Comparative Histomorphology of Endocrine Pancreas in Deccani Sheep and Bidri Goat

Abstract
The present study was conducted in six adult Deccani sheep and Bidri goats to know the histological structure of endocrine pancreas. Islets were distributed irregularly within the exocrine portion of the gland. The reticular fibers formed a boundary between the endocrine and exocrine portion. The islets were oval, round or irregular in shape and were classified into small, medium and large, based on their size on its long axis. Three types of cells α, β and δ were identified within the islets by special stains like Masson’s trichrome, Modified aldehyde fuchsia stain method and Maldonado’s method.

Keywords: Histology; Endocrine pancreas; Islets; Sheep; Maldonado’s method; Deccani; Histomorphological

Introduction
Small ruminants pancreas is a major accessory organ and has an important role in digestion. Endocrine part of pancreas is Islets of Langerhans which are highly vascularized, consisting of several endocrine cell types that function together to maintain glucose homeostasis. There has been limited scientific work done on the histomorphological study of pancreas in Deccani sheep Bidri goat. The present research was conducted to analyze and compare the general histomorphology of endocrine pancreas in both the species. It may also useful in future for transplantation of pancreatic cells, stem cell regeneration, tissue engineering and in vitro studies. Therefore, for better understanding of pancreatic histomorphology in humans. Hence the present research work was undertaken.

Material and Methods
The present study was carried out in the Department of Veterinary anatomy and Histology, Veterinary College, Bidar, Karnataka. The materials for the study were collected from six adult Deccani sheep and Bidri goat, immediately after slaughter from local slaughter houses. The collected tissue pieces were washed in normal saline later, they were fixed in different fixatives like normal buffered formalin, bouin’s solution and zenker’s solution. The tissue pieces were processed and embedded in paraffin by routine method. Sections were cut at 4-6 µm thickness and were utilized for histomorphological studies.

Results and Discussion
There was no particular pattern of distribution of islets of Langerhans in different parts of the gland in both Deccani sheep and Bidri goat. Similar findings were reported by Meshram [1] in goat. However, Baltazar et al. [2] reported that islets were more concentrated in the right lobe than in the left lobe and body of pancreas in Philippine carabao.

Islets were round, oval or irregular in shape and were present irregularly within the exocrine portion and also in the interlobular connective tissue (Figure 1 & 2). These results were in agreement with findings of Malik et al. [3] in cattle and buffalo, Banks [4] in domestic animals and Ladukar et al. [5] in buffalo. Pancreatic islets were partially encapsulated by few reticular fibers in both species. These findings were in accordance with the observations of Gangul et al. [6] in goat and Trautmann et al. [7] in mammals. However, the number of reticular fibers was more in Deccani sheep than in Bidri goat. Some of the small islets contained only one, two or three cells in both species which was in agreement with Mukherjee et al. [8] who reported unicellular islets in sheep (Figure 3 & 4).

Figure 1: Photomicrograph of pancreas of adult Bidri goat showing islets with different shape and sizes (arrow) within the exocrine portion (a). H & E ×4.
In the present study small, medium and large islets were observed in both species and the length of islets was measured in its long axis. Statistically there was no significant difference in the length of islets in both Bidri goat and Deccani sheep. Small islets ranged from 16.05µm to 48.58µm in Bidri goat and 10.26µm to 45.61µm in Deccani sheep. Medium size islets ranged from 49.98µm to 87.11µm in Bidri goat and 49.21µm to 88.88µm in Deccani sheep. Large islets ranged from 91.75µm to 164.44µm in Bidri goat and 90.70µm to 162.83µm in Deccani sheep. These findings were contrary to the findings of Vijayaraghavan et al. [9] who reported that small islets ranged from 17.06µm to 70.30µm and large islets from 98.42µm to 168.72µm on longitudinal axis in buffalo.

Three types of cells viz; alpha, beta and delta cells were observed within the islets of Langerhans in both Deccani sheep and Bidri goat (Figure 5 & 6). Similar observations were reported by Dhoolappa [10] in Indian donkey and Eurell et al. [11] in domestic animals who also observed alpha, beta and delta cells. However, Mukherjee et al. [8], Bacha et al. [12] in sheep, Gangul et al. [6] in goat; Ladukar et al. [13] and Vijayaraghavan et al. [9] in buffalo reported only two types of cells.

Alpha cells were placed peripherally, while beta cells towards the centre of islets. Similar observations were reported by Monroe et al. [14] in dog and man and Ladukar et al. [13] in buffalo. However, Frossmann [15] in equine and Dhoolappa [10] in Indian donkey reported alpha cells towards centre and beta cells towards the periphery. This variation may be attributing to species variation. Beta cells were purple - violet, alpha cells were

**Figure 2**: Photomicrograph of pancreas of adult Deccani sheep showing interlobular duct (a), goblet cell (b), islet (c) within the interlobular connective tissue (d) Gomori’s Aldehyde Fuchsin stain × 40.

**Figure 3**: Photomicrograph of pancreas of adult Deccani sheep showing reticular fibres (arrow) around the islet. Gomori’s reticulum stain × 100.

**Figure 4**: Photomicrograph of pancreas of adult Deccani sheep showing only two, three cells in a pancreatic islets (arrow).

**Figure 5**: Photomicrograph of pancreas in Bidri goat showing alpha cells (a), beta cells (b) and delta cells (c) Gomori’s method for pancreatic islets × 100.

**Figure 6**: Photomicrograph of pancreas in Deccani sheep showing alpha cells (a), beta cells (b) and Delta cell (c) modified Aldehyde fuchsin stain × 40.

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yellow and delta cells appeared green with modified aldehyde fuchsin stain (Figure 6).

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References