

Mini Review





Natural products remain an important source for drug development in the post—genomic era

Abstract

Natural products or their related derivatives were the most important source of officially-approved drugs owning to the huge chemical structure diversities and their biodiversities. Compared with traditional combinatorial chemistries and high-throughput screening methods, genomics-driven natural product discovery is a rational and an efficient alternative for the discovery of novel structures. In this mini review, process of genomics-driven natural product discovery is highlighted including the main steps, the relevant technologies and bioinformatics tools.

Keywords: natural product, drug discovery, genomics-driven

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Abbreviations: BGCs, biosynthetic gene clusters; antiS-MASH, antibiotics and secondary metabolite analysis shell; iChip, isolation chip; PK(S), polyketide (synthase); NRP(S), nonribosomal peptide (synthase)

Introduction

Drug discovery, the first critical step to identify rational lead candidates in the novel drug development, has always been a challenging, time-consuming and laborious scientific task requiring expertise and experience. Historically, natural products or their related derivatives were the main source of officially-approved drugs, with more than 50% of clinical drugs approved between 1981and 2014 were derived from natural products. Even in recent days, natural products have still continued to enter clinical trials or to be approved to market, including Trabectedin (ET-743),2 Halaven (eribulin mesylate), 3,4 Bryostatin⁵ and so on. It is believed that the huge chemical structure diversity and the biodiversity of natural products make the greatest contributions to the success of natural products. However, combinatorial chemistries and high-throughput screening in drug discovery over the past decades have darkened the honorable outlook of natural products to some extent, 6 which lead to the growing studies on the novel drug discovery methods in the post-genomic era.

Drug discovery in the post-genomic era

With the growing development of DNA sequencing and synthetic biology technologies (e.g. proteomics, metabolomics, bioinformatics), great interest has been renewed in natural product discovery. It has been already accepted that natural products are synthesized by specific metabolic pathways encoded by biosynthetic gene clusters (BGCs), based on which virtually all natural products could be identified by DNA sequencing and metagenomic analysis theoretically. In this view, the chemical space could be far more covered and much more novel chemical structures might be discovered such as those encoded by silent biosynthetic gene clusters and uncultured microorganisms.

Genomics-driven natural product discovery usually includes three main steps:

a. Identification of BGCs. Antibiotics from microbes or plants are directly linked to BGCs coding for proteins associated

with biosynthesis, resistance, regulation and transport. So how to prioritize and characterize orphan BGCs is the first crucial procedure from the growing genome sequences.^{8,9} Genome mining^{10,11} is a novel technology developed for the identification and characterization of orphan BGCs, which is designed to analyze the sequenced genome of specific organisms to identify and determine whether the gene clusters involved in the production of new antibiotics. Several bioinformatics approaches have been proposed to satisfy this purpose, such as HMMER, GOLD, NORINE, SBSPKS, SEARCHPKS, NRPSpedictor₂, plantiSMASH and so on, among which AntiSMASH (antibiotics and secondary metabolite analysis shell)¹² might be the most widely tools used. It is a comprehensive bioinformatic tool for automated genome mining including the annotation of entire gene clusters.

b. Activation of silent gene clusters. Natural products are usually synthesized through secondary metabolism, which is tightly regulated and considerable BGCs remain silent under laboratory growth conditions. Owning to this, activating these BGCs will make it possible to obtain the potential encoded natural products. Usually, strategies to activate the silent BGCs including physicochemical methods and genomics-driven approaches. It was shown that if the producing host was grown under alternative physicochemical conditions compared with the standard laboratory growth conditions, it will possess the ability to express the originally silent gene clusters. 13-15 Co-cultivation of microorganisms and the application of iChip (isolation chip) have also been proved to be successful in activating the silent gene cluster. 16 Genomics-driven approaches 17 including mainly genome editing and genome engineering technologies by means of genetic manipulation of the target genome is believed to the most efficient but challenging and time-consuming methods to activate the silent or weakly-expressed gene clusters. Researchers have proposed and verified different strategies in this field, such as heterologous expression of BGCs in heterologous hosts functionally, 18 manipulation of global and pathway-specific transcriptional regulators(and/or RNA polymerase or ribosomal proteins), 19 perturbation of epigenetic control(DNA/his tone methylation and acetylation).20



c. Combinatorial biosynthesis of natural products. With the advance in synthetic biology and genome engineering technologies, it becomes rational and possible to obtain produce key precursors and the core scaffolds exploiting rational engineering of biosynthetic pathways.^{21–24} On this basis, increasing attention have been paid to modular PKS/NRPS megaenzymes^{25,26} for rational production of complex PK/NRP scaffolds. It was reported firstly by Christian Hertweck et al.26 that the reprogrammed modular (type I) aur polyketide synthase was successful in generating luteoreticulin through rational genetic recombination and domain exchanges. Combinatorial biosynthesis of target products is a relatively much more sophisticated process including many successive procedures. In brief, it contains 6 main steps:²⁴ sequencing and bioinformatic analysis, selection of heterologous expression hosts, DNA assembly, refactoring pathways, improved selection of mutants and genome editing.

