

Involvement of innate and adaptive immunity in parkinson's disease

Abstract

Central nervous system (CNS) was considered as an “immunologically privileged site”. However, accumulating evidence supports a role for neuro inflammation in progress in Parkinson's disease (PD). Not only the activated resident microglia in brains, cytokine levels in CNS and blood, the presence of auto antibodies, and the infiltration of T-cell in CNS also contribute disease progression. The interplay between innate and adaptive immunity in the pathobiology of PD will be focused on this article.

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Innate immunity in PD

The presence of HLA-DR (human MHC class II cell surface receptor) positive activated microglia in Parkinson's disease (PD) patients' brain including Substantia Nigra (SNs) has been described.¹⁻³ In vivo imaging using positron emission tomography (PET) suggests widespread activated microglia in PD patients.⁴ Moreover, HLA-DQ as well as HLA-DR, both expressed by monocytes in the CSF and peripheral blood of PD patients are significantly higher compared with controls.⁵ More recently, genome-wide association studies (GWASs) of PD patients,⁶⁻¹⁰ including a meta-analysis of the GWASs,¹¹ verified an increased relative risk for PD and expression of HLA-DR or HLA-DQ MHC II molecules, leading to the designation of HLA-DRA as PARK18, a genetic marker recently found to be associated with susceptibility to PD.⁶ Activated microglia and monocytes in PD brains secrete proinflammatory neurotoxic cytokines. Indeed, levels of IL-1 β , IL-6, and TNF- α are elevated in the CSF of PD patients.^{11,12} Increased expression of NF κ B (nuclear factor κ B) in the SN of PD patients is found in CD11b+ microglia and also in affected neurons.¹³ These data support the hypothesis that activation of cells of the innate immune system, such as microglia and monocytes, directly contribute to the pathobiology of PD.

It is likely that α -synuclein (α Syn) associated pathology modulates the microglia response as α Syn deposition correlates with the presence of MHC II expressing microglia.¹⁴ Nitrated α Syn within Lewy bodies, released from dying or dead dopaminergic neurons, was reported to induce microglia activation.¹⁵ In addition, α Syn-induced microglia activation was mediated through PRRs binding.¹⁶ Although α Syn is a typical cytosolic protein, it has also been found not only in CSF but also blood. Changes in the levels and characteristics of extracellular α Syn are associated with the disease and extracellular α Syn has been shown to be taken up by cultured microglia as well as neuron.¹⁷ Moreover, presence of abnormal α Syn expression in cells surrounding neuroinflammatory lesions was reported within the brains of patients with multiple sclerosis.¹⁸ Therefore, neurotoxicity related to accumulation of α Syn in PD may occur through an excessive microglia stimulation.

A harmful role of reactive microglia has also been found in several PD animal models. Behavioral changes or dopaminergic neurodegeneration, which are caused by neurotoxins MPTP and 6-OHDA, are associated with microglial activation and increased production of proinflammatory cytokines in the SNc.¹⁹⁻²² MPTP treated mice showed behavioral dysfunction, activated microglia, and increased the levels

of IL-10, IL-12(p40), IL-13, IFN- γ , and MCP-1 in CSF.²² Moreover, both peripheral and intranigral administration of lipopolysaccharide (LPS), a potent microglial activator and a ligand of TLR-4, induces a rapid microglial response and increased levels of pro-inflammatory cytokines and free radicals in the brain, which is followed only at a later time by dopaminergic degeneration.²³⁻²⁷ Finally, several studies have demonstrated a clear relationship between pro-inflammatory cytokines and nigral degeneration; over expression of TNF- α via virus delivery system causes dopaminergic cell death, while deficiency of TNF- α receptor is neuroprotective against MPTP toxicity²⁸⁻²⁹ and IL-1 β over expression exacerbates LPS or 6-OHDA-induced neurodegeneration.³⁰⁻³¹

Adaptive immunity in parkinson disease

Today, accumulating evidence suggested the adaptive immune system also involves PD pathology. Both patients and animal models showed exacerbation of the neurodegenerative process after a peripheral inflammatory stimulus.³² Increasing inflammation and breakdown of the blood-brain barrier (BBB) forces increased communication between the CNS and peripheral immune systems as evidenced in several neurodegenerative diseases with increased leukocyte migration within the brain parenchyma.³³

