

# Anti Genotoxic Effect of TiO<sub>2</sub> Nanoparticle Biosynthesized from *Sargassum polycystum* - a Marine Macroalgae

## Abstract

Nanotechnology is the application of science and technology to control matter at the molecular level, which is also referred to as the ability for designing, production, characterization and application to structures, devices and systems by controlling shape and size at the nanometer scale. The present study was an attempt to investigate antigenotoxic effect of TiO<sub>2</sub> nanoparticle biosynthesized from *Sargassum polycystum*. Our work aimed at conducting Genotoxicity studies for the screening of Mutagenicity using Ames test and to prove this as an antigenotoxicant. The mutagens (positive controls) treated without metabolic activation system showed a 3 fold increase of average revertant colonies per plate when compared with that of concurrent vehicle controls, thus exhibiting the ability to identify the mutagen by the tester strains. The mutagenicity assay was performed with three dose levels (0.312, 1.25, and 5.0 mg/ml) in the absence of metabolic activation system. Inhibition of background growth of non-revertant bacteria was not found at any of the three dose levels. Furthermore works are to be carried out in future to find the exact mechanism of its genoprotective nature.

**Keywords:** Seaweed; Ames test; Antigenotoxicity; Chromosome aberration

## Clinical Paper

Volume 5 Issue 4 - 2017

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**Received:** April 12, 2017 | **Published:** May 19, 2017

## Introduction

The dramatic expansion in the nanotechnology industry over the last decade has resulted in the development of a myriad of novel materials specifically in the nano-size range (sub-100 nm). These nanomaterials (NM) are promising to revolutionise our life-style as they have unique physico-chemical features encompassing beneficial properties surpassing those of traditional substances. Such features include enhanced electrical or thermal conductivity, more efficient catalysts, high tensile strength (yet lighter weight) or improved drug delivery vehicles. Consequently, NM has potential applications in a wide range of industrial settings in addition to medical healthcare and consumer products [1].

In laboratory tests with Nanoparticles, the bacteria, viruses, and fungi are killed within minutes of contact. The effect of nanoparticles on bacteria is very important, since they constitute the lowest level and hence enter the food chain of ecosystem [2,3]. TiO<sub>2</sub> nanoparticles (TiO<sub>2</sub>-NPs), approximately less than 100 nm in diameter, have become a new generation of advanced materials due to their novel and interesting optical, dielectric, and photo-catalytic properties from size quantization [4]. Titanium dioxide (TiO<sub>2</sub>) is a photo catalyst and widely utilized as a self-cleaning and self-disinfecting material for surface coating in many applications. Titanium dioxide has a more helpful role in our environmental purification due to its nontoxicity, photo induced super-hydrophobicity and antifogging effect [5]. The biomedical applications of TiO<sub>2</sub>, which includes the photodynamic therapy for cancer treatment, drug delivery systems, cell imaging, biosensors for biological assay, and genetic engineering. The major advantage

of TiO<sub>2</sub> is that it is retained for a longer time within biological systems and they are nontoxic and stable without light irradiation.

Genotoxicity describes the property of chemical agents that damages the genetic information within a cell causing mutations, which may lead to cancer. Genotoxicity studies is to investigate the potency of a compound to interact with the genetic constitution. A modification of the genetic code via gene mutations (point mutations) or DNA strand breaks may lead to fundamental changes in the behaviour of bacteria and cells. In evolution these changes can be of advantage for the species, or these changes may cause diseases in vertebrates. Genetic toxicity testing is routinely performed to identify potential genotoxic carcinogens and germ cell mutagens [6]. Exposure to genotoxic chemicals that are present in food, environment and also in medical treatments can alter the genetic material permanently and thus may lead to cancer. Antigenotoxic compounds from medicinal and aromatic plants are used over several years. The studies of this type are aimed at understanding the protective mechanisms which may be relevant for the primary prevention of cancer and other mutation related diseases. Plant-derived phenolic compounds exert anti genotoxic property. Antigenotoxic plant extracts can encounter or prevent the adverse effects caused by DNA damaging chemicals. This is due to the fact that dietary intake of phytochemicals has shown protective effect against the dreaded disease. Since many of the marine phyto-resources derived bioactive compounds can combat such mutagenic effect, screening of antigenotoxic potential of marine macroalgae is now gaining importance. *Sargassum* species are found throughout tropical and subtropical areas of the world

and are reported to produce metabolites of structural classes such as terpenoids, polysaccharides, polyphenols, sargaquinoic acids, sargachromenol, plastoquinones, steroids, glycerides etc., which possesses several therapeutic activities. As it possesses many pharmacological properties, it has been considered as a medicinal food of the twenty-first century, and research is being carried out on it to reveal its other pharmacological properties.

