

Zaltoprofen Loaded Solid Lipid Nanoparticles for Topical Delivery: Formulation Design, *In Vitro* and *Ex Vivo* Evaluation

Abstract

The objective of present study is development and evaluation of solid lipid nanoparticles (SLNs) of Zaltoprofen (ZLT) for topical delivery to formulate topical formulation with prolonged effects. ZLT loaded SLNs (ZLT-SLN) were formulated using Taguchi design of Experiment and the effect of lipid ratio, surfactant ratio, stirring time and concentration of surfactant was studied on entrapment efficiency. The optimized formulation was spherical in shape with mean particle size distribution of 208nm and zeta potential of -25mV. They were characterized for structural changes using Differential Scanning Colorimetry and Fourier Transform Infrared Spectroscopy. Optimized nanoparticles were further incorporated in carbopol hydrogel and characterized for pH, homogeneity, viscosity and spreadability. *In vitro* and *Ex vivo* studies were carried out for ZLT-SLN loaded carbopol gel and compared against ZLT-SLN suspension. ZLT-SLN suspension followed Higuchi cube root model whereas ZLT-SLN loaded gel followed Korsmeyer Peppas model in *in vitro* as well as *ex vivo* studies which may be due to increase in viscosity. SLN loaded gel showed sustained release as compared to plain drug loaded hydrogel confirming that SLN are efficient to encapsulate ZLT and effective for sustained release of drug.

Keywords: Zaltoprofen; Solid lipid nanoparticles; Topical delivery; Solvent injection method

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Abbreviations: ZLT: Zaltoprofen; BCS: Bio-Pharmaceutics Classification System; DDS: Drug Delivery System; FTIR: Fourier Transformed Infrared; Gm: Gram; HCL: Hydrochloric Acid; Hrs: Hour; IP: Indian Pharmacopeia; JP: Japanese Pharmacopeia; LOD: Loss On Drying; SLNs: Solid Lipid Nanoparticles; GMS: Glycerol Monostearate; DRG: Dorsal Root Ganglion

Introduction

The number of new pharmaceutical compounds with poor aqueous solubility has grown steadily over the last two decades due to the progress in discovery technologies, e.g. high throughput and combinatorial screening tools, during lead development and optimization phase. Most of these compounds are mainly categorized as bio pharmaceutics classification system (BCS) class II for which their oral absorption is limited by their extremely poor solubility in the gastrointestinal tract [1]. Many potential compounds frequently drop out of pharmaceutical development process because of their insufficient bioavailability. Therefore, the improvement of solubility and dissolution of poorly water-soluble drugs has become vital for formulation researchers [2]. Delivering the drug accurately and safely to its target site at precise time in order to have a controlled release and achieve maximum therapeutic effect remains a benchmark in the design and development of newer drug delivery systems. Nanoparticles aid to provide targeted delivery of many drugs, to sustain the effect of drug in target tissue, to improve oral bioavailability and to increase the stability of therapeutic agents against degradation

[3].

SLNs are colloidal carrier system composed of a high melting point lipid as a solid core coated by aqueous surfactant and the drugs used are of BCS Class II and IV [4]. Solid lipid nanoparticles (SLN) represent a promising approach as a drug delivery system for topical application [5,6]. Similar studies have been performed on number of drugs like Meloxicam [7], soy isoflavones [8], flurbiprofen [9] and Ibuprofen [10], was formulated with the aim of better penetration through stratum cornea and efficient deposition of drug in dermal matrix. Further they were evaluated for *in vitro* drug release characteristics, rheological behavior, pharmacokinetic and pharmacodynamics studies and showed excellent physical stability, high entrapment efficiency and controlled drug release can be produced representing a promising carrier for topical delivery of drugs [11].

Zaltoprofen (2-(10-oxo-10, 11, dihydrodibenzo [b, f] thiepin-2-yl) propionic acid belongs to the class of NSAIDs, a low molecular weight drug that acts by inhibiting the bradykinin-induced responses by blocking bradykinin interaction with the bradykinin β_2 receptor on dorsal root ganglion (DRG) neurons [11]. Recent study on Zaltoprofen opened up its potential to be delivered as nanosuspension [12], Nanoemulsion gel [13] and as drug loaded biopolymeric nanoparticles [14]. All these results suggest that zaltoprofen possesses novel anti-inflammatory mechanism, which inhibits β_2 -type BK receptor function in nerve endings and shows potential to be delivered using novel drug delivery systems.

Materials and Methods

Materials

Zaltopfen API was received as a gift sample from IPCA Laboratories, Mumbai. Glycerol Monostearate and Tween 80 were purchased from Loba Chemie, Poloxamer 188 was obtained from BASF, Methanol and Acetonitrile (HPLC grade) were purchased from SRL Fine chemicals, Carbopol 940, Triethanolamine, Methyl paraben, Propyl paraben were purchased from SD Chemicals. Millipore water was used throughout the study.