Along with activated microglia and astrocytes, T cells may also comprise components of PD pathobiology. More recently, both CD4+ and CD8+ T cells have been discovered within the SN of PD patients and MPTP treated mice.³³ Intercellular adhesion molecule-1 (ICAM-1) is known to play a key role in T-cell mediated host defense mechanisms and ICAM1-positive glia are also increased in the SN of PD brains and MPTP treated monkey brain,³⁴ as well as association with lesioned areas of Alzheimer disease, amyotrophic lateral sclerosis, Pick's disease and progressive supranuclear palsy.³⁵

Although several autoantibodies for dopamine neuron antigens are reported in sera and CSF of PD patients,^{36,37} the role of the adaptive immune system has only recently begun to be investigated in depth. IgG from PD patients (PD-IgG) activated microglia via the Fc γ receptor (FcR) and induce dopaminergic cell injury, while PD IgG injection in FcR $^{-/-}$ mice resulted in no significant increase of microglia and no loss of TH-positive cells in the SNpc.³⁸ Accumulating evidence suggested that PD patients showed brain-associated autoantibodies including those directed against, GM1, S100B, glial fibrillary acidic protein (GFAP), NGF, neurofilament, myelin basic protein, tau, A β , and neuronal calcium channels, as well as α -syn and its modified and

fibrillary forms.³⁸⁻⁴⁵ Immunohistochemical staining of tissues from PD patients show that Lewy bodies were strongly immunolabelled with IgG.⁴⁷ In MPTP-intoxicated mice, α -syn drains to cervical lymph nodes where it activates antigen-presenting cells and T cells.⁴⁸ Moreover, antibodies to α -syn and catecholamine-derived melanin (neuromelanin) are increased in PD patients with antineuromelanin immunoglobulin binding shown to be more active in early disease.⁴⁹ Indeed, these data suggest that endogenous antibodies of unknown specificity have the capacity to cross the BBB and bind cognate antigens expressed by dopaminergic neurons.

Conclusion

I have reviewed here the overwhelming evidence that supports a role for neuroinflammation in PD. The number of activated microglia in brains, cytokine levels in CNS and blood, the presence of autoantibodies, and the infiltration of T-cell in CNS suggest that not only the local immune system but the peripheral immune system involve disease progression. However, more evidence is necessary for immunomodulatory strategies in PD treatment.

Acknowledgments

None.

Conflicts of interest

None.

