Risk Factors and Contemporary Treatment of Orthodontically Induced Apical Root Resorption: A Review

Abstract

Aim: External apical root resorption is one of the most common side effects of orthodontic treatment. Some amount of root resorption is inevitable in orthodontic. Most of the times root resorption is not severe and goes unnoticed but in few cases root resorption is severe. The cause of root resorption is multifactorial. Aim of this article is to give an insight of root resorption caused by orthodontic tooth movement.

Material and method: Articles pertinent to root resorption caused by orthodontic tooth movement were first searched electronically and then hand searched. Relevant articles were topic wisely critically reviewed by all the reviewers.

Conclusion: It was concluded that many factors are involved in root resorption. In this article possible method of curbing root resorption have also been described.

Keywords: External apical root resorption; Osteopontin; Rankl; Orthodontic treatment; iatrogenic root resorption

Introduction

External apical root resorption is an unwanted complication of orthodontic treatment which leads to permanent shortening of the roots. There are many factors attributed to root resorption. Some factors causing root resorption are natural some are pathologic and some are iatrogenic. Orthodontic treatment is one of the main causes for iatrogenic root resorption. Amongst the many factors definitely orthodontic treatment is an important trigger factor for root resorption [1]. Root resorption can begin in the early leveling stages of orthodontic treatment which can continue throughout the treatment [2].

Computerized tomography is a superior tool than conventional radiography to estimate root resorption. Conventional radiographic image is a 2D image of 3D object whereas computerized tomography tool which is capable of giving a 3D image. Computed tomography studies have shown that orthodontically induced root resorption occurs in up to 91% of teeth [3].

CBCT is a superior tool to assess root resorption. Whenever patients have a high risk factor for external apical root resorption (EARR) then imaging by CBCT should be done especially when decision has to be taken to modify or terminate the treatment [4]. Even periapical radiographs may underestimate the actual amount of root resorption when compared to computerized tomography [5]. Digital Subtraction radiography has also been used to estimate the amount of root resorption along with corresponding changes in the bone [6,7].

The method of Malmgren et al. [8] has been the gold standard for measurement of root resorption. Scoring system of Malmgren described grade 0, no root resorption; grade 1, mild resorption, root with a normal length and only displaying an irregular contour; grade 2, moderate resorption, small area of root loss with the apex exhibiting an almost straight contour; grade 3, accentuated resorption, loss of almost one-third of root length; grade 4, extreme resorption, loss of more than one-third of the root length [8,9] (Figure 1).

Discussion

There are two types of factors related to external apical root resorption. One is a patient related factor that includes genetics, systemic factors, allergy and asthma. Severity of malocclusion, previous history of root resorption, and root morphology and root proximity with cortical bone also affect root resorption. Other is orthodontic treatment-related risk factors that include the treatment duration, magnitude of applied force, direction of...
tooth movement, amount of apical displacement, and method of force application [10]. However Taner was not able to establish any significant correlations between the amount of apical root resorption and tooth inclination, and the duration of active treatment [11]. Roots of maxillary incisors undergo maximum amount of root resorption. The risk of resorption is greatly increased with prolonged treatment time [12]. Root resorption of maxillary lateral incisors is the second most common to undergo root resorption followed by mandibular lateral incisor [13].

The incidence of root resorption is almost hundred percent with 1.3% of cases having severe root resorption exceeding more than 3mm [14]. Brin et al. [15] reported that the incidence of root resorption was almost equal in teeth with trauma and without trauma [15]. Severity of apical root resorption does not seem to be affected by gender of the patient [16]. There is no difference in root resorption in orthodontically treated and endodontically treated teeth [17,18].

Root of teeth can be of varying shapes like short, blunt, apically bent or pipette shaped. The degree of root resorption with blunt or pipette shaped is significantly higher than normal teeth [19]. The severity of external apical root resorption is significantly affected by the duration of treatment. Severity of treatment increases with prolonged treatment [20].

Mean root shortening for the four incisors has been reported to be 0.73 mm and 0.67 mm for girls and boys respectively. When using the most severe single root resorption per patient as a parameter the mean was 1.34 mm for both sexes [21]. Apical root resorption in orthodontic cases is not affected by the dental invagination and cannot be considered as a risk factor [22].