Screening of lipids and solvents

The aim of this study was to develop a solid lipid nanoparticle for topical formulation. Considering solubility of drug in lipids as one of the most important criteria, lipid and solvents were selected cautiously. The solubility of drug in lipid phase can affect the ability of the nanosuspension to maintain the drug in a solubilized form which is important in maintaining stability. The solubility of ZLT in various lipids was determined by adding an excess amount of drug in 100mg of selected lipid (Stearic acid, Glycerol monostearate, Compritol 888 ATO, Precirol 888 ATO and Lipoid S-75), in 10mL capacity beaker using a magnetic stirrer. Similarly different volatile organic solvents (Acetone, Dichloromethane, Ethanol and Methanol) exhibiting good solubility for zaltopfen were screened for its ability to solubilize the lipid.

Preparation of solid lipid nanoparticles

Based on the solubility studies of drug, Glycerol monostearate [15] was selected as the lipid phase and Methanol as solvent. Tween 80 and poloxamer 188 were selected as surfactant and stabilizer respectively. Millipore water was used as an aqueous phase for optimization. Several batches of SLNs were prepared using different combinations of lipid and surfactant ratio. Modified Solvent Injection Method [9,16] was used to formulate ZLT loaded solid lipid nanoparticles. Glycerol monostearate (GMS) and drug Zaltopfen were dissolved in methanol and warmed to $70 \pm 2^\circ\text{C}$. Tween 80 and Poloxamer 188 were dissolved in distilled water to prepare aqueous phase. The organic phase of alcoholic solution containing lipid mixture was added drop wise to pre-warmed aqueous solution ($70 \pm 2^\circ\text{C}$) with the help of a hypodermic needle on a magnetic stirrer for varying time periods to obtain nanoparticles with optimum size.

Optimization of solid lipid nanoparticles

In this experiment, Taguchi design (34) was adopted to optimize ZLT-SLN formulation process in order to derive optimal conditions. The trial studies were based on three levels (high, medium, low). Drug: Lipid ratio, Lipid: Surfactant ratio, Concentration of stabilizer and stirring time were considered as independent factors. Entrapment efficiency was considered as dependent factor in the proposed study. All statistical studies were performed using Design-Expert 9.0 software, manufactured by Stat-Ease, Inc. Analysis of variance was carried out to determine the significance of the fitted equation. The formulation was chosen with the criteria of high entrapment efficiency and absence of sedimentation. The process variables, their coded experimental values, are listed in Table 1.

Characterization of solid lipid nanoparticles

Entrapment efficiency: 2ml of solid lipid nanoparticles were centrifuged at 23500rpm for 90 minutes. The free drug content in supernatant after centrifugation was measured by Ultraviolet spectroscopy [17]. The calibration curve of absorbance against concentration of Zaltopfen was plotted. Equation obtained from the graph is shown below:

$$y = 0.0126x - 0.0835, r^2 = 0.9996$$

where;

y=Absorbance,

x=Zaltopfen concentration

From the above equation, entrapment efficiency was calculated based on given formula:

$$\% \text{Entrapment Efficiency} = (\text{Amount of drug added} - \text{Amount of drug free in supernatant} \times 100) / (\text{Amount of drug added})$$

Measurement of particle size: Photon co-relation spectroscopy using Zetasizer (Malvern Instruments, UK) determined globule size of the nanoemulsion by the principle of fluctuation in light scattering due to Brownian motion of the particles [18].

Measurement of zeta potential: Zetasizer (Malvern Instruments) was used to measure Zeta Potential [18]. Samples were prepared for analyzing zeta potential by diluting 1ml of the SLN dispersion with 10ml Millipore water.

Transmission electron microscopy: The morphology of particles was examined by TEM (Hitachi 2000 Japan). Combination of bright field imaging at increasing magnification and of diffraction modes was used to reveal the form and size of the nanosuspension [18].

Differential scanning calorimetry: Scan of about 2mg, accurately weighed zaltopfen and ZLT-SLN formulation using a differential scanning calorimeter was recorded. Sealed and perforated aluminium pans were used in experiments for all the samples. Sample was run at the scanning rate of $10^\circ\text{C}/\text{minute}$ from $30 - 150^\circ\text{C}$.

Incorporation of solid lipid nanoparticles into gel: 1% w/w Carbopol 940 was dispersed in drug loaded solid lipid nanosuspension. Carbopol 940 was incorporated in nanosuspension and soaked overnight. The formulation was stirred with alkalizing agent triethanolamine till pH 6 and the gel of desired viscosity is obtained [19,20].