Conclusion

Natural products (or the semisynthetic/ synthetic derivatives) were widely used in medicine for the past decades. In the genomics era, with the growing development of genomics and metagenomics, natural products has regained the spotlight for drug discovery due to their huge chemical structure diversity and their biodiversity. I believe that more novel bioactive chemical entities will be identified through genomics— driven natural product discovery, and drug development pipelines promise to be revitalized by this method in the long run.

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Conflict of interest

The author declares that there is no conflict of interest.

References

- Newman DJ, Cragg GM. Natural Products as Sources of New Drugs from 1981 to 2014. J Nat Prod. 2016;79(3):629–661.
- D'Incalci M, Galmarini CM. A review of trabectedin (ET–743): a unique mechanism of action. *Mol Cancer Ther*. 2010;9(8):2157–2163.
- McBride A, Butler SK. Eribulin mesylate: a novel halichondrin B analogue for the treatment of metastatic breast cancer. *Am J Health Syst Pharm*. 2012;69(9):745–755.
- Huyck TK, Gradishar W, Manuguid F, et al. Eribulin mesylate. Nat Rev Drug Discov. 2011;10(3):173–174.
- Morgan RJ Jr, Leong L, Chow W, et al. Phase II trial of bryostatin–1 in combination with cisplatin in patients with recurrent or persistent epithelial ovarian cancer: a California cancer consortium study. *Invest New Drugs*. 2012;30(2):723–728.
- Seneci P, Miertus S. Combinatorial chemistry and high-throughput screening in drug discovery: different strategies and formats. *Mol Divers*. 2000;5(2):75–89.
- Harvey AL, Edrada–Ebel R, Quinn RJ. The re–emergence of natural products for drug discovery in the genomics era. *Nat Rev Drug Discov*. 2015;14(2):111–129.

- Cimermancic P, Medema MH, Claesen J, et al. Insights into secondary metabolism from a global analysis of prokaryotic biosynthetic gene clusters. Cell. 2014;158(2):412–421.
- 9. Land M, Hauser L, Jun SR, et al. Insights from 20 years of bacterial genome sequencing. *Funct Integr Genomics*. 2015;15(2):141–161.
- Challis GL. Genome mining for novel natural product discovery. J Med Chem. 2008;51(9):2618–2628.
- Zerikly M, Challis GL. Strategies for the discovery of new natural products by genome mining. *Chembiochem*. 2009;10(4):625–633.
- Weber T, Blin K, Duddela S, et al. AntiSMASH 3.0-a comprehensive resource for the genome mining of biosynthetic gene clusters. *Nucleic Acids Res*. 2015;43(W1):W237-243.
- Inaoka T, Ochi K. Scandium stimulates the production of amylase and bacilysin in Bacillus subtilis. Appl Environ Microbiol. 2011;77(22):8181– 8183
- Scherlach K, Schuemann J, Dahse HM, et al. Aspernidine A and B, prenylated isoindolinone alkaloids from the model fungus *Aspergillus nidulans*. J Antibiot (Tokyo). 2010;63(7):375–377.
- Tanaka Y, Hosaka T, Ochi K. Rare earth elements activate the secondary metabolite-biosynthetic gene clusters in Streptomyces coelicolor A₃(2). *J Antibiot (Tokyo)*. 2010;63(8):477–481.
- Rutledge PJ, Challis GL. Discovery of microbial natural products by activation of silent biosynthetic gene clusters. *Nat Rev Microbiol*. 2015;13(8):509–523.
- Ochi K, Hosaka T. New strategies for drug discovery: activation of silent or weakly expressed microbial gene clusters. *Appl Microbiol Biotech*nol. 2013;97(1):87–98.
- Zhang MM, Wang Y, Ang EL, et al. Engineering microbial hosts for production of bacterial natural products. *Nat Prod Rep.* 2016;33(8):963– 987.
- Bergmann S, Schümann J, Scherlach K, et al. Genomics-driven discovery of PKS-NRPS hybrid metabolites from *Aspergillus nidulans*. *Nat Chem Biol*. 2007;3(4):213–217.
- Cichewicz RH. Epigenome manipulation as a pathway to new natural product scaffolds and their congeners. Nat Prod Rep. 2010;27(1):11–22.
- Shao Z, Zhao H, Zhao H. DNA assembler, an *in vivo* genetic method for rapid construction of biochemical pathways. *Nucleic Acids Res*. 2009;37(2):e16.
- Luo Y, Huang H, Liang J, et al. Activation and characterization of a cryptic polycyclic tetramate macrolactam biosynthetic gene cluster. *Nat Commun.* 2013;4:2894.
- Medema MH, Cimermancic P, Sali A, et al. A systematic computational analysis of biosynthetic gene cluster evolution: lessons for engineering biosynthesis. *PloS Comput Biol*. 2014;10(12):e1004016.
- Winn M, Fyans JK, Zhuo Y, et al. Recent advances in engineering nonribosomal peptide assembly lines. *Nat Prod Rep.* 2016;33(2):317–347.
- Weissman KJ. Genetic engineering of modular PKSs: From combinatorial biosynthesis to synthetic biology. Nat Prod Rep. 2016;33(2):203–230.
- Sugimoto Y, Ding L, Ishida K, et al. Rational design of modular polyketide synthases: morphing the aureothin pathway into a luteoreticulin assembly line. *Angew Chem Int Ed Engl.* 2014;53(6):1560–1564.