

Verbascoside reverses TGF- β 1-induced renal cellular fibrosis

Abstract

Background: Renal fibrosis is characterized by interstitial cells hypercellularity and matrix protein accumulation, cause of loss normal function and finally kidney failure. Transforming growth factor- β (TGF- β) is a profibrotic cytokine as a key mediator for Smads pathways. *Verbascoside* is a pure compound most from *Osmanthus spp.* In addition, *Verbascoside* has an antioxidants, anti-inflammatory and immunosuppressive functions. In here, we investigated the underlying mechanism of *Verbascoside* in the regulation of TGF β 1-induced cellular fibrosis in NRK-49F.

Materials and Methods: NRK-49F (Rat kidney fibroblast Cells) were cultured in TGF- β 1 (5ng/ml) for days. The cells treatment with different concentrations of *Verbascoside* (0.1 μ M, 1 μ M, 10 μ M) in last 24 hours. The effect of *Verbascoside* on cell viability in renal fibrosis by MTT test. Here we evaluated the in vitro cytotoxic effect of *Verbascoside* in renal fibrosis using the LDH release assay. We determined to EMT relevant markers including fibronectin and TGF- β 1/Smads transducer including Smad4, Smad2/3 and Smad7 by Western blot assay. The expression of the extracellular fibronectin by ELISA. Results: MTT analysis showed that *Verbascoside* did not significantly affect levels of cell viability. Treating cells with *Verbascoside* did not release a significant amount of LDH compared to control. Western blotting showed that *Verbascoside* dose-dependent reverse TGF β 1-induced expression of fibronectin in NRK49F cells. Immunofluorescence assay showed that *Verbascoside* (10mM) reverse TGF β 1-induced expression of fibronectin in NRK49F cells.

Conclusion: We propose that *Verbascoside* is a potential fibrosis antagonist for renal cells by TGF- β 1/Smads signaling pathway.

Keywords: renal fibrosis, tgf- β 1, verbascoside, fibronectin, tgf- β /smad signaling pathway

Volume 3 Issue 6 - 2016

Jue-Long Wang,¹ Yi-Jyun-Chen,² Pei-fang Hsieh,^{3,4} Shu-Fen Liu,⁵ Tsung-Jen Hung,⁶ Chien-Ya Hung,⁷ Ming-Der Shi,^{2,8} Yu-Ju Hung,^{4,9} Tzu-sui Hung,¹⁰ Yow-Ling Shiue,³ Yu-Lin Yang^{2,4}

¹Department of Physical Medicine and Rehabilitation, Kaohsiung Veterans General Hospital, Taiwan

²Department of Biological Science and Technology, Chung Hwa University of Medical Technology, Taiwan

³Graduate Institute of Biomedical Science, National Sun Yat-sen University, Kaohsiung, Taiwan

⁴Graduate Institute of Medical Laboratory Science and Biotechnology, Chung Hwa University of Medical Technology, Taiwan

⁵Department of Internal Medicine, Kaohsiung Medical University Chung-Ho Memorial Hospital, Kaohsiung, Taiwan

⁶Graduate Institute of Biomedical Science, Chung Hwa University of Medical Technology, Taiwan

⁷Department of Food nutrition, Chung Hwa University of Medical Technology, Taiwan

⁸Department of Medical Technology, Kaohsiung Veterans General Hospital Tainan Branch, Taiwan

⁹Department of Public Health, National Taiwan University, Taiwan

¹⁰Department of sport, Health and leisure, Chung Hwa University of Medical Technology, Taiwan

Correspondence: Yang Yu-Lin, Graduate Institute of Medical Laboratory Science and Biotechnology, Chung Hwa University of Medical Technology, Taiwan, Tel +886 6 267 7250, Fax +886 267 7250, Email Call 0955443221@gmail.com

Received: July 07, 2016 | **Published:** October 28, 2016

Introduction

Renal interstitial fibrosis is the common feature of all forms of renal failure, is characterized by interstitial cells hypercellularity and matrix protein accumulation, considered the hallmark of progressive renal disease.¹⁻³ The important evaluating of fibrogenesis is fundamental to such efforts, and that includes an investigating of renal fibroblasts, which are the cells that are primarily responsible for fibrogenesis.⁴⁻⁷ Fibroblasts were principal mediators of renal fibrosis, which contribute cellular elements being the sole source of ECM.^{2,5,8-10} Many studies have determined that the extent of interstitial involvement the accumulation of extracellular matrix (ECM) components including collagen types I, III, and IV, as well as proteoglycans and fibronectin.^{4,11-14} The fibroblasts in produce consequent to a fibronectin-rich and fibrillar ECM that fills up the interstitium, which leads, in turn, to nephron loss and the associated decline in kidney function. In this study, we investigation of the phenotypes of the interstitial fibroblasts, in addition to investigating the processes that may cause their modulation.

