

Review on Testosterone Delivery by Natural and Synthetic Nanoparticles

Abstract

The loading efficacies of testosterone with polyamidoamine PAMAM-G3 and PAMAM-G4 and chitosan-15 and chitosan-100 kDa nanoparticles were compared in aqueous solution at pH 7.4. The results of multiple spectroscopic methods, transmission electron microscopy (TEM) and molecular modeling were used to characterize the testosterone binding process to polymer nanoparticles. Structural analysis showed testosterone-polymer bindings occur via hydrophobic, H-bonding contacts. The binding affinity is testosterone-chitosan > testosterone-PAMAM. Transmission electron microscopy showed significant changes in carrier morphology with major changes in the diameter of the polymer aggregates as steroid loading occurred. Chitosan nanoparticles are more effective carriers than PAMAM dendrimers.

Keywords: Testosterone; Chitosan; Dendrimer; Delivery; TEM; Modeling; Synthetic polymers; Dendrimers; A Family of cationic polymers; Filter paper; Surface groups; Water solubility; Drug delivery; Spectroscopic; Biological properties; Intensity

Review Article

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Abbreviations: Ch: Chitosan; PAMAM: Poly(Amidoamine); FTIR: Fourier Transform Infrared; TEM: Transmission Electron Microscopy

Introduction

Natural and synthetic polymers are widely used as drug delivery tools in pharmaceutical and nanomedicine biotechnology [1-5]. Among synthetic polymers, dendrimers, a family of cationic polymers, are promising nonviral vectors for gene and drug delivery because of a well-defined molecular shape, controlled chemical structure, high water solubility, large number of chemically versatile surface groups, and unique architecture [6-8]. Natural cationic polymers such as chitosan are attractive candidates for therapeutic applications as they generally are non-toxic, being derived from renewable resources [9]. Biodegradable, biocompatible and nontoxic chitosan nanoparticles are of a major interest in drug delivery systems. They form complexes with antibiotics, anticancer drugs and therapeutic proteins [10-12]. Chitosan and its derivatives have the desired properties for safe use as pharmaceutical drug delivery tools. This has prompted accelerated research activities worldwide on chitosan nanoparticles as drug delivery tools [13-16].

Testosterone is the main androgenic hormone which controls many physiological processes such as, sexual functions and secondary sex characteristics, muscle protein metabolism, plasma lipid and bone metabolism [17]. Testosterone can also be used as a natural template to construct semi-synthetic analogues such as dimers, testosterone amide derivatives, as well as testosterone-cytotoxic hybrid molecules [18-22]. These novel semi-synthetic molecules can be tested for their biological properties, using

diverse spectroscopic methods [18-25]. Synthetic polymers are used as potential nanocarriers to deliver steroids in vitro and in vivo [26-28]. Furthermore, chitosan and its derivatives were also tested as delivery tools for transporting steroids [29,30].

In this review, the binding efficacies of testosterone with PAMAM dendrimers and chitosan nanoparticles were compared, using multiple spectroscopic, TEM images and molecular modeling results. Structural analysis regarding testosterone binding process and the effect of steroid-polymer conjugation on polymer morphology as well as the possibility of testosterone delivery by PAMAM and chitosan nanoparticles are discussed here.

Experimental

Transmission electron microscopy

The TEM images were taken using a Philips EM 208S microscope operating at 180 kV. The morphology of the testosterone conjugates with PAMAM dendrimers and chitosan nanoparticles in aqueous solution at pH 7.4 were observed, using transmission electron microscopy. One drop (5-10 μL) of the freshly-prepared mixture [polymer solution (60 μM) + steroid solution (60 μM)] in Tris-HCl buffer ($24 \pm 1^\circ\text{C}$) was deposited onto a glow-discharged carbon-coated electron microscopy grid. The excess liquid was absorbed by a piece of filter paper, and a drop of 2% uranyl acetate negative stain was added before drying at room temperature [28].

UV spectroscopy

The UV-Vis spectra were recorded on a Perkin-Elmer Lambda

spectrophotometer with a slit of 2 nm and scan speed of 400 nm min⁻¹. Quartz cuvettes of 1 cm were used. The absorbance measurements were performed at pH 7.4 by keeping the concentration of polymer constant (60 μM), while increasing testosterone concentrations (1 μM to 60 μM). The binding constants of steroid-polymer conjugates were obtained according to the published method [31].