The common edible brown seaweed *Sargassum polycystum* (C. Agardh) (SP) reportedly alleviated hyperglycaemia and dislipidemia in diabetic rats [7], possibly due to its good antioxidant and free radical scavenging properties [8]. SP is reportedly used for eczema, scabies, and psoriasis, ulcer and lung diseases, renal dysfunction, viral hepatitis and heart ailments and to promote bile secretion [9], besides having antilipidemic, antioxidant and membrane stabilizing properties [10,11]. SP also has drug metabolizing enzymes protective effects, prevents TNF- $\alpha$  elevation [12], inhibits lipid peroxidation, and preserves hepatic antioxidant defence system *in vivo* [13,14]. It was reported to be hepatoprotective under high-fat/highcholesterol diet [15].

However, Antigenotoxic potential of this species was yet to be explored. Hence, the nontoxic and highly stable characteristic of TiO<sub>2</sub> nanoparticle mooted us to investigate antigenotoxic effect of TiO<sub>2</sub> nanoparticle biosynthesized from *Sargassum polycystum*.

## Materials and Methods

The species of brown algae *Sargassum polycystum* were collected from Mandapam, South Coast of Tamilnadu, India. The algal materials were washed thoroughly with distilled water to remove debris and other associated biota. The samples were shade dried, powdered and stored at 4°C for further use. The dried algae of *S. polycystum* were washed several times with distilled water to remove the waste materials. 5 g of thoroughly washed and boiled in 100 mL double-distilled water for 15 min. Filtered algae extract was stored at -15°C for further use, being usable for several weeks

### Green synthesis of TiO<sub>2</sub>

Titanium tetra iso propoxide [1.5N] was dissolved in 100 ml of distilled water for synthesis the TiO<sub>2</sub> nanoparticles. The algal extract is added drop wise under constant stirring in the ratio of 1:10 [v/v]. The mixture was subjected to constant stirring for 24 hours continuously at room temperature using orbital shaker. In this process formation of nanoparticles were observed by the formation of tiny coarse particles. The biosynthesized nanoparticles were filtered using what man filter paper and washed with distilled water repeatedly to remove the by-products. The obtained wet nanoparticles were dried at 80°C overnight for the removal of moisture.

### Characterization of synthesized TiO<sub>2</sub> nanoparticles from *S. polycystum*

The initial spectral characterization of the biosynthesized TiO<sub>2</sub> NP was carried out after 24hrs between 300 nm to 800 nm using Perkin-Elmer lambda 25 UV-Vis spectrometer. The absorbance was taken after 5-fold diluting the sample with distilled water against distilled water as blank. FTIR was used to identify the

possible functional groups responsible for the reduction of the metal ions and capping of the bio-reduced TiO<sub>2</sub> nanoparticles. The samples were analyzed on a ABB Horizon MB 3000 spectrum instrument in the diffuse reflectance mode operation with the scanning range of 4000-400cm<sup>-1</sup> at a resolution of 0.7 cm<sup>-1</sup> utilising the DTGS [Deuterium Triglycane sulphate] detector. In order to obtain good signal/noise ratio 512scans were recorded. The peaks obtained were plotted as % transmittance in Y axis and wave number (cm<sup>-1</sup>) in x axis. Combined with the intuitive Horizon MBTM FTIR software, the MB3000 will facilitate easy acquisition, processing and analysis of samples. Scanning Electron Microscopy is done for revealing the surface morphology of particles. Structural studies of TiO<sub>2</sub> NPs were done by AURIGA- Cross beam FESEM (M/s Carl Zeiss, Germany). The characterization of the purified nanoparticles were conducted with an XRD 6000 X-ray diffractometry (shimadzu, Japan) operated at voltage of 40 kV and current of 30 mA with cu k radiation in  $\theta$  2 $\theta$  configurations. The crystallite domain size was calculated from the width of the XRD peaks by assuming that they were free from non uniform strains and using the sherrer formula:

$$D = \frac{0.94}{\beta \cos \theta}$$

Where the D is the average crystalline domain size perpendicular to the reflecting planes,  $\lambda$  is the X-ray wave length,  $\beta$  is the full width at of maximum (FWHM) and  $\theta$  is the diffraction angle. To eliminate the additional instrumental broadening, the FWHM was corrected using FWHM from a large grained Si sample.  $\beta$  corrected = (FWHM<sup>2</sup> sample - FWHM<sup>2</sup>n)<sup>1/2</sup> This modified formula is valid only when the crystallite size is smaller than 100 nm.