A number of growth factors have been found to be involved in the pathogenesis of renal interstitial fibrosis, including transforming growth factor (TGF- β 1), which stimulates the deposition of both EMT

and ECM.^{2,6,8,15} Many studies showed that TGF- β is major profibrotic cytokines and play a key role in fibrogenesis. TGF- β stimulate both type I and type II TGF- β receptors and upregulated the Smad family of transcriptional activators to transmit various signals. Such as receptor-regulated Smad (R-Smad), Smad2/3 are phosphorylated by activated type I receptors, which serves as a mediator for all of them.^{5,6,8,11,14,15} On the other hand, smad7 is firmly combined with TGF- β 1 receptor, direct to the inability of smad 2/3 to be activated as well as inhibition of the signal transduction pathways. Smad7 inhibit the p-smad2/3, nuclear translocation of activated smad complexes, resulting in decreased fibronectin expression and complete inhibition of TGF- β signal transduction.^{2,5} When transcriptional activators are translocated into the nucleus, where they bind to specific Smad-binding elements in the regulate of TGF- β target genes and then effectively guide transactivation. TGF- β 1/Smad signaling plays an important role in the ECM and, in turn, the pathogenesis of renal fibrosis. That said, further investigations are needed because the development of renal fibrosis is highly complex and our understanding of the condition remains far from complete.

Verbascoside can be found in species in *Osmanthus spp.*¹⁶ Verbascoside is hydrophilic in nature and relevant effects in the reduction of symptoms correlated to chronic pathologies and

conditions such as Arthritis, Hypertension, Parkinson's disease, Alzheimer's disease, Estrogenic mediated diseases, Allergy type 1, Intestinal mucositis.¹⁷⁻²¹ More importantly, *Verbascoside* also demonstrated various biological properties including anti-hepatotoxic, antinflammatory, anti-nociceptive, antioxidant and antihemolytic effects and antineoplastic properties in addition to numerous wound-healing and neuroprotective properties.^{17,19,20,22-32} However, literature no survey supporting the therapeutic effects of *Verbascoside* in treatment of renal fibrosis.

The present study was investigate the underlying mechanism of *Verbascoside* in renal cellular fibrosis, with the results showing that *Verbascoside* does, indeed, play a pivotal role in its regulation. The results further suggested that *Verbascoside* reversal of TGF- β ₁-induced cellular fibrosis while simultaneously suppressing the expression of fibronectin and fibrogenic signal proteins. More specifically, *Verbascoside* could potentially serve as an antagonist against both TGF- β ₁ signaling and fibrosis, in part through its down-regulation of TGF- β RII and pSmad2/3. In addition, *Verbascoside* increased negative-regulation of TGF- β proteins such as Smad 7 and E-cadherin. The findings of first evidence report that *Verbascoside* is a novel agent against TGF- β signaling and renal interstitial fibrosis.

Materials and methods

Cell culture and treatment

NRK-49F cells (CRL-1570) were procured from the American Type Culture Collection (ATCC), while Dulbecco's modified Eagle's medium (Gibco, Carlsbad, CA) supplemented with 5% bovine calf serum (BCS), 100U/ml penicillin, and 100mg/ml streptomycin (HycloneLabs, Logan, UT) was used to culture a normal Rattus norvegicus kidney cell line at 37°C under 5% CO₂. The cells were then trypsinized using 0.05% trypsin-EDTA (Hyclone).

Cell proliferation by MTT assay

MTT assays were performed to evaluate the cell viability of NRK-49F cells. Cells (1×104 cells/dl) were plated and incubated for 24hrs in wells of a 96-well plate. Then treated with culture medium including on contraction of TGF- β (5ng/ml) and/or Verbascoside. After 24hrs incubation, 10 μ l of sterile MTT dye were added, and the cells were incubated for 6 hrs at 37°C. Then, 100 μ l of acidic isopropanol (0.04M HCl in isopropanol) were added and thoroughly mixed. Spectrometric absorbance at 595nm (for formazan dye) was measured with the absorbance at 655 nm for reference.

LDH Assay for cytotoxicity

Cells were maintained and passaged as described above. The cells were seeded in 96 well plates at a density of 2×104 cells/well in complete medium and incubated at 37°C in 5% CO₂ overnight. Supernatant from the conditioned cells was collected and stored. Supernatant from maintained cells treated with 1% Triton X-100 was regarded as a positive control for maximum lactate dehydrogenase (LDH) release. After 24h incubation at 37°C in 5% CO₂, the supernatants were collected and centrifuged at 4,500g for 5min to remove contaminating cells, and the level of LDH measured in duplicate using a cytotoxicity detection kit (Clontech, CA, US)

Enzyme-Linked Immunosorbent Assay (ELISA)

ELISA was used to evaluate the expression of secreted fibronectin. To quantify fibronectin in the supernatant of cultured NRK-49F

cells, conditioned culture medium was collected and centrifuged at 1200rpm for 5min to remove particulates. The clear supernatant was then collected and concentrated, then stored at -80°C for further use. A commercial sandwich enzyme-linked immunosorbent assay kit was used for detection of extracellular fibronectin (Takara Bio, Inc., Shiga, Japan) or TGF- β ₁ (R&D Systems, Minneapolis, MN). Detection was performed according to the manufacturer's instructions. Sample absorbances at 450nm were analyzed using an ELISA reader, and the concentration of each sample determined by interpolation with a standard curve, generated using an exogenous fibronectin (12.5, 25, 50, 100, 200, 400, 800ng/ml) or TGF- β ₁ (0-2,000pg/ml) as the standard.