External root resorption although inevitable is not influenced by type of appliance. There is no difference in the amount of the root resorption between self ligating brackets and conventional brackets [23,24]. Lingual orthodontic appliance is also associated with slight periapical root resorption [25]. Mean root resorption in cases treated with straight wire method (MBT) was 18.26% compared to 14.82% in patients treated with standard edgewise appliance. Straight wire patients showed more resorption which could be attributed to more root movement because of built in torque [26,27].

Genetic factors

The factors influencing the root resorption due to orthodontic treatment can be broadly divided into genetic and environmental. There is tremendous variation in the severity of root resorption which cannot be explained by environmental factors alone. Genetic factors account for one half to two third of the variation [28], (Table 1). Interleukin 1 gene cluster is present on the long arm of chromosome 2 (2q13). It includes 3 genes. Two genes interleukin 1A (IL-1A) and interleukin 1B (IL-1B) encode for pro inflammatory cytokine protein. IL-1B is responsible for the formation of protein IL-1β and IL-1A is responsible for the synthesis of IL-1α. Third gene is IL-RN encodes a related protein IL-1ra. The protein IL-1β has been implicated in bone resorption (catabolic modeling). Allele 2 of IL-1B at +3954 is associated with four fold increase in production of IL-1B. In future it will be possible to analyze the DNA from a cheek swab of the patient. The patients having two copies of IL-1B gene would be high risk patients [28-30]. Variations in the interleukin 1 receptor antagonist gene (rs419598) and the IL1B gene (rs1800587) are determinants of a predisposition to postorthodontic external apical root resorption [31].

Table 1: Possible genes involved in eternal apical root resorption.

<table>
<thead>
<tr>
<th>Possible Gene involved in Root Resorption</th>
<th>Gene</th>
<th>SNP</th>
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<tbody>
<tr>
<td>1</td>
<td>Interleukin 1 Receptor Antagonist Gene</td>
<td>rs419598</td>
</tr>
<tr>
<td>2</td>
<td>Purinergic-receptor-P2X, Ligand-gated ion Channel 7 (P2RX7)</td>
<td>rs208294</td>
</tr>
<tr>
<td>3</td>
<td>Osteopontin Gene</td>
<td>rs9138 and rs11730582</td>
</tr>
<tr>
<td>4</td>
<td>Interleukin-1β Genes</td>
<td>rs143634</td>
</tr>
<tr>
<td>5</td>
<td>IL1B Gene</td>
<td>rs1800587</td>
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P2X purinoceptor 7 is a protein that in humans is encoded by the P2RX7 gene. The product of this gene belongs to the family of purinoceptors for ATP. P2X receptors have been implicated in ATP-mediated cell death, regulation of receptor trafficking, and inflammation. P2X, receptor signaling increases the release of proinflammatory molecules such as IL-1β, IL-6, and TNF-α. Presence of specific genotypes for P2RX7 SNP rs208294 (purinergic-receptor-P2X7, ligand-gated ion channel 7 (P2RX7; rs208294),) were significantly associated with external apical root resorption (EARR) [32,33].

Genetic variants of the interleukin-1 gene cluster (IL1) are associated with a possible genetically induced variability in post-orthodontic external apical root resorption (EARR) in root filled teeth and their control counterparts with vital pulps Genetic variations in the interleukin-1β gene (rs1143634) predispose root filled teeth to EARR [34].

The main focus of the present study is to clarify whether variants in the interleukin-1 receptor antagonist gene coding for the IL-1ra protein have both positive and negative effect on EARR of endodontically-treated teeth. Variants in allele 1 of the interleukin-1 receptor antagonist gene (rs419598) are associated with a high risk of suffering post-orthodontic EARR in endodontically treated teeth [35].

Non-parametric sibling pair linkage analysis identified evidence of linkage of EARR affecting the maxillary central incisor with the microsatellite marker D18S64 (tightly linked to TNFRSF11A). This indicates that the TNFRSF11A locus, or another tightly linked gene, is associated with EARR [36]. Osteopontin gene is an essential mediator in the odontoclast fusion and attachment process. Variations in the osteopontin gene (rs9138 and rs11730582) are determinants of a genetic predisposition to suffer EARR secondary to orthodontic treatment [37]. During orthodontic tooth movement RANKL expression is induced on the compressed side of the tooth. RANKL activates osteoclastogenesis, and this is better demonstrated by the acceleration of tooth movement, which is achieved after transfer of the RANKL gene to the periodontal tissue [38].