### Ames test

Cells were treated with various concentrations of Ti-NPs as described in the toxicological study. Mutagenicity was assessed by the pre-incubation assay as described by Maron and Ames [16]. Briefly, 100  $\mu$ L of overnight cultures (1-2  $\times$  10<sup>8</sup> cfu/mL) of strain TA98 were treated separately for 30 minutes at 37°C [8] with various concentrations of Ti-NPs in the absence or presence of S9 mix. The Ti-NPs were suspended in sterile DW and were used at a final concentration of less than 5% (v/v). For the Ames test, the controls and Ti-NP-treated cells were mixed with 2 mL of sterile top agar (0.6% agar and 0.5% NaCl containing 0.5 mM histidine and 0.5 mM biotin) and poured onto minimal glucose agar plates (1  $\times$  Vogel-Bonner salts (0.2 g/L magnesium sulfate, 2 g/L citric acid monohydrate, 10 g/L dipotassium hydrogen phosphate, and 3.5 g/L sodium ammonium phosphate), 2% glucose, and 1.5% agar). The plates were then incubated at 37°C for 48 hours, after which revertants and surviving colonies were counted. Three independent experiments were conducted and each experiment consisted of three replicate plates for each treatment. The positive control used in the presence of S9 mix was 2-aminofluorene.

### Plating procedures

These procedures were used in the dose range-finding and mutagenicity assays. Each plate was labeled with the test item, test phase, tester strain, activation condition, and dose. Treatments in

the absence of S9 were performed by adding 100 µl tester strain and 100 µl test or control article to 2.5 ml molten diluted top agar (maintained at 45 ± 2°C). The mixtures were vortexed and overlaid onto the surface of bottom agar dishes. After the overlay solidifies; the plates were inverted and incubated for 72 hrs at 37 ± 2°C. After incubation the plates were evaluated for the condition of the background lawn for the evidence of cytotoxicity and test item precipitate in comparison with the control and the plates were evaluated for the number of revertant colonies.

### Human lymphocytes culture and chromosomal aberration studies

Heparinized blood samples (0.5 ml) was collected from healthy individuals and were placed in sterile culture flasks with 0.7 ml of RPMI1640 supplemented with fetal bovine serum (1.5 ml), antibiotic-antimycotic mixture (1.0 ml), phyto haemagglutinin (0.1 ml). The cultures are placed in incubator at 37°C for 24 h. After 48 h incubation, test compound (biosynthesis TiO<sub>2</sub>) / positive mutagen of known concentration prepared in suitable vehicle or as such, will be added to the culture at a volume of 0.1 ml to achieve desired final concentration after all other constituents, if any, are added. Negative control cultures received vehicle-DMSO alone at a volume of 0.1 ml. Negative control culture without metabolic activation will receive only 0.5ml of phosphate buffer. The vials/centrifuge tube/flasks will be transferred to CO<sub>2</sub> incubator. The culture will be incubated at 37 ± 1°C and 5% CO<sub>2</sub> for 3-6 h. flasks were transferred to labeled sterile 15 ml centrifuge tubes, centrifuged at room temperature at 1600 rpm for 5-10 min, supernatant will be aspirated gently and to the pellet a freshly made working growth medium RPMI 1640 without PHA-M were added. The total volume of the culture will be made up to 10 ml using the culture medium. The centrifuge tubes were transferred to CO<sub>2</sub> incubator. The culture will be incubated at 37 ± 1°C and 5% CO<sub>2</sub> for 18-21 h. 100µl of 10µg/ml of colchicine and incubate for additional 2 hours. The entire content of the flask was transferred to a sterile centrifuge tube and centrifuged at 800-1000rpm for 10 minutes. The supernatant was discarded and the pellet was suspended in 5ml of hypotonic 0.075M KCl solution and incubated in a water bath at 37°C for 15-20 minutes. The equal amount of freshly prepared ice cold fixative were added (Acetic acid: methanol, 1:3 parts). The cells were centrifuged at 800-1000rpm for 10min. The fixative was removed by centrifugation and this process is repeated twice. The slides were prepared and they were stained with 3 % Giemsa stain solution in phosphate buffer (pH 6.8) for 15 min. At least 300 metaphases were scored in each slide for examining different types of abnormality according to standard protocol of Savage [17]. Mitotic index (MI) was calculated by using formula, MI = number of dividing cells/ total number of cells × 100, where MI, Mitotic index.