Immunofluorescence staining

Cells were cultured in glass chamber slides (Nunc, Rochester, NY) then fixed with 4% paraformaldehyde for 10min, permeabilized with 0.1% Triton X-100 in phosphate-buffered saline (PBS). After incubation blocked with 1% bovine serum albumin in PBS for 1hr. The Cells were incubated with primary antibody overnight, washed with PBS, and incubated with the FITC-conjugated secondary antibody for 1hr. The slides were mounted with Fluorescent Mounting Medium (Santa Cruz), and observed with a fluorescence microscope (Olympus, Japan).

Western blot analysis

Western blot assay was used to evaluate protein expression of the RII TGF- β receptors, and their downstream signal transducers (e.g., Smad7). In brief, cells were lysed using lysis buffer (10mM Tris, 1mM EDTA, 1% Triton X-100, 1mM Na3VO4, 20 μ g/ml aprotinin, 20 μ g/ml leupeptin, 1mM dithiothreitol, and 50 μ M PMSF). The crude protein lysate was resolved by 10% sodium dodecyl sulphate-polyacrylamide gel electrophoresis (SDS-PAGE) under reducing conditions and transferred to polyvinylidene difluoride membranes (Millipore, Bedford, MA). Samples were then blocked with 10% (w/v) non-fat milk in Tris buffer saline tween (TBS-T) for 1h at 37°C. Individual membranes were probed with a 1:2000 (v/v) ratio of rabbit polyclonal antibodies to anti-Smad2/3 (sc-8332), anti-pSmad2/3 (sc-11769), anti-Smad7 (sc-11392), anti-E-cadherin (sc-7870), anti-T β RII (sc-1700; Santa Cruz Biotechnology, Santa Cruz, CA), and a 1:2000 ratio of anti- β -actin (Sigma-Aldrich, St Louis, MO A-5316). After hybridization at 37°C, blots were washed and hybridized with 1:4000 (v/v) dilutions of goat anti-rabbit IgG or horseradish peroxidase-conjugated secondary antibody (Jackson ImmunoResearch Laboratories, West Grove, PA) or donkey anti-mouse IgG- or horseradish peroxidase (Santa Cruz). The signals were visualized using enhanced chemiluminescence (ECL), with β -actin as an internal control.

Statistics

The results were expressed as mean \pm SEM. The unpaired Student's t test was used for comparison between two groups. Values of P $<$ 0.05 were considered to be statistically significant. For the in vivo experiments, a one-way analysis of variance (ANOVA) was used to determine if the means were significantly different (P $<$ 0.05). If the means were significantly different, a Tukey-Kramer multiple group comparison test was used to compare the individual groups. The standard error was indicated for each value by a bar, and the significance was indicated for each comparison. A one-way ANOVA test was used to calculate P values for all the ratios. All values were calculated using GraphPad Prism, version 3.00, for Macintosh (GraphPad Software, San Diego, CA).

Results

We want to understand the fibrosis-regulatory effects of *Verbascoside* extract on renal fibroblast cells. To examine whether *Verbascoside* blocks TGF- β -induced change of NRK-49F cells, we

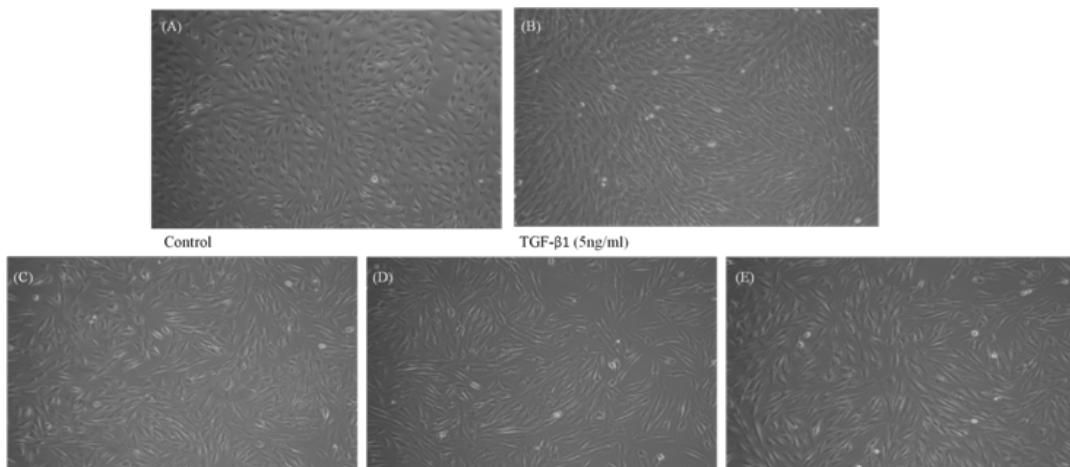


Figure 1 Effects of *Verbascoside* on the cell morphological transformations.