FTIR Spectroscopic measurements

Infrared spectra were recorded on a FTIR spectrometer (Impact 420 model, Digilab), equipped with deuterated triglycine sulphate (DTGS) detector and KBr beam splitter, using AgBr windows. Solution of testosterone was added drop wise to the polymer solution with constant stirring to ensure the formation of homogeneous solution and to reach the target steroid concentrations of 15, 30 and 60 μM with a final polymer concentration of 60 μM. Spectra were collected after 2 h incubation of testosterone with polymer solution at room temperature, using hydrated films. Interferograms were accumulated over the spectral range 4000-600 cm⁻¹ with a resolution of 2 cm⁻¹ and 100 scans. The difference spectra [(polymer + steroid solution) - (polymer solution)] were generated using water combination band at 2300 cm⁻¹.

Molecular modeling

The PAMAM-G4, chitosan and testosterone structures were generated using Chem Office Ultra 6.0 software suite [32,33]. The drug was then automatically docked to the rough PAMAM-G4 and chitosan structures using ArgusLab 4.0.1 (ArgusLab 4.0.1, Mark A. Thompson, Planaria Software LLC, Seattle, WA, <http://www.arguslab.com>). The docked testosterone-polymer structures were optimized by means of molecular dynamics, using the MM+ force field available in Hyper Chem Pro 7.0. The free binding energies of the optimized steroid-polymer complexes were calculated using the Ascore scoring function provided in the ArgusLab software [13,14,28].

Results and Discussion

TEM images and the morphology of testosterone-polymer aggregates

TEM images showed major changes in the morphological of PAMAM dendrimers and chitosan nanoparticles upon conjugation with testosterone Figure 1. It can be seen that the free PAMAM-G3 and PAMAM-G4 Figure 1A & 1B are spherical crystalline structures, with mean dimensions of 10 ± 2 nm [34-41], while testosterone-PAMAM, present average dimensions of 19 ± 3 nm (Figures 1A, 1B, 1E & 1F) [28]. The results show that the steroid-PAMAM nanoparticles were larger in size than the free PAMAM due to testosterone encapsulation. This size differences could be attributed to expansion of the branches of PAMAM dendrimer to accommodate steroids within the hydrophobic core to reach high loading. It should be noted that the increase in particle size was more pronounce in the case of testosterone-PAMAM-G4 than testosterone-PAMAM-G3 (Figures 1E & 1F). PAMAM-G4 forms more stable conjugates than PAMAM-G3. Similar structural

changes were observed for several drug-PAMAM-conjugates [42,43].

The shapes of the free Ch-15 and Ch-100 kDa alongside with their testosterone conjugates are shown in the TEM images (Figures 1C, 1D, 1G & 1H). TEM micrographs show that free chitosan had a markedly different shape depending on its size; Ch-15 has spherical-shaped, while Ch-100 is needle-shaped with smooth surface and narrow size distribution of about 90 nm [44,45]. Similar differences were observed for AFM images of Ch-15 and Ch-100 kDa where the result is attributed to the degree of polymer aggregation, as chitosan size increased [46]. However, marked differences were observed in the morphology of the testosterone-chitosan aggregates. TEM images clearly showed the appearance of the aggregates of irregular shapes dispersed in solution when Ch-15 conjugates with testosterone (Figures 1G & 1H). In addition, the bound Ch-100 with testosterone showed major changes of the polymer morphological shape (Figure 1H). An increase of the spherical-shaped aggregates can be seen from TEM micrograph, suggesting that the elongated shapes were lost in favor of spherical-shaped in the testosterone-chitosan conjugates (Figures 1G & 1H). The loss of the elongated shape of chitosan nanoparticles after complex formation with testosterone is likely to be the result of the testosterone encapsulation. This is consistent with major particle size increase as encapsulation occurs (Figures 1G & 1H). Testosterone binding to chitosan, which is a linear polysaccharide that has multiple sites of interaction and therefore should be regarded as core-shell system with steroid (core) and chitosan (shell) [47-51].