T-lymphocytes secrete soluble s-RANKL. RANKL mRNA is expressed mostly in bone and bone marrow, as well as in lymphoid tissues. The role of RANKL, together with another very important
protein ligand, MCSF (which binds to its receptor c-fms), promotes osteoclast formation, fusion, differentiation, activation and survival, thus enhancing bone resorption. The effects of RANKL are produced when it binds to RANK. The biological effects of OPG are opposite to the RANKL-mediated effects, because OPG acts as a soluble receptor antagonist, which competes with RANKL and therefore prevents RANKL-RANK interaction [39], (Figure 2).

The resorptive activity of osteoclasts, induced by soluble RANKL or cell-bound RANKL, is completely inhibited by the simultaneous addition of OPG. Conclusively, OPG, RANKL and RANK form a key network that regulates bone metabolism and osteoclast biology. RANKL to OPG ratio in periodontal ligament (PDL) cells contribute to root resorption during orthodontic tooth movement. The compressed PDL cells in cases of severe external apical root resorption produce a large amount of RANKL and increases osteoclastogenesis. This explains why there is greater increase of RANKL and decrease of OPG in cases of severe root resorption [40,41].

Repair

It has been reported that repair of denuded root takes place after the active force has been removed. The repair takes predominantly by the formation of secondary cementum. The amount of secondary cementum formed is about three times more after 8 weeks than at 1 week [42].

Macrophages are large cells and are important mediators of inflammation. There are two types of macrophages which are involved in the inflammatory process. The macrophages which encourage inflammation are called M1 and those that decrease inflammation and promote tissue repair are called M2. M1 macrophage metabolize arginine to nitric oxide whereas M2 macrophage convert it into repair molecule ornithine. During active treatment M1 macrophages are upregulated on the compression side when the forces are removed then M2 macrophages are upregulated. Root resorption is aggravated by an enhanced M1/M2 ratio and partially rescued by a reduced M1/M2 ratio [43,44].

Patients at risk of severe apical root resorption can be identified during the initial treatment stages. It advisable to take periapical X rays at frequent intervals in high risk patients to determine progress of resorption [45].

Pressure causes resorption of root but the resorption in such case does not show any radiolucency and the tooth is mostly asymptomatic or associated with mild pain. Root resorption due to pulp pathology will have periapical radiolucency and tooth will be tender on percussion. Root resorption is almost unavoidable in orthodontic patients as such. It needs to be a part of the informed consent process. Root resorption seen due to orthodontic treatment is rarely detrimental and it should be explained to the patient before informed consent.

Drugs

Many drugs effect induced root resorption (Table 2). Corticosteroids are compounds that are derived from cholesterol and have widespread effects on various organs. Synthetic steroids are used as anti-inflammatory and immunosuppressive agents in the treatment of wide range of chronic and medical condition. Prednisolone is a moderately potent synthetic steroid which has been used in animals and humans to study the effects of steroids. Prolonged administration of synthetic corticosteroids is associated with bone loss and osteoporosis. In low dose prednisolone has been associated with decrease in bone resorption.

Table 2: Possible role of drugs in controlling root resorption.

<table>
<thead>
<tr>
<th>Name</th>
<th>Group</th>
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<tbody>
<tr>
<td>1 Prednisolone</td>
<td>Corticosteroid</td>
</tr>
<tr>
<td>2 L-thyroxine</td>
<td>Hormone</td>
</tr>
<tr>
<td>3 Celebrex</td>
<td>Nonsteroidal Anti-inflammatory Cox2 Inhibitor</td>
</tr>
<tr>
<td>4 Clodronate</td>
<td>Bisphosphonates</td>
</tr>
<tr>
<td>5 Doxycyclin</td>
<td>Broad spectrum Antimicrobial Agent</td>
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In animal studies prednisolone has shown to reduce resorption by suppressing the inflammation and clastic activity [46]. Administration of prednisolone and high-dose celecoxib reduces root resorption and interferes with tooth movement in rats. Both drugs may interfere in the arachidonic acid cascade depending on dose thresholds. L-thyroxine decreased the extent of root resorption while increasing the amount of tooth movement. Administration of Celebrex during the application of orthodontic forces does not interfere with tooth movement and appears to offer some slight protection against root resorption [47]. The short and long-term celecoxib administration did not suppress the root resorption in case of experimental orthodontic force application [48].