We evaluate the underlying effects of *Verbascoside* on TGF- β -induced pharmacology. Cell survival and protein content were analyses in cells treated with TGF- β (5ng/ml) and *Verbascoside* (0.1, 1 and 10 μ M) by LDH and MTT were performed (Figure 2A) (Figure 2B). We found that *Verbascoside* does not affect the viability

pretreated cells with TGF- β for 24hours, followed by *Verbascoside* at the indicated concentrations for 24hours. As figure 1, cellular morphology was significantly and dose-dependently (i.e. 0.1, 1, 10 μ M) restored compared with TGF- β -treating group (n=4, p<0.05).

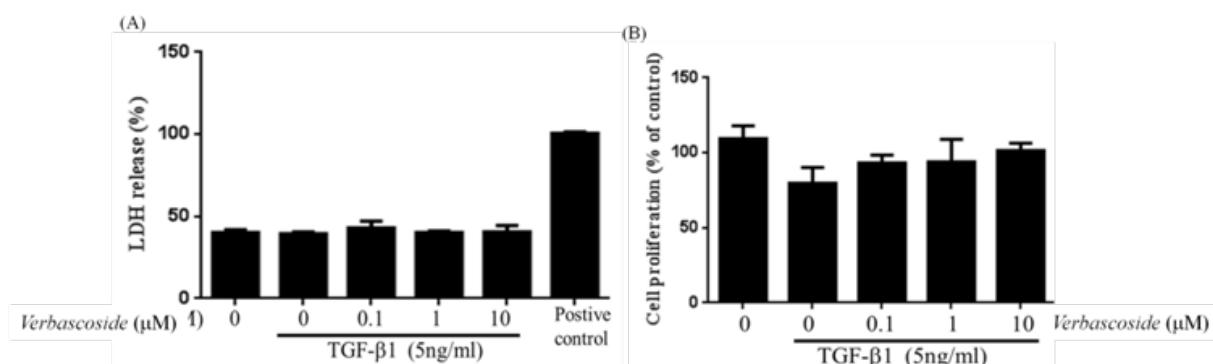


Figure 2 Effects of *Verbascoside* on levels of Cell proliferation by (A) LDH assay and (B) MTT assay.

To understanding the effects of anti-fibrosis on the regulation of TGF- β and *Verbascoside* dose-dependently in fibronectin bioactivity by ELISA assay. TGF- β (5ng/ml) significantly increase extracellular fibronectin in NRK-49F cells compared to control in Figure 3A. More importantly, the *Verbascoside* dose-dependently (0.1, 1, 10 μ M)

of cultured fibroblasts. These observations demonstrated that either *Verbascoside* or TGF- β does not statistically affect cellular survival and protein content according to Figure 2. Thus, *Verbascoside* extract does not exert its effects by inhibiting cell growth or by inducing cell death.

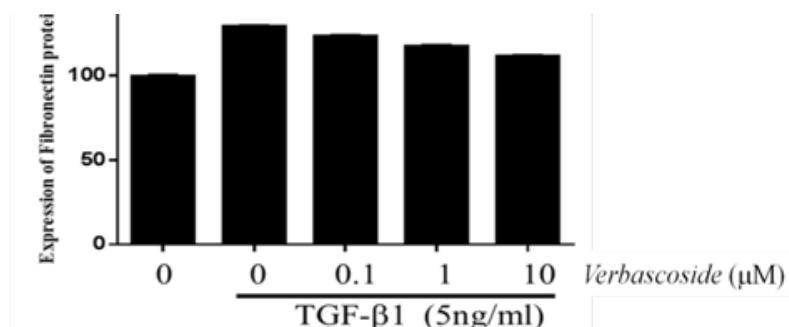


Figure 3 Effects of *Verbascoside* on levels of secreted fibronectin.

To clarify the mechanism by which *Verbascoside* regulates renal cellular fibrosis, the expressions of TGF- β signal pathway were investigated. As shown in Figure 4, the administration of exogenous TGF- β (5ng/ml) significant increases in the expression of fibronectin in the cells, whether of extracellular origin (as determined by ELISA) (Figure 3) or intracellular origin (as determined by Western blot) (Figure 4). More importantly, *Verbascoside* dose-dependently (0.1, 1, 10 μ M) and dramatically suppressed TGF- β -induced increases in fibronectin levels. The presence of TGF- β receptors strongly correlates with susceptibility to cellular fibrosis. As shown in Figure 4, TGF- β induced a significant increase in the level of type II TGF- β receptors. Most importantly, Western blot analysis showed a statistically significant reduction expression of T β RII for the *Verbascoside*. Since the Smad family is the most important mediator for TGF- β signaling. As shown in Fig. 4, TGF- β (5ng/ml) significantly increased Smad2/3 levels. Intriguingly, *Verbascoside* dose-dependently (0.1, 1, 10 μ M) dramatically suppressed TGF- β -induced increases in Smad2/3 in a dose-dependent manner. These observations show that *Verbascoside* may reverse TGF- β -induced cellular fibrosis by regulating and suppressing TGF- β down-stream signals. We examined Smad7 and E-cadherin, a powerful intracellular TGF- β antagonist. Treatment with TGF- β (5ng/ml) induced significant decreases in Smad7 and E-cadherin. The *Verbascoside* dose-dependently (0.1, 1, 10 μ M) and dramatically reversed TGF- β -induced decreases in Smad7 and E-cadherin. In other words, *Verbascoside* might ameliorate renal cellular fibrosis by inducing an increase in inhibitory Smad7 and E-cadherin. We found that 10 μ M *Verbascoside* also dramatically decreased Smad 2/3. Thus, *Verbascoside* has potential to induce TGF- β signaling and so regulate renal cellular fibrosis.

