Ct is very close to the 25 points between the cycles and the fluorescence and therefore the concentration is always inversely proportional to the Ct.

The mean of the concentrations obtained is as follows: for the reference gene 273.42 ng / mL, gene expression is presented with 220.33 ng / mL for RuMYB10 and 212.75 ng / ml for the RuANS.
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Table 1: Concentrations (ng/mL) of the β-Actin, RuANS and RuMYB10 genes in Rubus glaucus.

<table>
<thead>
<tr>
<th>Genes</th>
<th>Repetitions</th>
<th>β - Actin</th>
<th>ANS</th>
<th>MYB10</th>
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<td>240</td>
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<td>average(x)</td>
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<td>273.42</td>
<td>213</td>
<td>220.33</td>
</tr>
</tbody>
</table>

Quantitative PCR, due to its sensitivity, speed, accuracy and specificity, can be used to analyze gene expression in a more precise way than conventional techniques, allowing quantitative measurements to be obtained in small quantities of samples that are difficult to collect and generate valuable contribution in the isolation of genes of interest and their application in the industrial field [1].

Conclusions

The objectives set out in the present research were fulfilled and the gene expression of β-actin, RuANS and RuMYB10, involved in the metabolic pathway of anthocyanins in rubus glaucus by RT-qPCR; concluding that the species studied presents a similar level of expression among the three analyzed genes.

Currently in Ecuador, we are working on other native species that could even be vulnerable to extinction, the use of molecular biology as a tool for biodiversity management can support the monitoring of natural species and populations, as well as their repowering, use and genetic improvement.

Acknowledgement

To María Elena Maldonado.

Conflict of Interest

None.

References


Citation: Chiluisa-Utreras V, Vaca I, Chicaiza O, Pehaherrem S, Acuicio RD (2017) Isolation and Quantification of three Genes involved in the Expression of Anthocyanins in Rubus glaucus Benth by RT-qPCR. MOJ Food Process Technol 1(3): 00129. DOI: 10.15406/mojfpt.2017.05.00129