Characterization of gel: ZLT-SLN loaded gel was characterized for drug content, pH, Homogeneity, Viscosity and Spreadability. Every test was performed in triplicate [21].

Drug content: Gel equivalent to 100mg of ZLT was taken and dissolved in 100mL of methanol in a volumetric flask. Gel solution was subjected to shaking for 2h on mechanical shaker to obtain complete solubility of drug. The solution was estimated spectrophotometrically at 331nm using methanol as solvent.

pH of gel: 1gm of gel was taken and dissolved in 100ml of purified water and stored for two hour. The three samples were checked for pH using pH meter (Lab India).

Table 1: Design for optimization of solid lipid nanoparticles using Taguchi design of experiment.

Batch No	Factor 1	Factor 2	Factor 3	Factor 4
	Drug: Lipid Ratio	Lipid: Surfactant Ratio	Conc of Stabilizer (%)	Stirring Time (Hours)
ZLT001	01:05	01:00.5	0.10%	4
ZLT002	01:05	01:02	0.50%	8
ZLT003	01:07	01:01	0.10%	8
ZLT004	01:06	01:00.5	0.30%	8
ZLT005	01:07	01:00.5	0.50%	6
ZLT006	01:05	01:01	0.30%	6
ZLT007	01:06	01:01	0.50%	4
ZLT008	01:06	01:02	0.10%	6
ZLT009	01:07	01:02	0.30%	4

Homogeneity: All developed gels were tested for homogeneity by visual inspection after the gels have been set in the container.

Viscosity: Brookfield digital viscometer was used to measure the viscosity of prepared gel formulations. The spindle no. 6 was rotated at 10rpm. The reading, near to 100% torque was noted. Samples were measured at $30 \pm 1^\circ\text{C}$.

Spreadability study of topical gel: Spreadability was determined using apparatus consisting of wooden block and 11 pulleys at bottom. 'Slip' and 'Drag' principle were used to measure the spreadability of gel. About 2gm of gel was placed on ground slide. Another glass slide was placed on ground slide and entrapped air between the slides was removed. 1Kg of weight was put on the slide for 5 minutes. Excess of gel was cleaned from the corner of slide 80gm weight was placed on the pulley and time required by the top slide to move a distance of 7.5cm was reported. A short interval indicates better spreadability [22]. Spreadability was then calculated using the following formula:

$$S = M \times L / T$$

Where,

S: Spreadability,

M: Weight in the pan (tied to the upper slide),

L: Is the length moved by the glass slide and

T: The time taken to separate the slide completely from each other.

In vitro drug release study: *In vitro* release studies were performed using modified Franz diffusion cells. Dialysis membrane having pore size 2.4nm molecular weight cut off 1200-14000 (HIMEDIA) was used. Membrane was soaked in phosphate buffer pH 6.8 overnight. Zaltoprofen formulation was filled in donor compartment and the recipient compartment was filled with phosphate buffer pH 6.8. The content of the cell was stirred with the help of magnetic stirrer at 32°C . At fixed time interval predetermined aliquots of sample was withdrawn from receiver compartment through side tube. Fresh phosphate buffer of pH 6.8 was used to replace the withdrawn medium to maintain sink

condition. Samples were analyzed by UV spectrophotometer [23].

Ex vivo drug release study: *Ex vivo* release studies were performed using modified Franz diffusion cells. Porcine ear skin was used. Skin was soaked in phosphate buffer pH 6.8 overnight. Zaltoprofen formulation was filled in donor compartment was filled in donor compartment and the recipient compartment was filled with phosphate buffer pH 6.8. The content of the cell was stirred with the help of magnetic stirrer at 32°C . At fixed time interval predetermined aliquots of sample was withdrawn from receiver compartment through side tube. Fresh phosphate buffer of pH 6.8 was used to replace the withdrawn medium to maintain sink condition. Samples were analyzed by UV spectrophotometer [23].

Drug diffusion kinetics study: The *in vitro* and *ex vivo* release pattern was evaluated to study flux. Mass transport of molecules in a solution or molecular transport across a barrier is normally measured by flux. The flux of a solute is simply defined as the mass or number of molecules moving through a given cross-sectional area during a given period of time:

$$J = m / At$$

Where;

J: Is the flux of a mass of compound

m: Moving through a cross-sectional area

At: During time t.

The unit for a flux value could thus be $\text{mol cm}^{-2} \text{min}^{-1}$, or alternatively $\text{mol cm}^{-2} \text{min}^{-1}$, or alternatively $\text{mg cm}^{-2} \text{h}^{-1}$ [21].

The *in vitro* and *ex vivo* release pattern was evaluated to check the goodness of fit to the zero order kinetics, first order kinetics, Higuchi's square root of time equation, Hixson Crowell cube root equation, Korsmeyer Peppas power law equation.