### Statistical analysis

The Mean and standard deviation was calculated for each parameter. The data was analyzed by 'SPSS 17.0' software. Two ways ANOVA was performed to determine significance of treatment. The mean separation was performed according to Duncan's New multiple range test (P ≤ 0.05).

## Result and Discussion

The use of plant based natural products as chemopreventive agents is drawing a lot of attention and considered to be practically beneficial in certain cell/tissue based systems and animal model systems. It is necessary to provide scientific proof to justify the use of a plant or its active principles for medicinal purposes [18]. Modern drugs, plants and plant extracts must be characterized after their pharmacological screening for their pharmacokinetic and pharmacodynamic properties, including toxicity. A large number of potential chemopreventive agents have been identified, and they function by mechanisms directed at all major stages of carcinogenesis [19]. The absorption or reflectance in the visible range directly affects the perceived color of the chemicals involved. In this region of the electromagnetic spectrum, molecules undergo electronic transitions. It is based on the principle that molecules containing π-electrons or non-bonding electrons (n-electrons) can absorb the energy in the form of ultraviolet or visible light to excite these electrons to higher anti-bonding molecular orbitals.

Characterization process of TiO<sub>2</sub> NPs includes UV-Vis absorbance measurement. TiO<sub>2</sub> NP dispersed in lake water gave a near-UV absorbance peak between 280 to 336nm. The dynamic aggregation process of TiO<sub>2</sub> NPs was monitored using a UV-Vis spectrophotometer at 378 nm, analyzing the sedimentation of the NPs in different water matrices. The metal particles were observed to be stable in solution even 4 weeks after their synthesis. By stability, we mean that there was no observable variation in the optical properties of the nanoparticle solutions with time.

FTIR measurement was carried out for functional molecules or constituents in the *S. polycystum* responsible for the reduction of TiO<sub>2</sub> ions to nanoparticles and stabilizing the nanoparticles. The TiO<sub>2</sub> nanoparticles synthesized from *S. polycystum* exhibit a lot of biomolecules which were involved in the synthesis of TiO<sub>2</sub> process (Figure 1 & 2). FTIR shows the presence of different functional groups, which give rise to the well-known signatures in the IR region of electromagnetic spectrum. The strong and broad band observed at 3,750 cm<sup>-1</sup> indicates the presence of polyphenolic O-H group and primary amine O-H band, C-H stretching vibrations of alkanes group was formed at the absorption band at 2,230 cm<sup>-1</sup>, a narrow band at 1,638 cm<sup>-1</sup> indicates the presence of amide I, 1,418 cm<sup>-1</sup> corresponding to C-C stretching aromatic ring, C-O stretching carboxylic acid group assigned at 1,243 cm<sup>-1</sup>, the narrow peak at 1,030 cm<sup>-1</sup> is because of the presence of C-N stretching vibrations of aliphatic amines of proteins and 813 cm<sup>-1</sup> is assigned to S-O stretching of sulfonates. The weak bands at 639 and 586 cm<sup>-1</sup> correspond to alkyl halides. This study also confirms that the carbonyl group from amino acids or proteins has stronger ability to bind metal so that the proteins or enzymes could most possibly cap the metal nanoparticles to prevent the agglomeration of the particles.

Scanning Electron Microscopy is done for revealing the surface morphology of particles. Here, the bead for the SEM analysis was prepared by placing a drop of the nanoparticle suspension on the carbon tape attached to the head of cylindrical bead



and it was dried inside a vacuum dryer for a couple of hours. The particles on the top of the bead were scanned by Scanning Electron Microscope and the following image was obtained. The grain shape and surface morphology as investigated by the SEM is shown in Figure 2. The TiO<sub>2</sub> nanoparticles were showing irregular particle structure.