References

- McGeer PL, Itagaki S, Boyes BE, et al. Reactive microglia are positive for HLA-DR in the substantia nigra of Parkinson's and Alzheimer's disease brains. *Neurology*. 1988;38(8):1285-1291.
- Banati RB, Daniel SE, Blunt SB. Glial pathology but absence of apoptotic nigral neurons in long-standing Parkinson's disease. *Mov Disord*. 1998;13(2):221-227.
- Hirsch EC, Hunot S. Neuroinflammation in Parkinson's disease: a target for neuroprotection? *Lancet Neurol*. 2009;8(4):382-397.
- Gerhard A, Pavese N, Hotton G, et al. In vivo imaging of microglial activation with [¹¹C](R)-PK11195 PET in idiopathic Parkinson's disease. *Neurobiol Dis*. 2006;21(2):404-412.
- Fiszer U, Mix E, Fredrikson S, et al. Parkinson's disease and immunological abnormalities: increase of HLA-DR expression on monocytes in cerebrospinal fluid and of CD45RO+ T cells in peripheral blood. *Acta Neurol Scand*. 1994;90(3):160-166.
- Hamza TH, Zabetian CP, Tenesa A, et al. Common genetic variation in the HLA region is associated with late-onset sporadic Parkinson's disease. *Nat Genet*. 2010;42(9):781-785.
- Saiki M, Baker A, Williams-Gray CH, et al. Association of the human leucocyte antigen region with susceptibility to Parkinson's disease. *J Neurol Neurosurg Psychiatry*. 2010;81:890-891.
- Nalls MA, Couper DJ, Tanaka T, et al. Multiple loci are associated with white blood cell phenotypes. *PLoS Genet*. 2011;7(6):e1002113.
- Puschmann A, Verbeeck C, Heckman MG, et al. Human leukocyte antigen variation and Parkinson's disease. *Parkinsonism Relat Disord*. 2011;17(5):376-378.
- Simón-Sánchez J, van Hilten JJ, van de Warrenburg B, et al. Genome-wide association study confirms extant PD risk loci among the Dutch. *Eur J Hum Genet*. 2011;19(6):655-661.
- Blum-Degen D, Müller T, Kuhn W, et al. Interleukin-1 beta and interleukin-6 are elevated in the cerebrospinal fluid of Alzheimer's and de novo Parkinson's disease patients. *Neurosci Lett*. 1995; 202(1-2):17-20.
- González-Scarano F, Baltuch G. Microglia as mediators of inflammatory and degenerative diseases. *Annu Rev Neurosci*. 1999;22:219-240.
- Ghosh A, Roy A, Liu X, et al. Selective inhibition of NF-kappaB activation prevents dopaminergic neuronal loss in a mouse model of Parkinson's disease. *Proc Natl Acad Sci U S A*. 2007;104(47):18754-18759.
- Croisier E, Moran LB, Dexter DT, et al. Microglial inflammation in the parkinsonian substantia nigra: relationship to alpha-synuclein deposition. *J Neuroinflammation*. 2005;2:14.
- Reynolds AD, Stone DK, Mosley RL, et al. Nitrated {alpha}-synuclein-induced alterations in microglial immunity are regulated by CD4+ T cell subsets. *J Immunol*. 2009;182(7):4137-4149.
- Béraud D, Twomey M, Bloom B, et al. α -Synuclein Alters Toll-Like Receptor Expression. *Front Neurosci*. 2011;5:80.
- Bae EJ, Lee HJ, Rockenstein E, et al. Antibody-aided clearance of extracellular α -synuclein prevents cell-to-cell aggregate transmission. *J Neurosci*. 2012;32:13454-13469.
- Lu JQ, Fan Y, Mitha AP, et al. Association of alpha-synuclein immunoreactivity with inflammatory activity in multiple sclerosis lesions. *J Neuropathol Exp Neurol*. 2009;68(2):179-89.
- Kohutnicka M, Lewandowska E, Kurkowska-Jastrzebska I, et al. Microglial and astrocytic involvement in a murine model of Parkinson's disease induced by 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP). *Immunopharmacology*. 2008;39(3):167-180.
- Luchtman DW, Shao D, Song C. Behavior, neurotransmitters and inflammation in three regimens of the MPTP mouse model of Parkinson's disease. *Physiol Behav*. 2009;98(1-2):130-138.
- Schintu N, Frau L, Ibba M, et al. Progressive dopaminergic degeneration in the chronic MPTP mouse model of Parkinson's disease. *Neurotox Res*. 2009;16(2):127-139.
- Yasuda Y, Shimoda T, Uno K, et al. The effects of MPTP on the activation of microglia/astrocytes and cytokine/chemokine levels in different mice strains. *J Neuroimmunol*. 2008;204(1-2):43-51.
- Arai H, Furuya T, Yasuda T, et al. Neurotoxic effects of lipopolysaccharide on nigral dopaminergic neurons are mediated by microglial activation, interleukin-1beta, and expression of caspase-11 in mice. *J Biol Chem*. 2004;279(49):51647-51653.
- Dutta G, Zhang P, Liu B. The lipopolysaccharide Parkinson's disease animal model: mechanistic studies and drug discovery. *Fundam Clin Pharmacol*. 2008;22(5):453-464.
- Herrera AJ, Castaño A, Venero JL, et al. The single intranigral injection of LPS as a new model for studying the selective effects of inflammatory reactions on dopaminergic system. *Neurobiol Dis*. 2000;7(4):429-447.
- Irvani MM, Leung CC, Sadeghian M, et al. The acute and the long-term effects of nigral lipopolysaccharide administration on dopaminergic dysfunction and glial cell activation. *Eur J Neurosci*. 2005;22(5):317-330.
- De Lella Ezcurra AL, Chertoff M, Ferrari C, et al. Chronic expression of low levels of tumor necrosis factor-alpha in the substantia nigra elicits progressive neurodegeneration, delayed motor symptoms and microglia/macrophage activation. *Neurobiol Dis*. 2010;37(3):630-640.
- Sriram K, Matheson JM, Benkovic SA, et al. Deficiency of TNF receptors suppresses microglial activation and alters the susceptibility of brain regions to MPTP-induced neurotoxicity: role of TNF-alpha. *FASEB J*. 2006;20:670-682.
- Ferrari CC, Pott Godoy MC, Tarelli R, et al. Progressive neurodegeneration and motor disabilities induced by chronic expression of IL-1beta in the substantia nigra. *Neurobiol Dis*. 2006;24(1):183-193.
- Long-Smith CM, Collins L, Toulouse A, et al. Interleukin-1 β contributes to dopaminergic neuronal death induced by lipopolysaccharide-stimulated rat glia *in vitro*. *J Neuroimmunol*. 2010;226(1-2):20-26.

