

The immortal *Salmonella typhimurium*. (Amnesia among resting cells)

Abstract

Our laboratory contains a collection of thousands of *Salmonella typhimurium* LT2 and LT7 cultures, including mutants (the Demerec collection). Milislav Demerec started the collection of mutants over five decades ago at Carnegie Institute of Genetics, Cold Spring Harbor, L.I., NY. Upon his retirement, Dr. Demerec and the collection moved to Brookhaven National Laboratories. Upon Dr. Demerec's death, the collection (archived in quintuplicate) was divided and distributed to Kenneth Sanderson (who now maintains the official *Salmonella* Center, Calgary, Canada), Philip Hartman, Princeton University, and to Abraham Eisenstark, U. Missouri (see History of Collection, below). Thus, each of us had a complete collection

Keywords: *salmonella typhimurium*, archival storage, viability, aged bacteria

Volume 5 Issue 3 - 2017

Eisenstark A,^{1,2} Mehr JK,¹ Ransdell J^{1,2}

¹Cancer Research Center, USA

²University of Missouri-Columbia, USA

Correspondence: Eisenstark A, Cancer Research Center and Division of Biological Sciences, University of Missouri, 3501 Berrywood Dr, Columbia, MO, USA, Tel (573) 8752255, Fax (573) 4431202, Email Eisenstarka@missouri.edu

Received: June 07, 2017 | **Published:** August 21, 2017

Introduction

These cultures of *Salmonella typhimurium*, both wild type and mutants, were stored mainly for the ability to start fresh cultures and to note any genomic changes that may have occurred during storage. In this report there is no intent to relate to any evolutionary events. To do this would require precisely designed experimental methods. In addition to this brief Mini Review, we now report an example use of these archived *Salmonella* mutants in exploring genomic changes among survivors after the last 4-6 decades of storage in LB agar stabs stored in sealed small tubes (Eisenstark lab), the genomic change results observed in recent experiments (with co-investigators, J. Kian Mehr and J. Ransdell). The successful use of immortal HeLa cells in research initiated deliberation about life in assumed non-viable cells. After decades of stem cell research, scientists are deliberating the use of mammalian skin stem cells even in the initiation of normal baby dogs and perhaps human babies.

Our research efforts have focused on immortality among bacterial cells. In search of survival strategies, our recent studies revealed numerous mutations in *Salmonella typhimurium* cells that had been sealed in LB agar stab vials and stored for the last 4-6 decades. The mutant isolates that were conserved in over 20,000 vials were progeny derived from the same *S. typhimurium* LT2 and LT7 strains (Figure 1).



Figure 1 Sample of storage of archived cultures of *S. typhimurium* LT2 and LT7. Over 2000 vials remain in 10 boxes containing immortal cultures that remain important for genomic studies.

Note – Figure 1 can go here. + legend

Among studies in our laboratory of decades-old archived cultures of resting stage *Salmonella typhimurium* LT2, we have observed a series of diverse mutation patterns. The mutations included deletions, duplications, frame shifts, inversions and transpositions, and losses in carbon and nitrogen metabolism.¹⁻⁶

Most recently, an archived genetically engineered strain of LT2 has been used to target and impede metastasis in human prostate tumor cells.^{7,8}

History of the collection

(This history is now archived by American Society of Microbiology in a series of letters from Joshua Lederberg). To inaugurate the collection, the initial Lilleengen *Salmonella typhimurium* LT1-LT23 was sent to Demerec by Joshua Lederberg, who obtained them through the Swedish Embassy. From these, Demerec and colleagues chose LT2 to construct a gene linkage map, using the newly discovered transducing phage P22.⁹ Phage P22 was found in the glycogenic P22 strain.

Materials and methods

In addition to the use of the archived strains, standard bacteriology materials and methods were used.¹⁰ In a recent experiment, 100 new archived tubes of *S. typhimurium* LT2 were opened, one-third after storage of 19 months, another third after 33 months, and another third after 12 years and nine months. The methods for scoring genomic changes are described under Results.

Results

For this mini review, earlier results will only be summarized, and below are the results from the 100 newly archived tubes. Note that samples were taken at three different time periods. The basis for this was to verify that, with increasing time, there was increasing variance in phenotypes. For decades, experiments using these archived mutants have provided interesting insights on genomic changes that occur in assumed dormancy.^{1-6,11} In our most recent set of experiments, we focused on scoring three specific phenotypic changes:

A. Altered growth spreading when archived

LTs are touched on LB agar plate, as observed in Figure 2.