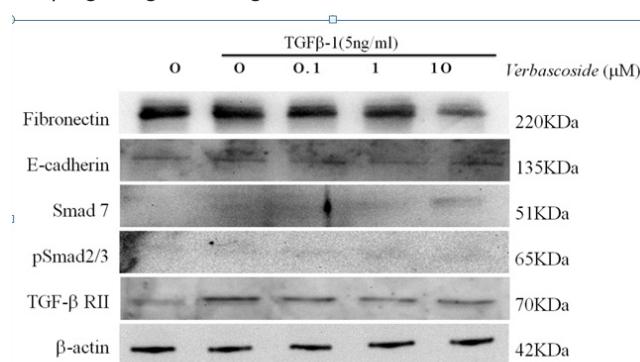


Figure 4 Effects of *Verbascoside* on TGF- β 1-induced both of fibronectin expression and TGF- β 1/Smads signaling pathway in NRK-49F cells.

The morphologies of cells undergoing TGF- β induced fibroblast transition were observed through a immunofluorescence microscope. The expression of fibronectin of extracellular origin in the cells, however, was significantly increased by exogenous TGF- β , as determined by immunofluorescence staining in Figure 5. Treatment with *Verbascoside* 10 μ M restored the normal expression of the NRK-49F cells. On the basis of our results, we were able to identify a clear mechanism by which *Verbascoside* reverse renal cellular fibrosis. Specifically, the fibrosis appeared to be induced via the upregulation of the type II receptor of TGF- β , as well as the upregulation of Smad 2/3 and fibronectin. At the same time, TGF- β caused a significant decrease in the expression of Smad7. Meanwhile, *Verbascoside* caused significant attenuation of the TGF- β -induced upregulation of the type II receptor of TGF- β , Smad2/3, as well as significant attenuation of the TGF- β -induced downregulation of fibronectin. These results

suggest that *Verbascoside* might be possible to downregulate TGF- β signal proteins, allowing it to be used as a novel treatment of renal fibrosis.

Discussion

Our results demonstrate that *Verbascoside* plays a protective role against TGF- β 1-induced renal fibrosis. *Verbascoside* antagonizes TGF- β 1-induced renal fibrosis possibly by enhancing the expression of Smad7 in renal cells. Meanwhile, we show that *Verbascoside* increases the expression of E-cadherin. Thus, *Verbascoside* has the potential to inhibit TGF- β -induced renal tubular fibrosis possibly by regulating Smads pathway. Due to the range of pharmacological activities and biological of *Verbascoside* and insufficient data on the safety reports it may be needed for assessing toxicity profiles. In this study, cellular toxicity of main constituent of *Verbascoside* was determined. We showed *Verbascoside* does not statistically affect cellular survival and viability according to Figure 2. These observations were consistent with those reported Aleo et al.³³ Dell'Aquila et al.³⁴ indicating revealed a significant antioxidant effect of *Verbascoside*.^{18,33,34} In 2013, Potapovich et al.³⁵ showed *Verbascoside* offers additional skin-protection capacity against harmful ultraviolet radiation and inflammatory insults.³⁵ Both of Georgiev et al.,²⁰ has been demonstrated HepG2 and NIH cells were exposed to different concentrations of *Verbascoside* in MTT assay.¹⁸ All of the above are consistent with this study.

When kidney injury occurs, activated renal cells can release TGF- β , that in turn activates fibroblasts, therefore, TGF- β has a close relationship with fibrosis.^{6,9} The formation of renal fibrosis is a complex process of multifactor and multicell involvement. In this pathological process, the cell-cell, cell-cytokines and cell-matrix interactions constitute a cumbersome network. In this network, the development of renal fibrosis can be regulated by different signaling pathways and means. TGF- β is a major activator of kidney and a key mediator in the pathogenesis of renal fibrosis. TGF- β stimulates both type I and type II TGF- β receptors and then utilizes the Smad family of transcriptional activators to transmit various signals. Such as receptor-regulated Smad (R-Smad), Smad2, and Smad3 are phosphorylated by activated type I receptors, after which they bind to Smad4, which serves as a mediator for all of them.^{1,4,6,11,13,36} On the other hand, smad7 is firmly combined with TGF- β receptor, leading to the inability of smad 2/3 to be activated as well as inhibition of the signal transduction pathways. We speculate that *Verbascoside* may be used as a kind of treatment of renal fibrosis, and inhibit the formation of renal fibrosis.