Stability studies: The optimized SLN loaded gel formulation was stored at $40^\circ\text{C}/75\% \text{RH}$, 4°C and room temperature for a period of three months and *in vitro* release was determined to evaluate its stability.

Results and Discussion

Screening of lipids and solvent

To assess solubility of Zaltopufen, various lipids were screened. Steric acid, Glycerol Monostearate, Compritol 888 ATO, Precirol 888 ATO and Lipoid S-75 were chosen from preliminary screening of lipids. Zaltopufen showed excellent solubility in Glycerol monostearate and Lipoid S-75 having a melting point of 60°C and 40°C respectively. Zaltopufen showed good solubility in Acetone, Ethanol, Methanol and Dichloromethane. Depending on solubility of the lipids in solvent, Methanol was selected as a solvent. Solid lipid nanoparticles were formulated using Solvent Injection Emulsification and Evaporation technique using Glycerol monostearate and Lipoid S-75 as lipid. Formulation with Lipoid S-75 showed larger particle size (2 microns) than that with Glycerol monostearate (400nm). Latter showed an acceptable particle size. Therefore Glycerol monostearate was selected as lipid, Tween 80 as surfactant, poloxamer 188 as a stabilizer, Methanol as solvent and water as continuous phase.

Analysis of optimization design

In the present study different formulation batches were prepared on the basis of independent factors including concentration of lipid, surfactant, stabilizer and stirring time and were analyzed for maximum entrapment efficiency. Taguchi's Design of Experiment was adopted for optimization studies which listed a run of nine designs. Shapiro Wilk test showed p value more than 0.1 with no terms selected, which meant that there are no statistically significant differences on studied effect. Hence it may be concluded that no factors were significantly affecting entrapment efficiency of Solid Lipid Nanoparticles for Zaltopufen. The formulation batches exhibiting high entrapment efficiency were selected for further screening. The three formulations were studied for average particle size (Table 2). From the above results Batch no. ZLT007 with an average particle size of 208nm (Table 3).

Table 2: Results for entrapment efficiency to optimize the solid lipid nanoparticles formulation.

Batch No	% Entrapment Efficiency (%w/w)	Sedimentation
ZLT001	65	Observed
ZLT002	48.1	Observed
ZLT003	64.35	Not Observed
ZLT004	35.62	Observed
ZLT005	55.43	Not Observed
ZLT006	65.5	Not Observed
ZLT007	70.6	Not Observed
ZLT008	33.21	Not Observed
ZLT009	46.43	Observed

Characterization of optimized batch

Batch no. ZLT007 with an average entrapment of 70.6% and average particle size of 208nm and polydispersity index 0.456

was optimized formulation. Zeta potential of optimized batch was found to be -25mV indicating stability of the formulation.

Table 3: Results for particle size analysis to optimize the solid lipid nanoparticles formulation.

Batch	Particle Size	Polydispersity Index
ZLT003	277.7nm	0.608
ZLT006	359.6nm	0.672
ZLT007	208.0nm	0.456

Transmission electron microscopy

TEM was conducted to investigate the morphology of Zaltopufen loaded SLNs. It was evident from TEM images that nanoparticles were almost spherical with smooth morphology, well dispersed and separated on the surface (Figure 1).

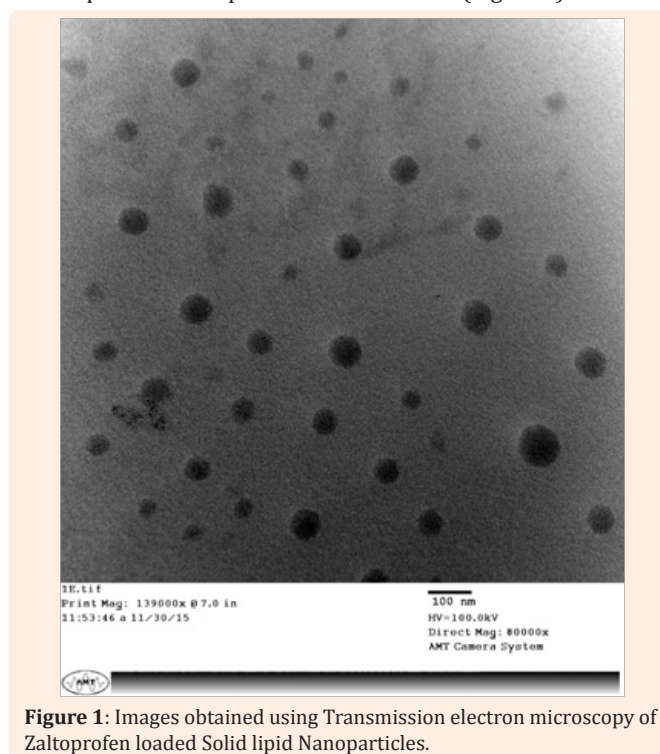


Figure 1: Images obtained using Transmission electron microscopy of Zaltopufen loaded Solid lipid Nanoparticles.