B. Diversity of phage types among archived cells

Phage Tests

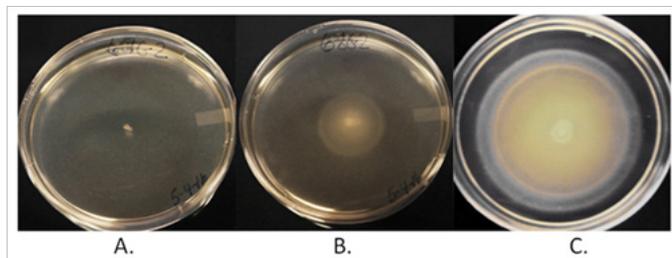


Figure 2 This migration phenotype was chosen because different migration patterns reflected a mutation in any one of a number of genes. There are over 70 genes in wt. *S. typhimurium* LT2. Example of migration patterns among archive mutants.

- A. Loss of motility mutant
- B. Partial loss of motility mutant
- C. LT2 wild type motility

Although phage testing has been a very useful scheme for

Table 1 *Salmonella typhimurium* and bacteriophages used in study our phage stock no. Strain History

WT	<i>S. Typhimurium</i> LT2, Wild-type sequenced strain Contains fels-1, fels-2, gifsy-1 and gifsy-2 from Maloy
2818	Phage-free <i>S. Typhimurium</i> strain received from A. Segall; SDT2739 <i>S. entericaserovar Typhimurium</i> LT2MA8507 (Gifsy-1– Gifsy-2– Fels-2–) Δ Fels-1::frt; characterized further in reference: Gunderson
Phage	
O-1	<i>S. Typhimurium</i> -specific bacteriophage characterized by Kropinski
P22	<i>S. Typhimurium</i> –specific bacteriophage characterized by Kronpinski
P27	<i>S. Typhimurium</i> -supernatant of CRC strain 1949
P53	<i>S. Typhimurium</i> bacteriophage P22 on strain CRC3895
P126	<i>S. Typhimurium</i> supernatant of CRC strain 2010 which contains gifsy-1, gifsy-2
P95	<i>S. Typhimurium</i> supernatant of CRC 1944 Pur E150 LT2
P153	<i>S. Typhimurium</i> Isolated from archival strain 2239 vs. nonarchival ATCC



Figure 3 Example of differences in colony size and morphology when a single archived culture is streaked on an LB agar plate. The larger blotched colonies are not contaminants. They are mutants that have lost typical colony edges.

identifying differences among bacterial strains, it displayed fewer differences in his case than either of the other two tests. The phage that we chose is listed in Table 1. We used these to phage test 65 archived strains.

14028 competition (see Phage P153 lysis pattern below)

The lysis patterns were:

Phage O-1- WT clear plaques 2/65, extra ring of growth in plaque.

Phage P22- Displayed cloudy plaques.

Phage P27- Cloudy plaques.

Phage P53- Cloudy, 14/65 clear, 1/65 extra ring of growth.

Phage P95- Cloudy, 21, 65 clear.

Phage P126- Normal clear plaques, 4/65 no visible plaque, 61/65 tiny clear spot only.

Phage P153- 15/65 normal clear plaques, 2/65 cloudy, 48/65 tiny clear spots. Plaques of Phage P153 only arose when two other strains, *S. typhimurium* 2239 and 14024 were placed in competition with each other, resulting in the induction of a prophage harbored by one of the competing hosts.

C. Diversity of colony sizes of archived cells on LB agar plates.

Discussion

We now know that facultative *S. typhimurium* can survive in sealed agar stabs, with depleting nutritional and oxygen supply. How much longer? The future generations of bacteriologists will be receiving the collection with the mandate to test for immortality and for future genomic changes (the genomic DNA sequences of a few archived strains are on record; also, all the Lillengen strains have been micro arrayed). The concept of mutational change has had an interesting history. Starting with that of Darwin [offspring diversity-natural selection], challenged by Lysenko [environment as mutagen], clarified by Watson and Crick [chromosomal double helix knowledge], [occurrence only upon DNA replication], observations of mutations in resting, non-DNA synthesizing cells by Ryan,¹² and the abundance of mutants among non-growing archived cells.