In Figure 4, *Verbascoside* could be considered as an attractive therapeutic strategy for inhibition of cellular fibrosis after NRK-49F was stimulated with TGF- β in vitro. More specifically, *Verbascoside* can inhibit the expressions of TGF- β /smad pathway in NRK-49F. *Verbascoside* can decrease smad2/3 expressions, and increase smad7 expression and increase E-cadherin expression suggesting that *Verbascoside* also plays a role in inhibiting NRK-49F activation and thus inhibit renal fibrosis, highlighting a potential anti-fibrotic mechanism, as illustrated in Figure 4. The findings of first evidence report that *Verbascoside* is a novel agent against TGF- β signaling and renal interstitial fibrosis. Reportedly, *Verbascoside* is regulated in different organ cells, including *Verbascoside* promoted apoptosis via p53 in human CRC cells.²⁷ Furthermore reported that *Verbascoside* attenuates LPS-induced pro-inflammatory mediator production via TAK-1/JNK/AP-1 signalling in U937 cells.³⁷ Therefore, the role of

Verbascoside as a molecule able to regulated via different signal pathway in organ cells.

In summary, our study provides the new medicine *Verbascoside*, which exhibits potent effects in *Verbascoside* reverse renal fibrosis via TGF- β signal pathway. Thus, *Verbascoside* has potential to be developed into an therapeutic against renal fibrosis.

Figure 1 Effects of *Verbascoside* on the cell morphological transformations. The observation shows TGF- β ₁ induced NRK-49F cells fibrosis at different concentrations of *Verbascoside* to be treated by an inverted microscope. The experimental group of cells are without any morphological abnormalities, which represents *Verbascoside* does not affect the growth of NRK-49F cells. The experimental results showed that *Verbascoside* will not cause harm to the cells, and therefore does not have cell cytotoxicity.

Figure 2 Effects of *Verbascoside* on levels of Cell proliferation by (A) LDH assay and (B) MTT assay. NRK-49F cells were treated with TGF- β (5ng/ml) in 0.5% BCS for 48h, followed by treatment with *Verbascoside* (0.1, 1,10 μ M) for a further 24h. The absorbance of each sample was then analyzed by an ELISA reader. Supernatant was collected and subjected to MTT and LDH analysis. We found that *Verbascoside* does not affect the viability of cultured fibroblasts and inhibiting cell growth or inducing cell death.

Figure 3 Effects of *Verbascoside* on levels of secreted fibronectin. The observation shows (A) Standard curves were generated using known concentrations of fibronectin (0, 0.39, 1.56, 6.25,25ng/ml). The absorbance (450 nm) of each sample was then analyzed by an ELISA reader. (B) NRK-49F cells were treated with TGFb1 (5ng/ml) in 0.5% BCS for 48h, followed by treatment with *Verbascoside* for another 24h. Supernatant was collected and subjected to fibronectin

ELISA analysis. Fibronectin level was determined by interpolation with the standard curve. The fibronectin level of each condition was normalized to the cell number of each well. And contrast the normal control group, showing the amount of pathology group extracellular fibronectin has significantly increased, which means that changes in TGF- β ₁ can induce NRK-49F cells to secrete more fibronectin. The observation ELISA showed that *Verbascoside* (10 μ M) dose-dependent reverse TGF- β ₁-induced increase in fibronectin in NRK49F cells.

Figure 4 Effects of *Verbascoside* on TGF- β ₁-induced both of fibronectin expression and TGF- β ₁/Smads signaling pathway in NRK-49F cells. The observation shows NRK-49F cells were treated with TGF- β 1 (5ng/ml) in 0.5% BCS for 48h, followed by treatment with *Verbascoside* for another 24h. Western blot analysis, Cell extracts were subjected to SDS-PAGE and immunoblot with a primary antibody against fibronectin. The expression of b-actin was used as an internal control. Western blotting showed that *Verbascoside* (10 μ M) dose-dependent reverse TGF- β ₁-induced increase in fibronectin expression and in NRK49F cells. And TGF- β ₁ through TGF- β ₁/Smads signaling pathway induced renal cellular fibrosis.

Figure 5 Effects of *Verbascoside* on TGF- β ₁-induced fibronectin expression in NRK-49F cells by Immunofluorescence assay. The observation shows NRK-49F cells were treated with TGF- β ₁ (5ng/ml) in 0.5% BCS for 48h, followed by treatment with *Verbascoside* for another 24h. Immunofluorescence assay, grow cultured cells on chamber slides, fix cells by 4% Paraformaldehyde and add ice cold Triton X-100 break through the cells. At last immunoblot with a primary antibody against fibronectin. Immunofluorescence assay showed that *Verbascoside* (10 μ M) reverse TGF- β ₁-induced increase in fibronectin expression in NRK49F cells.