31. Pott Godoy MC, Tarelli R, Ferrari CC, Sarchi MI, Pitossi FJ (2008) Central and systemic IL-1 exacerbates neurodegeneration and motor symptoms in a model of Parkinson's disease. *Brain* 131(Pt 7): 1880-1894.
32. Ferrari CC, Tarelli R (2011) Parkinson's disease and systemic inflammation. *Parkinsons Dis* 2011: 436813.
33. Brochard V, Combadière B, Prigent A, Laouar Y, Perrin A, et al. (2009) Infiltration of CD4+ lymphocytes into the brain contributes to neurodegeneration in a mouse model of Parkinson disease. *J Clin Invest* 119(1): 182-192.
34. Miklossy J, Doudet DD, Schwab C, Yu S, Mc Geer EG, et al. (2006) Role of ICAM-1 in persisting inflammation in Parkinson disease and MPTP monkeys. *Exp Neurol* 197: 275-283.
35. Ikeda K, Akiyama H, Kondo H, Ikeda K (1993) Anti-tau-positive glial fibrillary tangles in the brain of postencephalitic parkinsonism of Economo type. *Neurosci Lett* 162(1-2): 176-178.
36. McRae-Degueurce A, Rosengren L, Haglid K, Bööj S, Gottfries CG, et al. (1998) Immunocytochemical investigations on the presence of neuron-specific antibodies in the CSF of Parkinson's disease cases. *Neurochem Res* 13(7): 679-684.
37. Dahlström A, Wigander A, Lundmark K, Gottfries CG, Carvey PM, et al. (1990) Investigations on auto-antibodies in Alzheimer's and Parkinson's diseases, using defined neuronal cultures. *J Neural Transm Suppl* 29: 195-206.
38. He Y, Le WD, Appel SH (2002) Role of Fcγ receptors in nigral cell injury induced by Parkinson disease immunoglobulin injection into mouse substantia nigra. *Exp Neurol* 176(2): 322-327.
39. Elizan TS, Casals J, Yahr MD (1983) Antineurofilament antibodies in postencephalitic and idiopathic Parkinson's disease. *J Neurol Sci* 59(3): 341-347.
40. Karcher D, Federspiel BS, Lowenthal FD, Frank F, Lowenthal A (1986) Anti-neurofilament antibodies in blood of patients with neurological diseases. *Acta Neuropathol* 72(1): 82-85.
41. Appel SH, Smith RG, Alexianu M, Engelhardt J, Mosier D, et al. (1994) Neurodegenerative disease: autoimmunity involving calcium channels. *Ann N Y Acad Sci* 747: 183-194.
42. Terryberry JW, Thor G, Peter JB (1998) Autoantibodies in neurodegenerative diseases: antigen-specific frequencies and intrathecal analysis. *Neurobiol Aging* 19(3): 205-216.
43. Poletaev AB, Morozov SG, Gnedenko BB, Zlunikin VM, Korzhenevsky DA (2000) Serum anti-S100b, anti-GFAP and anti-NGF autoantibodies of IgG class in healthy persons and patients with mental and neurological disorders. *Autoimmunity* 32(1): 33-38.
44. Zappia M, Crescibene L, Bosco D, Arabia G, Nicoletti G, et al. (2002) Anti-GM1 ganglioside antibodies in Parkinson's disease. *Acta Neurol Scand* 106(1): 54-57.
45. Papachroni KK, Ninkina N, Papapanagiotou A, Hadjigeorgiou GM, Xiromerisiou G, et al. (2007) Autoantibodies to alpha-synuclein in inherited Parkinson's disease. *J Neurochem* 101(3): 749-756.
46. Yanamandra K, Gruden MA, Casate V, Meskys R, Forsgren L, et al. (2011) α-synuclein reactive antibodies as diagnostic biomarkers in blood sera of Parkinson's disease patients. *PLoS One* 6(4): e18513.
47. Orr CF, Rowe DB, Mizuno Y, Mori H, Halliday GM (2005) A possible role for humoral immunity in the pathogenesis of Parkinson's disease. *Brain* 128(pt 11): 2665-2674.
48. Benner EJ, Banerjee R, Reynolds AD, Sherman S, Pisarev VM, et al. (2008) Nitrated alpha-synuclein immunity accelerates degeneration of nigral dopaminergic neurons. *PLoS One* 3(1): e1376.
49. Double KL, Rowe DB, Carew-Jones FM, Hayes M, Chan DK, et al. (2009) Anti-melanin antibodies are increased in sera in Parkinson's disease. *Exp Neurol* 217(2): 297-301.