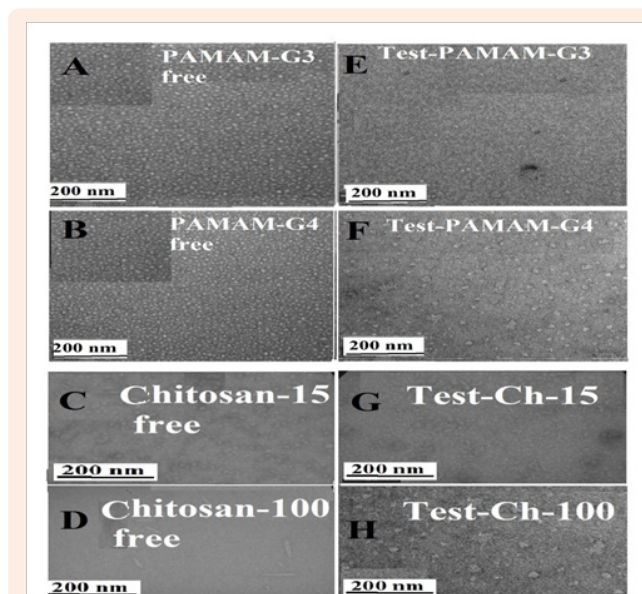


Figure 1: TEM photographs showing the morphology of the free PAMAM-G3 and PAMAM-G4 (A and B) and chitosan-15 and chitosan-100 kDa (C and D) at pH 7.4 and 24°C: in the presence of testosterone (E, F, G and H). The concentrations of steroid and polymer were 60 μM in all samples.

Stability and loading efficacy of testosterone-polymer conjugates by UV spectroscopy

Steroid-polymer binding results in changes in the absorption spectra of the PAMAM and chitosan and the observed changes can be used to calculate the testosterone-polymer binding constants [28,31,52,53]. Steroid-polymer complexation occurred with an increase in the intensity of polymer and testosterone absorption bands around 250 nm (Figure 2 & 3). The steroid-polymer binding constants were calculated as described [31], using plots of $1/(A-A_0)$ vs $(1/\text{steroid concentrations})$ (Figures 2 & 3).

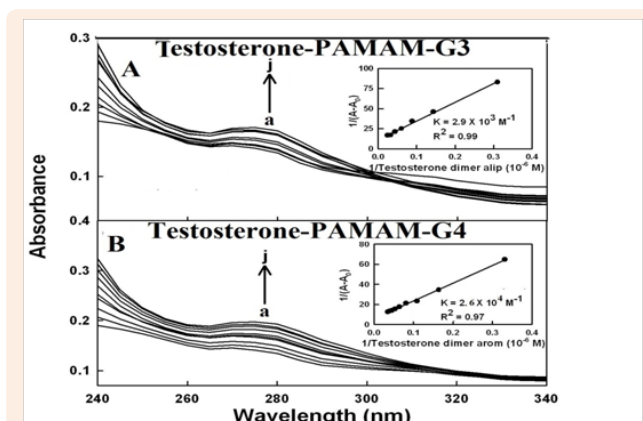


Figure 2: UV-visible spectra of PAMAM and its complexes with testosterone (A) with free PAMAM-G3 and (B) PAMAM-G4 (a) at 60 μM and testosterone (b-j) at 1, 3, 5, 10, 20, 30, 40, 50 and 60 μM . Inset: plot of plot of $1/(A-A_0)$ vs $(1/\text{steroid concentration})$ and binding constant (K) for testosterone-PAMAM conjugates. $1/(A-A_0)$

The double reciprocal plot is linear and gives the overall binding constants for each complex, $K_{\text{testosterone-PAMAM-G3}} = 2.9 (\pm 0.6) \times 10^3 \text{M}^{-1}$, $K_{\text{testosterone-PAMAM-G4}} = 2.6 (\pm 0.3) \times 10^4 \text{M}^{-1}$, $K_{\text{testosterone-chitosan-15}} = 5.0 (\pm 0.6) \times 10^4 \text{M}^{-1}$ and $K_{\text{testosterone-chitosan-100}} = 5.9 (\pm 0.9) \times 10^5 \text{M}^{-1}$ (Figure 2 & 3). The results showed that testosterone forms stronger adduct with chitosan nanoparticles than PAMAM dendrimers (Figures 2 & 3). The extra stability of acid-PAMAM-G4 conjugate is related to the presence of additional terminal charged NH_2 groups (64 amino groups) compared to PAMAM-G3 (32 amino groups), that are involved in testosterone-polymer interactions. Similarly chitosan-100 kDa forms more stable steroid conjugates than chitosan-15 kDa, due to the presence of more charged NH_2 groups associated with larger chitosan nanoparticles.