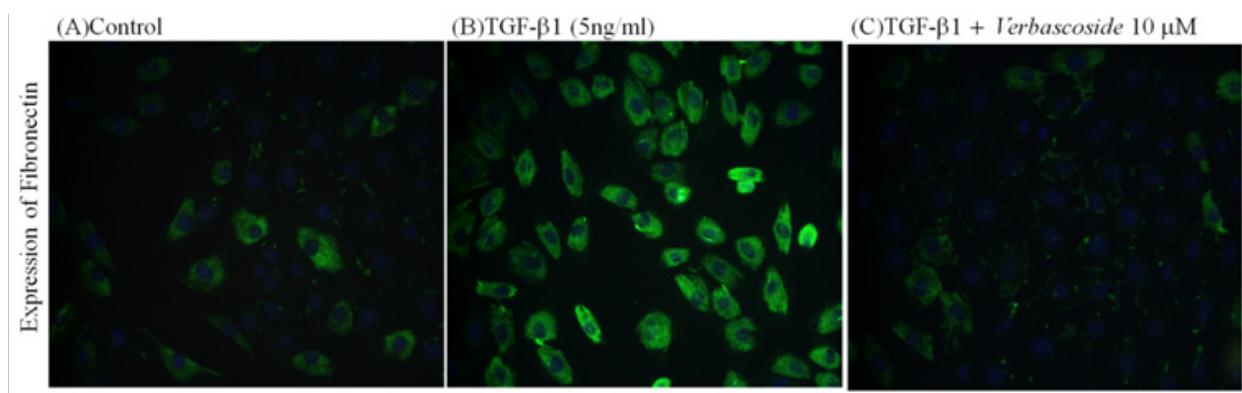


Figure 5 Effects of *Verbascoside* on TGF- β 1-induced fibronectin expression in NRK-49F cells by Immunofluorescence assay

Acknowledgements

None.

Conflict of interest

Author declares that there is no conflict of interest.

References

1. Cho MH. Renal fibrosis. *The Korean Pediatric Society*. 2010;53(7):735–740.
2. Jack Lawson JE, Caroline Wheeler-Jonesa, Harriet Symeb, et al. Renal fibrosis in feline chronic kidney disease: Known mediators and mechanisms of injury. *The Veterinary Journal*. 2015;203(1):18–26.
3. Jun Yang JC, Jingyin Yan, Liping Zhang, et al. Effect of Interleukin 6 Deficiency on Renal Interstitial Fibrosis. *PLoS One*. 2012;7(12):e52415.
4. Kis K, Liu X, Hagood JS. Myofibroblast differentiation and survival in fibrotic disease. *Expert Rev Mol Med*. 2011;13:e27.
5. Liu Y. Renal fibrosis: New insights into the pathogenesis and therapeutics. *Kidney Int*. 2006;69(2):213–217.
6. Guo-tao Chen, Xiao-hui Liao, Ru-yu Yan, et al. Augmenter of liver regeneration ameliorates renal fibrosis in rats with obstructive nephropathy. *Biosci Rep*. 2014;34(5).pii:e00135.
7. Xia Y, Entman ML, Wang Y. Critical Role of CXCL16 in Hypertensive Kidney Injury and Fibrosis. *Hypertension*. 2013;62(6):1129–1137.
8. Huang WY, Li ZG, Rus H, et al. RGC-32 Mediates Transforming Growth Factor- β -induced Epithelial-Mesenchymal Transition in Human Renal Proximal Tubular Cells. *J Biol Chem*. 2009;284(14):9426–9432.
9. Flaviu Bob GG. Processes of Fibrogenesis in Glomerular Nephropathies. *Journal of Timisoara medical*. 2004;3.