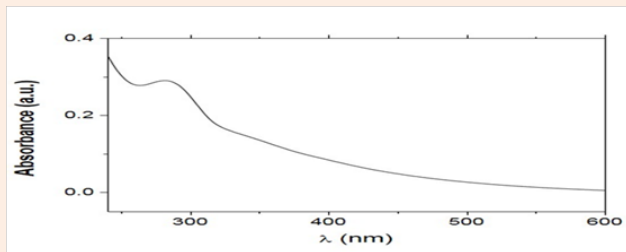


Figure 1: The absorbance spectrum of TiO<sub>2</sub> nanoparticles showing maximum absorbance near 288nm.

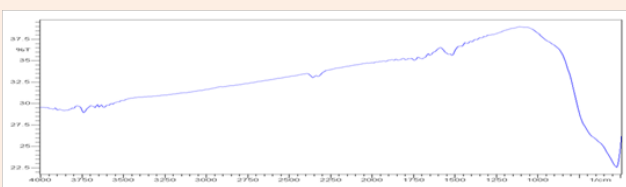


Figure 2: The FTIR absorbance spectrum of TiO<sub>2</sub> NP Biosynthesized Using *S. Polycystum*.

Our work aimed at conducting Genotoxicity studies for the screening of Mutagenicity using Ames test and to prove this as a precursor of drug. The mutagens (positive controls) treated without metabolic activation system showed a 3 fold increase of average revertant colonies per plate when compared with that of concurrent vehicle controls, thus exhibiting the ability to identify the mutagen by the tester strains (Figure 3).

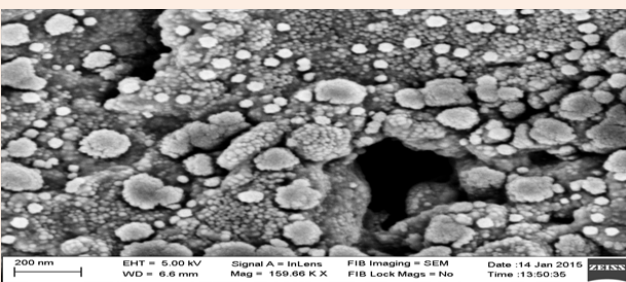


Figure 3: SEM image showing surface morphology of the TiO<sub>2</sub> NP Biosynthesized Using *S. Polycystum*.

The mutagenicity assay was performed with three dose levels (0.312, 1.25, and 5.0 mg/ml) in the absence of metabolic activation system. Inhibition of background growth of non-revertant bacteria was not found at any of the three dose levels.

The study data represented in Table 1 showed no significant increase of His<sup>+</sup> revertant colonies when exposed to the biosynthesized TiO<sub>2</sub> NPs at any of the dose levels incubated with any of the tester strains, without S9 addition, when compared to

the respective controls in the mutagenicity assay. The average revertant colonies per plate treated with the control in the absence of metabolic activation system were found to be within the acceptance limits of the spontaneous revertant control values of respective *Salmonella* strains. The results showed no significant increase in the His<sup>+</sup> revertant colonies following exposure to the samples at any tested concentration in any of the tester strains without S9 when compared with the negative control of each tester strain. Based on the above results, it is concluded that the synthesised nanoparticles were non-mutagenic at the dose levels ranging from 0.313 to 5000 µg/plate by the Ames bacterial reverse mutation assay in the absence of S9 mix, under the conditions of the test employed.

Table 1: Mean Colony Count - Strain TA 98 spontaneous mutation.

Test Item	Test Concentration	Histidine Revertant Colonies
	(µg/plate) / (mg/ml)	CFU/Plate
Positive Control	Sodium azide (1.5 µg/plate)	432
TiO <sub>2</sub> NP Biosynthesised Using <i>S. polycystum</i> (without S9)	5	84
	1.25	60
	0.312	72

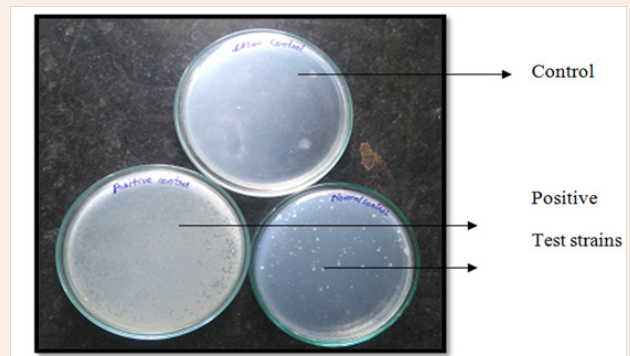


Figure 4: Normal, Positive control, DMSO for TA 98 (Set 1).