The loading efficacy for testosterone-polymer conjugates was determined as reported [54].

$$\% \text{ Efficiency} = \frac{\text{Final absorption intensity} - \text{Initial absorption intensity}}{\text{Initial absorption intensity}} \times 100$$

The loading efficacy of testosterone conjugates estimated to be 30% (PAMAM-G3), 40% (PAMAM-G4), 45% (chitosan-15) and 55% (chitosan-100 kDa) in these steroid-polymer conjugates.

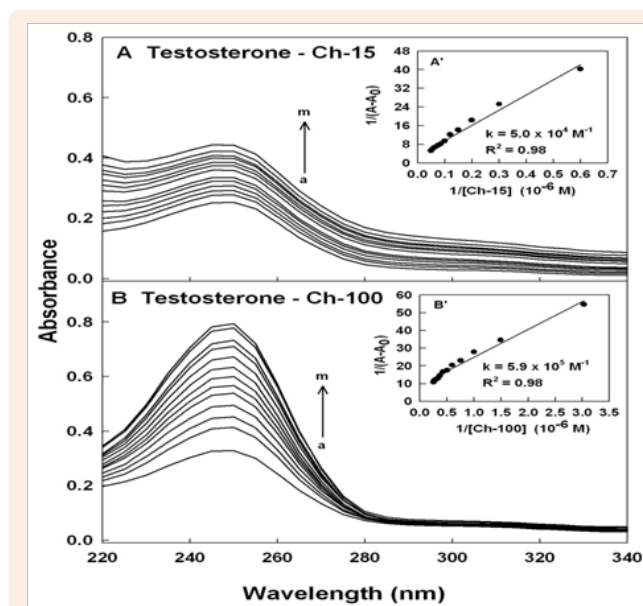


Figure 3: UV-visible spectra of chitosan and its complexes with testosterone (A) chitosan-15 and (B) chitosan-100 kDa (a) with free testosterone at 60 μM and chitosan (b-m) at 1, 3, 5, 10, 20, 30, 40, 50, 60, 70, 80 and 90 μM . Inset: plot of $1/(A-A_0)$ vs $(1/\text{chitosan concentration})$ and the binding constant (K) for testosterone-chitosan conjugates.

Binding analysis of testosterone-polymer conjugates by FTIR spectroscopy

Testosterone-polymer interactions come from infrared results presented in Figure 4. Spectral shifting was observed for the polymer C=O, C-N, C-O stretching and OH and NH bending [55,56], upon testosterone hydrophilic contacts with polymer polar groups. In the free PAMAM-G4 infrared spectrum, the bands at 1650 and 1558 cm^{-1} (OH and NH bending), 1471, 1380, 1303, 1159 and 1060 cm^{-1} (C-O and C-C stretch), exhibited shifting and intensity decreases, upon steroid-polymer complexation (Figure 4A, complexes, 60 μM). The observed spectral shifting was accompanied by a gradual decrease in the intensity of the above vibrational frequencies, in the difference spectra [(polymer solution + steroid solution) - (polymer solution)] of testosterone-polymer conjugates (Figure 4A, diffs 15 μM). As testosterone concentrations increased to 60 mM, major decreases in the intensity of PAMAM vibrational frequencies were observed (Figure 4A, diffs 60 μM). The strong negative features in the difference spectra at 1648, 1554 and 1051 cm^{-1} (testosterone-PAMAM) are due to the loss of intensity of polymer major vibrational bands (Figure 4A, diffs 60 μM). The spectral changes observed are attributed to the hydrophilic interactions of steroid polar groups with polymer OH, NH_2 , C-O and C-N groups. The hydrophilic interaction is more pronounced at high steroid concentrations (Figure 4A, diffs 60 μM). Similarly, chitosan vibrational frequencies at 1638, 1530, 1379, 1151, 1080 and 1035 cm^{-1} exhibited major

changes upon testosterone conjugation (Figure 4B). The positive features observed at 1596-1594, 1498-1488 cm^{-1} and negative features at 1073 and 1074 cm^{-1} , in the difference spectra of testosterone-chitosan complexes are due to the participation of chitosan OH, NH_2 , C-O and C-N groups in testosterone-polymer bindings (Figure 4B, diffs 15 and 60 μM).