Differential scanning calorimetry

DSC is a tool to investigate the melting and recrystallization behavior of crystalline material like SLNs. Figure 2 & 3 shows the DSC thermograms of pure Zaltopufen and Zaltopufen loaded SLNs respectively. Plain Zaltopufen showed a sharp endothermic peak at 136.6°C corresponding to its melting point, indicating its characteristic crystalline nature. As observed in Figure 3, the thermogram of ZLT-SLN (ZLT007) showed no peak near 136.6°C as shown by plain Zaltopufen Figure 2 indicating the conversion of crystalline ZLT to the amorphous form which confirmed complete dissolution of the drug in the molten lipid matrix.

Fourier transform infrared spectroscopy

Figure 4 shows an overlapped FTIR spectra of Placebo solid

lipid nanoparticles and ZLT loaded nanoparticles. The spectra suggest that no additional peak of Zaltopfen was seen which indicate the entrapment of API inside the lipid matrix. Absorption

peaks observed for Zaltopfen at 2980,1703,1671,1280,799,752 cm^{-1} are masked in the FTIR spectra observed for ZLT loaded solid lipid nanoparticles.

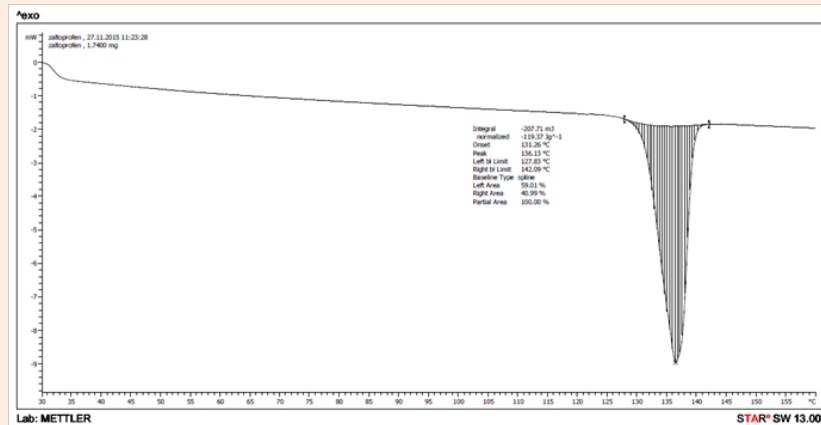


Figure 2: DSC thermogram of API Zaltopfen.

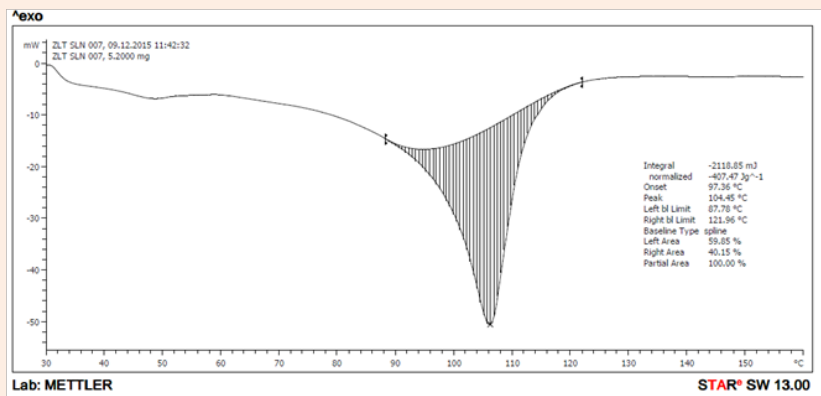


Figure 3: DSC thermogram of ZLT loaded SLN.