10. Shailendra P Singh ST, Timothy A Fields, Sydney Webb, et al. Glycogen synthase kinase-3 inhibition attenuates fibroblast activation and development of fibrosis following renal ischemia-reperfusion in mice. *Dis Model Mech.* 2015;8(8):931–940.
11. Liu Y. Epithelial to Mesenchymal Transition in Renal Fibrogenesis: Pathologic Significance, Molecular Mechanism, and Therapeutic Intervention. *J Am Soc Nephrol.* 2004;15(1):1–12.
12. Zeisberg FSA. Renal Fibroblasts and Myofibroblasts in Chronic Kidney Disease. *J Am Soc Nephrol.* 2006;17(11):2992–2998.
13. Deborah L Clarke AMC, Tomas Mustelin, et al. Matrix regulation of idiopathic pulmonary fibrosis: the role of enzymes. *Fibrogenesis Tissue Repair.* 2013;6(1):20.
14. Na Liu ET, Murugavel Ponnusamy, Haidong Yan, et al. Delayed Administration of Suramin Attenuates the Progression of Renal Fibrosis in Obstructive Nephropathy. *J Pharmacol Exp Ther.* 2011;338(3):758–766.
15. Ying Li YS, Fuyou Liu, Lin Sun, et al. Norcantharidin Inhibits Renal Interstitial Fibrosis by Blocking the Tubular Epithelial-Mesenchymal Transition. *PLoS One.* 2013;8(6):e66356.
16. Søren Rosendal Jensen, Henrik Franzky, Eva Wallander. Chemotaxonomy of the Oleaceae: iridoids as taxonomic markers. *Phytochemistry.* 2002;60(3):213–231.
17. Alipieva K, Korkina L, Orhan IE, et al. Verbascoside-A review of its occurrence, (bio)synthesis and pharmacological significance. *Biotechnol Adv.* 2014;32(6):1065–1076.
18. Leila Etemad RZ, Naser Vahdati- Mashhadian, Seyed Adel Moallem, et al. Acute, Sub-Acute and Cell Toxicity of Verbascoside. *Research Journal of Medicinal Plants.* 2015;9(7):354–360.
19. Covas MI NK, Poulsen HE, Kaikkonen J, et al. EUROLIVE Study Group. Prooxidant effects of verbascoside, a bioactive compound from olive oil mill wastewater, on in vitro developmental potential of ovine prepubertal oocytes and bioenergetic/oxidative stress parameters of fresh and vitrified oocytes. *Ann Intern Med.* 2006;145(5):333–341.
20. Georgiev M, Pastore S, Lulli D, et al. Verbascum xanthophoeniceum-derived phenylethanoid glycosides are potent inhibitors of inflammatory chemokines in dormant and interferon-gamma-stimulated human keratinocytes. *J Ethnopharmacol.* 2012;144(3):754–760.
21. Mosca M, Ambrosone L, Semeraro F, et al. Ocular tissues and fluids oxidative stress in hares fed on verbascoside supplement. *Int J Food Sci Nutr.* 2014;65(2):235–240.
22. Akdemir Z, Kahraman C, Tatlı II, et al. Bioassay-guided isolation of anti-inflammatory, antinociceptive and wound healer glycosides from the flowers of *Verbascum mucronatum* Lam. *J Ethnopharmacol.* 2011;136(3):436–443.
23. Alfonso Varela-López, Pedro Bullón FG, José L. Quiles. Non-Nutrient, Naturally Occurring Phenolic Compounds with Antioxidant Activity for the Prevention and Treatment of Periodontal Diseases. *Antioxidants (Basel).* 2015;4(3):447–481.
24. Ambrosone LG, Germano Cinelli, Mariapia Filippelli, et al. Corneal Epithelial Wound Healing Promoted by Verbascoside-Based Liposomal Eyedrops. *Biomed Res Int.* 2014;2014:471642.
25. Campo GM, Bristot L, Monti M, et al. The in vitro effects of verbascoside on human platelet aggregation. *J Thromb Thrombolysis.* 2012;34(3):318–325.
26. Kirmizibekmez H, Ariburnu E, Masullo M, et al. Iridoid, phenylethanoid and flavonoid glycosides from *Sideritis trojana*. *Fitoterapia.* 2012;83(1):130–136.
27. Zhou L, Feng Y, Jin Y, et al. Verbascoside promotes apoptosis by regulating HIPK2-p53 signaling in human colorectal cancer. *BMC Cancer.* 2014;14:747.
28. Luigi Ambrosone GG, Mariapia Cinelli, Mariaelena Filippelli, et al. Corneal Epithelial Wound Healing Promoted by Verbascoside-Based Liposomal Eyedrops. *BioMed Research International.* 2014;2014:8.
29. Mauriz E, Vallejo D, Tuñón MJ, et al. Effects of dietary supplementation with lemon verbena extracts on serum inflammatory markers of multiple sclerosis patients. *Nutr Hosp.* 2014;31(2):764–771.
30. Sheng GQ, Zhang JR, Pu XP, et al. Protective effect of verbascoside on 1-methyl-4-phenylpyridinium ion-induced neurotoxicity in PC12 cells. *Eur J Pharmacol.* 2002;451(2):119–24.
31. Scarpati MLaFDM. Isolation from *Verbascum sinuatum* of two new glucosides, verbascoside and isoverbascoside. *Ann Chim.* 1963;53:356–367.
32. Liang JQ, Wang L, He JC1, et al. Verbascoside promotes the regeneration of tyrosine hydroxylase-immunoreactive neurons in the substantia nigra. *Neural Regen Res.* 2016;11(1):101–106.
33. Aleo E, Ricciand R, Passi S, et al. A novel cyt-H2O2-chemiluminescences assay for measuring the reducing/antioxidant capacity on hydrophilic and lipophilic antioxidants and biological samples. *Prog Nutr.* 2005;7(3):154–182.
34. Dell'Aquila ME, Bogliolo L, Russo R, et al. Prooxidant effects of verbascoside, a bioactive compound from olive oil mill wastewater, on in vitro developmental potential of ovine prepubertal oocytes and bioenergetic/oxidative stress parameters of fresh and vitrified oocytes. *Biomed Res Int.* 2014;2014:878062.
35. Pastorelli G, R Rossi, C Corino. Influence of *Lippia citriodora* verbascoside on growth performance, antioxidant status and serum immunoglobulins content in piglets. *Czech J Anim Sci.* 2012;57(7):312–322.
36. Liu X, Hu H, Yin JQ. Therapeutic strategies against TGF- β signaling pathway in hepatic fibrosis. *Liver Int.* 2006;26(1):8–22.
37. Pesce M, Franceschelli S, Ferrone A, et al. Verbascoside down-regulates some pro-inflammatory signal transduction pathways by increasing the activity of tyrosine phosphatase SHP-1 in the U937 cell line. *J Cell Mol Med.* 2015;19(7):1548–1556.