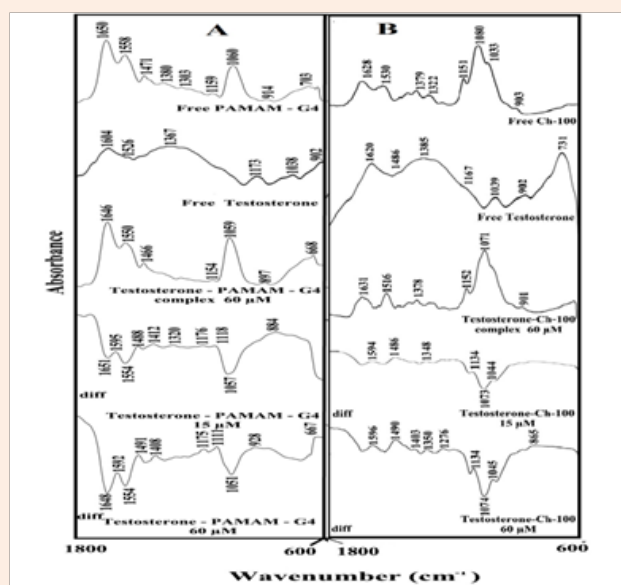


Figure 4: FTIR spectra in the region of 1800-600 cm^{-1} of hydrated films (pH 7.4) for free (A) PAMAM-G4 (60 μM) and (B) chitosan-100 kDa (60 μM) and their testosterone complexes with difference spectra (diff.) (bottom two curves) and obtained at different testosterone concentrations (indicated on the figure).

Docking study

Docking experiments of testosterone to PAMAM-G4 and chitosan nanoparticles revealed the preferred binding sites as shown in Figure 5. The docking results showed testosterone is located in the PAMAM internal cavities surrounded by polymer hydrophobic groups with the free binding energy -9.36 kcal/mol (Figure 5A), while testosterone is bound to the chitosan surface donor groups with the free binding energy -4.44 kcal/mol (Figure 5B). It should be noted that the free binding energy of docking showed more stable testosterone-PAMAM adducts than chitosan conjugates, which is in contrast with the spectroscopic results (Figures 1 & 2).

Concluding Remarks

Testosterone forms stronger conjugate with chitosan nanoparticles than PAMAM dendrimers. Hydrophobic and H-bonding are the main forces in the steroid-polymer conjugation. The stability and loading efficacy of testosterone-polymer conjugates were increased as polymer size increased. Steroid encapsulation alters markedly the size and the shape of PAMAM and chitosan nanoparticles. Chitosan nanoparticles are more effective carriers for testosterone than PAMAM dendrimers.

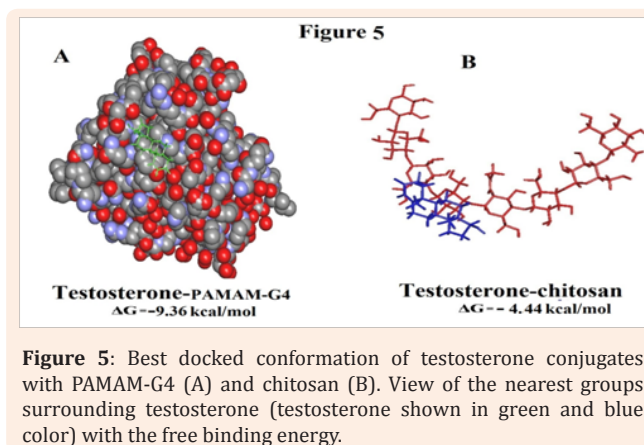


Figure 5: Best docked conformation of testosterone conjugates with PAMAM-G4 (A) and chitosan (B). View of the nearest groups surrounding testosterone (testosterone shown in green and blue color) with the free binding energy.

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