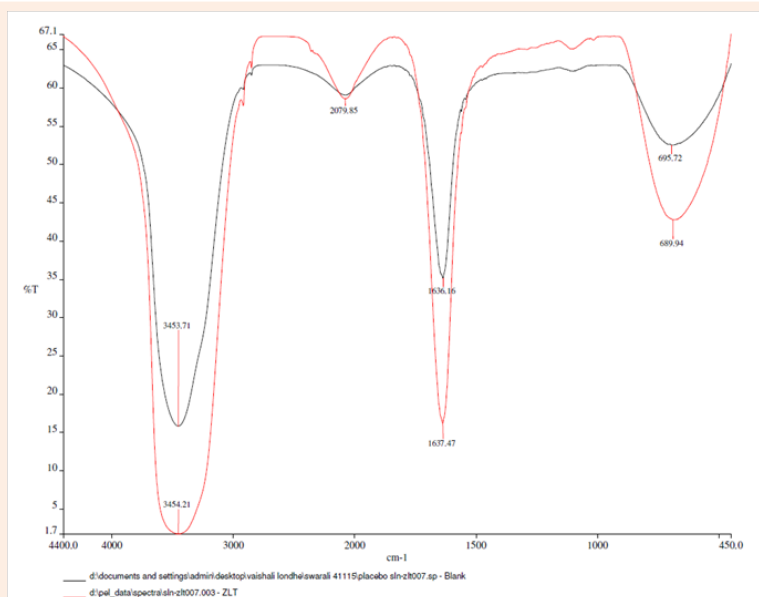


Figure 4: Comparison of FTIR spectra of Placebo and Zaltopfen loaded solid lipid nanoparticles.

Evaluation of SLN loaded gel

Formulated gel appeared to be white and translucent and displayed homogeneity in texture. The pH of gel was observed to be 6.5 ± 1 which is compatible with pH of skin. Total drug content was noted as 90%. The viscosity and spreadability of zaltoprofen solid lipid nanoparticle loaded topical gel was found to be 1898 poise and 206.8gm/sq.cm respectively.

In vitro diffusion studies

The cumulative drug release from optimized formulation batch of ZLT solid lipid nanoparticles and ZLT SLN loaded gel was found to be 53.7% and 46.3% respectively. The hydrophobic drug incorporated into solid lipid showed prolonged release of drug, which improves release profile of Zaltoprofen (Figure 5).

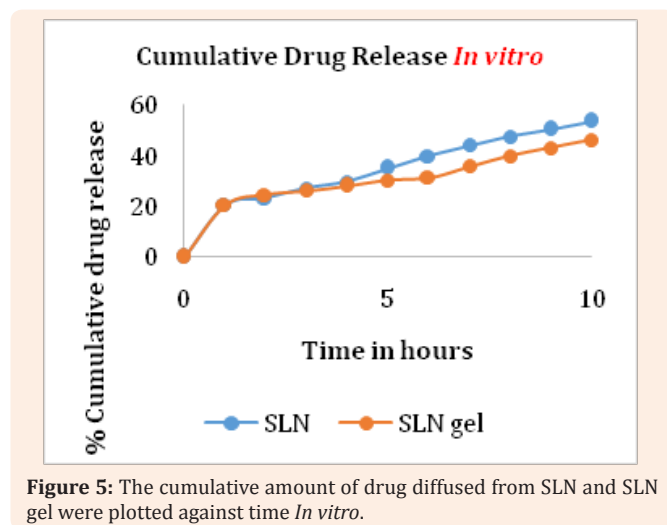


Figure 5: The cumulative amount of drug diffused from SLN and SLN gel were plotted against time In vitro.

Ex vivo release study

Ex vivo release study was performed using porcine ear skin. The cumulative drug release from optimized formulation batch of ZLT solid lipid nanoparticles and ZLT SLN loaded gel was found to be 49.6% and 36.8% respectively. The hydrophobic drug incorporated into solid lipid showed prolonged release of drug, which improves release profile of Zaltoprofen (Figure 6).

Drug diffusion kinetic study

Mass transport across a barrier, e.g. intestinal epithelium or other pharmaceutically relevant barriers, can be caused by diffusion or migration. Transport by simple diffusion can be measured as flux, and characterized by a permeability for the transported solute. Flux calculated by equation 4.1 at 10th hour is as given below (Table 4): The values of R² for different mathematical models are shown in table. The R² values for zaltoprofen loaded solid lipid nanoparticles follows Higuchi square root model and ZLT SLN loaded carbopol gel depicts that Korsmeyer Peppas model fits best (Table 5).

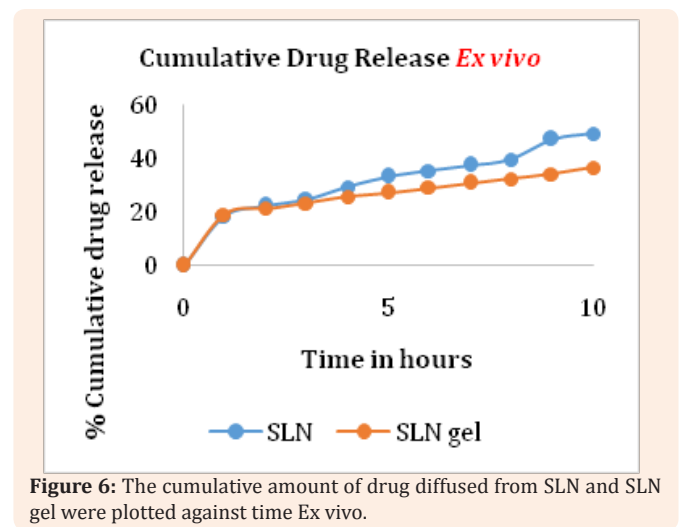


Figure 6: The cumulative amount of drug diffused from SLN and SLN gel were plotted against time Ex vivo.