When human leukocytes were treated with synthesis nanoparticle of *S. polycystum* alone at different doses; the incidence of cells having aberrants (including gap) in percentage of chromosomal aberration frequency maximum concentration of dose of 1.25 and 5 mg/ml no increase in number of CA were observed when compared to untreated. In contrast, the incidence of aberrant cells in each positive control (Mitomycin C) group increased greatly as compared with each solvent group (P<0.001) Table 2.

Nanomedicine can be broadly defined as a technology which uses molecular tools and knowledge about human body for medical diagnosis and treatment [20]. Here the nanomaterial plays the pivotal therapeutic role because of their ability to cross biological barriers. Furthermore they can stimulate self-healing cell responses and bio compatibility of implants [21]. Moreover

the developmental costs and risks of these materials are low as compared to other drugs [22]. Ti-NPs of different sizes, shapes and material properties have many applications in various areas such as in industrial, medical and cosmetic fields. However, there

is no current data on the characteristics of Ti-NPs such as size and shape in the workplace atmosphere. Many researchers have suggested that size is a critical factor for nanoparticle-induced toxicity and biological responses [23].

**Table 2:** Cytogenetic Assay of *S. polycystum* - Aberration Summary without S9.

Dose (mg/ml Culture)	Total Number of Cells Scored	Percent Numerical Aberration	Mean of Structural Aberration	Total Number of Aberration	Total no. of Cells with Aberration	No. of Aberration per Cell	Aberration Frequency (%)
Negative control (DMSO)	100	0	0	0	0	0	0
0.312	100	0	0	0	0	0	0
	100	0	0	0	0	0	
1.25	100	0	0	79	6	13.17	10.5
	100	0	0	73	9	8.11	
5	100	0	0	100	11	9.09	20
	100	0	0	101	18	5.61	
Positive control	50	0	0	169	29	5.82	45.5
Mitomycin C	50	0	0	151	33	4.58	

**Note:** The results are average of three sets of experiments. a- P<0.05 compared to untreated, b- P<0.05 compared to Mytomycin.

Li et al. [24] also reported that human lymphoblastoid cells exposed to 5 nm Ag-NPs showed a dose dependent increase in MN frequency. Furthermore, Foldbjerg et al. [25] reported that DNA damage was detected as an increase in bulky DNA adducts by 32P post-labeling in human alveolar cells exposed to 120-150 nm Ag-NPs. On the contrary, no significant genotoxic responses were observed in an Ames test (5 nm) [26], a mouse lymphoma assay (< 100 nm) [27] and an *in vivo* MN assay (60 nm) [28]. The Ames test is generally used as a first screening method to assess chemical genotoxicity. In the database published by Kirkland et al. [29], 542 had published Ames results out of 756 rodent carcinogens. Although Ames test has proven to be invaluable in the safety testing of chemical substances, it has been less commonly used with nanoparticles. Doak et al. [30] noted that 17 studies reported negative mutagenicity among the published 19 Ames test studies. In this study, the Ag-NPs were negative in the Ames test with or without S9 mix and this result are consistent with previous reports that indicated Ames tests on nanoparticles were predominantly negative. Many reasons suggested by numerous studies support that Ames test does not appear to be suitable for the assessment of nanoparticles.

### Conclusion

Nanomedicine is the phenomena which uses nanostructured or nanoscale products [size range up to 1,000 nm] in medicine which have some unique medicinal properties based upon their structure. And the biological way of synthesising nanoparticles has potential applications like cost effectiveness, eco-friendly and compatibility for large scale production. In the present study the TiO<sub>2</sub> nanoparticles biosynthesized from marine alagae extract has proved to be a powerful antigenotoxicant which is primarily due to the combined role of phenolics and flavonoids of *S. Polysystem*

that initiated formation of titanium nanoparticles. However our studies are preliminary and work has to be carried out in future to find the exact mechanism of genoprotecting activity of these biosynthesized metal oxide nanoparticles.

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