Systemic administration of low-dose doxycyclin in rats have an inhibits orthodontically induced resorptive activity to some extent [49,50]. A histological section observed under scanning electron microscope has confirmed that root resorption lacunae decrease on administration of thyroxine [51]. Doxycyclin is a potent antimicrobial agent has been shown to have some anti-inflammatory effect. Doxycyclin reduces the resorption by directly inhibiting the osteoclasts [52,53]. Doxycyclin inhibits the orthoclastic activity that is not related to infectious agents.

reduction in orthodontic activity is due to apoptosis of osteoclast. Low dose of doxycyclin may be useful in to controlling root resorption clinically [54,55].

Bisphosphonates are a class of drugs that prevent the loss of bone mass, used to treat osteoporosis and similar diseases. They are the most commonly prescribed drugs used to treat osteoporosis. Local application of riserdronate in rats arrests root resorption [56,57]. Similarly Clodronate, a non-N-containing bisphosphonate, inhibits bone resorption and also has anti-inflammatory properties. Localized use of doxodronate could be a useful in orthodontic treatment [58]. Root resorption also occurs in reimplanted teeth. Soaking the teeth in alandronate solution rather than Hank’s Balanced salt solution reduces the chance of root resorption [59].

Low Intensity Pulsed Ultra Sound

Low intensity pulsed ultrasound can enhance healing of connective tissue and dental tissue and has some benefit in cases of root resorption [60]. Low intensity pulsed ultrasound has a beneficial effect on fracture healing. Complex cellular reaction takes place in bone healing. Some of these processes are accelerated by low intensity pulsed ultrasound. Cells like fibroblast, chondrocytes osteoblast and osteocytes are mechanosensitive cells are affected by low intensity ultrasound. Low-intensity pulsed ultrasound acts on osteoblastic cells at a specific stage of differentiation and induces direct anabolic reactions, which results in bone matrix formation [61-63]. Effect of low-intensity pulsed ultrasound (LIPUS) on orthodontically induced root resorption. The LIPUS, decreases osteoclast numbers and activity levels, and increased OPG/RANKL expression ratios [64].

Effect of force and endodontic treatment on root resorption

Rey et al. [65] have suggested that orthodontic management of external apical root resorption should be done by simple mechanical techniques. Light and controlled forces, allowing predictable movements which are physiologically acceptable. Prevention of root resorption can occur by application of light and controlled forces [65].

Teeth with only the cervical third remaining from orthodontically induced external root resorption must remain in one’s mouth with function and esthetics preserved. In these cases, endodontic treatment is not recommended for affected teeth because the pulp is not involved in the process and the post-treatment phase of endodontic therapy might be a complicating factor due to risks of accidental contamination or filling material overflow [66].

The resorption and repair processes during the early stages of retention are balanced, and most of the reparative process occurs after 4 weeks of passive retention after the application of orthodontic force. Frequent orthodontic reactivations should be avoided to promote recovery and repair of root surface damage [67]. Kheirieh et al. [68] reported a case where they did extraoral retrograde root canal filling with CEM cement in a refractory apical lesion due to orthodontically induced extensive EARR [68].

Orthodontic treatment is frequently related to the occurrence of apical root resorption. The concentration of orthodontic forces on the root, especially on the apex, can cause biological changes in the cementum and periodontal ligament, resulting in root resorption [69,70]. Intrusion of incisor creates considerable amount of stress at the apex and is associated with greatest amount of root resorption. Comparable resorption is seen with anterior retraction [71-75].

Resorptive activity is more pronounced after the heavy forces. The reparative processes are different between the light and heavy forces, with marked individual variations. Repair becomes steady after 4 weeks of passive retention following 4 weeks of light force application, whereas most repair occurs after 4 weeks of passive retention following 4 weeks of heavy force application. Root resorption crater volume positively depended on tooth movement and negatively correlated with chronologic age [76]. The volume of root resorption craters formed by buccally directed forces for 12 weeks on the maxillary and mandibular first premolars are directly proportional to the magnitude of the force [77].

Conclusion

Root resorption is almost always associated with orthodontic treatment. At present our understanding about orthodontically induced root resorption is limited. Drugs like doxycyclin and bisphosphonate may be used in future to arrest root resorption. Low intensity ultrasound also has reparative effect on induced root resorption.

References