Table 4: Flux at 10th hour of diffusion study.

In vitro SLN	63.67µg/cm ² /hour
In vitro SLN gel	59.53µg/cm ² /hour
Ex vivo SLN	63.67µg/cm ² /hour
Ex vivo SLN gel	56.98µg/cm ² /hour

Table 5: Drug diffusion kinetics for In vitro and Ex vivo drug release study.

	Zero Order Model	First Order Model	Higuchi Square Root Model	Hixson Crowell Cube Root Model	Korsmeyer Peppas Model
In vitro ZLT SLN	0.94	0.97	0.98	0.97	0.97
In vitro ZLT SLN gel	0.89	0.93	0.96	0.92	0.97
Ex vivo ZLT SLN	0.93	0.96	0.98	0.95	0.97
Ex vivo ZLT SLN gel	0.85	0.89	0.96	0.88	0.97

Stability studies

Table 6 and Figure 7.

Table 6: Results for stability studies of ZLT loaded SLN gel.

Formulation	RT	40°C/75%RH	4°C
Physical Observation	No Change	Slight Yellow	No Change
% Assay	88%	56%	92%

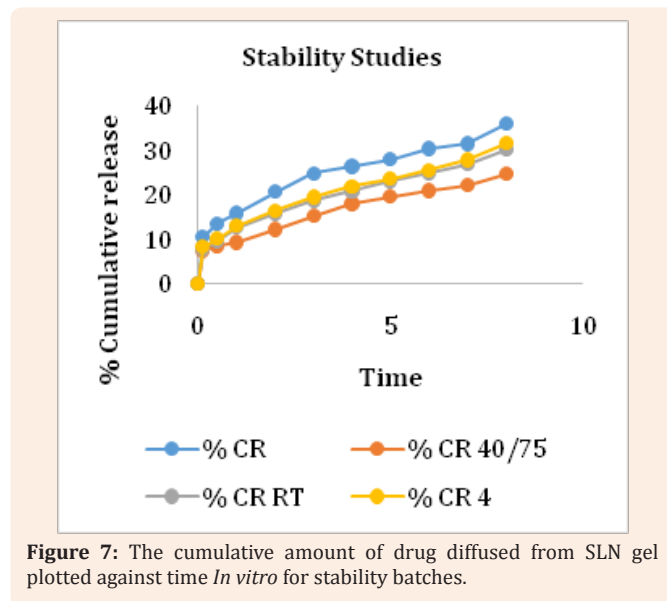


Figure 7: The cumulative amount of drug diffused from SLN gel plotted against time *In vitro* for stability batches.

Conclusion

The objective of the present investigation was to explore potential of Zaltoprofen loaded Solid Lipid Nanoparticles as a drug carrier system to be applied topically. Zaltoprofen, as a NSAID, was entrapped in glycerol monostearate using solvent injection, emulsification and evaporation method. The formulation was optimized by experimental design considering the concentrations of drug and lipid, surfactant, stabilizer and stirring time as independent variables. The entrapment of drug in lipid was confirmed by FTIR and DSC studies. Zaltoprofen loaded Solid lipid nanoparticles were characterized by particle size, morphology, zeta potential, and PDI and % entrapment efficiency. The optimized design was obtained using Taguchi design. The resultant solid lipid nanoparticles were spherical with particle size of 208 nm and entrapment efficiency of 70.6%. The -25.3 zeta potential indicated good stability of nanoparticles and PDI of 0.456 indicated aggregation free system. *In vitro* and *Ex vivo* diffusion studies showed that drug undergoes sustained diffusion from optimized formulation over a period of 10h. Zaltoprofen loaded solid lipid nanoparticles loaded in topical gel formulation is an alternative to tablet by virtue of its sustained and extended release and its ability to decrease side effects caused due to oral administration of Zaltoprofen. Also topical application of Zaltoprofen yields better patient compliance.

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Conflict of Interest

The authors of this article certify that they have no affiliations or involvement in any organization or entity with any financial or non-financial interest in the subject matter discussed in the manuscripts.

References

- Papdiwal A, Pande V, Sagar K (2014) Design and characterization of zaltoprofen nanosuspension by precipitation method. *Scholars Research Library Der Pharma Chemica* 6(3): 161-168.
- De Jong WH, Geertsma RE, Roszek B (2005) National Institute for Public Health and the Environment (RIVM). Bilthoven, Netherlands.
- Loxley A (2009) Solid Lipid Nanoparticles for the Delivery of Pharmaceutical Actives. *Drug Delivery Technology* 9(8): 1-5.
- Yadav N, Khatak S, Singh S (2013) Solid lipid nanoparticles - a review. *International Journal of Applied Pharmaceutics* 5(2): 8-18.
- Müller RH, Mäder K, Gohla S (2000) Solid lipid nanoparticles (SLN) for controlled drug delivery - a review of the state of the art. *Eur J Pharm Biopharm* 50(1): 161-177.
- Khalil RM, El-Bary AA, Kassem MA, Ghorab MM, Ahmed MB (2013) Solid lipid nanoparticles for topical delivery of Meloxicam: development and *in vitro* characterization. 1st Annual International Interdisciplinary Conference, AIIC, Azores, Portugal, p. 24-26.
- Deshmukh K, Amin P (2013) Formulation and evaluation of solid-lipid nanoparticle based 0.1% Soy isoflavone dermal gels. *Journal of Pharmaceutical and BioSciences* 1(1): 7-18.
- Bhaskar K, Anbu J, Ravichandiran V, Venkateswarlu V, Rao YM (2009) Lipid nanoparticles for transdermal delivery of flurbiprofen: formulation, *in vitro*, *ex vivo* and *in vivo* studies. *Lipids Health Dis* 8: 6.
- Kumar PD, Subas CD, Subrata C, Soumen R (2012) Formulation and evaluation of solid lipid nanoparticles of a poorly water soluble model drug Ibuprofen. *IRJP* 3(12): 132-137.
- Dasgupta S, Ghosh SK, Ray S, Kaurav SS, Mazumder B (2014) *In vitro* & *In vivo* studies on lornoxicam loaded nanoemulsion gels for topical application. *Curr Drug Deliv* 11(1): 132-138.
- Tang HB, Inoue A, Oshita K, Hirate K, Nakata Y (2005) Zaltoprofen inhibits bradykinin-induced responses by blocking the activation of second messenger signaling cascades in rat dorsal root ganglion cells. *Neuropharmacology* 48(7): 1035-1042.
- Aher SJ, Jagdale S (2014) Preparation and characterization of zaltoprofen nanosuspension through quassi emulsification solvent diffusion. *Indo American Journal of Pharmaceutical Research* 4(12): 5959-5965.

13. Chodhary P, Aparna C, Srinivas P (2014) Formulation and Evaluation of Zaltoprofen Nanoemulsion gel. *IJPT* 6(2): 6552-6571.
14. Shah HA, Patel RP (2015) Statistical modelling of zaltoprofen loaded biopolymeric nanoparticles-Characterization and anti-inflammatory activity of nanoparticles loaded gel. *Int J Pharm Investig* 5(1): 20-27.
15. Patel M (2012) Development, Characterization and Evaluation of solid lipid nanoparticles as a potential anticancer drug delivery system. Masters of Science Thesis, The University of Toledo, USA, 51(4): 95.
16. Trotta M, Debernardi F, Caputo O (2003) Preparation of solid lipid nanoparticles by a solvent emulsification-diffusion technique. *Int J Pharm* 257(1-2): 153-160.
17. Shafiq S, Shakeel F, Talegaonkar S, Ahmad FJ, Khar RK, et al. (2007) Development and bioavailability assessment of ramipril nanoemulsion formulation. *Eur J Pharm Biopharm* 66(2): 227-243.
18. Sharma P, Nikam S, Chavan M (2014) Solid Lipid Nanoparticles: A Lipid Based Drug Delivery. *IPP* 2(3): 365-376.
19. Verma A, Singh S, Kaur R, Kumar A, Jain UK (2013) Formulation, optimization and evaluation of clobetasol propionate gel. *Int J Pharm Sci* 5(4): 666-674.
20. Patel J, Patel B, Banwait H, Parmar K, Patel M (2011) Formulation And Evaluation of Topical Aceclofenac Gel Using Different Gelling Agent. *Int J Drug Dev Res* 3(1): 156-164.
21. Shetty C, Babubhai S, Pathan I (2010) Development of Valdecoxib Topical Gels-Effect of Formulation Variables on the Release of Valdecoxib. *Int J Pharm Pharm Sci* 2(1): 70-73.
22. Barhate SD, Potdar B, Nerker P (2000) Development of Meloxicam sodium transdermal gel. *IJPRD* 2(5): 1-4.
23. Cordero JA, Alarcon L, Escribano E, Obach R, Domenech J (1997) A Comparative study of the transdermal penetration of a series of nonsteroidal anti-inflammatory drugs. *J Pharm Sci* 86(4): 503-508.