Among studies in our laboratory of decades old archived cultures of resting stage *Salmonella typhimurium* LT2, we have observed a

series of diverse mutation patterns. The mutations included deletions, duplications, frame shifts, inversions and transpositions, and losses in carbon and nitrogen metabolism. A joy in the study of “Microbial Genetics” is to note “Eureka Moments,” especially those that involve contemporary scientists. Starting with some earlier history, the book by Charles Darwin *On the Origin of Species* (mutation-selection), Lysenko (environment as mutagen), the attack of Lysenkoism by Demerec, Dobzhanski of Lamarckian concepts of the heritability of acquired characteristics. Soon after, molecular geneticists tackled “mutation”. The Luria concept of errors in replicating DNA was refined to include alterations among resting, non-replicating DNA, especially the experiments of Frances Ryan,⁴ who showed that resting cell DNA could incorporate radioactive isotopes and 5-bromuracil. Among our own experiments, studies revealed that numerous mutations continue to occur in *S. typhimurium* LT2. Our immortal *S. typhimurium* LT2 strain, after some genomic engineering, is now a key experimental tool in targeting and impeding prostate tumor metastasis in TRAMP mice.^{12,13}

Aging in higher organisms

Although there are many scientific studies on human aging, such as at the Newcastle University Institute for Ageing in Newcastle, G.B., investigators have not overlooked the need to understand factors at the biochemical and molecular level among the simpler organisms, including bacteria and viruses, such as the information in this Mini Review. We have enjoyed this venture. As might be expected, other microbiologists have provided very interesting data to the aging process in bacteria.^{15–17} Investigators have viewed the question of whether aged organisms, from bacteria to humans, have longevity advantage over parental wild type.¹¹

Conclusion

This Mini Review emphasizes that *S. typhimurium*, stored for several decades in sealed vials, may still be capable of revival and genomic changes.

Acknowledgements

Numerous undergraduate, graduate, post doctorate, and laboratory technicians performed experiments that led to this review. Also, we recognize all individuals who have been involved in establishment of the strain collection, especially Dr. Demerec’s associate, Prof. James Wyche, who brought our share of the collection plus the original stock books, from Long Island to our lab. Currently, we acknowledge the tedious stenographic work through many edits of this mini review, Alycia McGee and Anna Vaclavek.

Author contribution

Professor Eisenstark designed all experiments described or referenced in this mini review. The most recent experiments were performed by co-authors Jackie Kian Mehr and Joey Ransdell.

Conflict of interest

The authors declare no conflict of interest. The funding sponsors had no role in the design of the study; in the collection, analyses,

or interpretation of data; in the writing of the manuscript, and in the decision to publish the result.

References

1. Nishioka Y, Demerec M, Eisenstark A. Genetic analysis of aromatic mutants of *Salmonella typhimurium*. *Genetics*. 1967;56(2):341–351.
2. Holmes AJ, Eisenstark A. The Mutation Effect of Thymine–Starvation on *Salmonella Typhimurium*. *Mutat Res*. 1968;5(1):15–21.
3. Eisenstark A, Eisenstark R, Van Dillewijn J, et al. Radiation—sensitive and recombinationless mutants of *Salmonella typhimurium*. *Mutat Res*. 1969;8(3):497–504.
4. Rabsch W, Helm RA, Eisenstark A. Diversity of phage types among archived cultures of the Demerec collection of *Salmonella entericaserovar Typhimurium* strains. *Appl Environ Microbiol*. 2004;70(2):664–649.
5. Nishioka Y, Eisenstark A. Sequence of genes replicated in *Salmonella typhimurium* as examined by transduction techniques. *J Bacteriol*. 1970;102(2):320–333.
6. Kazmierczak RA, Gentry B, Mumm T, et al. Salmonella Bacterial Monotherapy Reduces Autochthonous Prostate Tumor Burden in the TRAMP Mouse Model. *PLoS one*. 2016;11(8):e0160926.
7. Choe E, Kazmierczak RA, Eisenstark A. Phenotypic evolution of therapeutic *Salmonella entericaserovar Typhimurium* after invasion of TRAMP mouse prostate tumor. *mBio*. 2014;5(4):e01182–e01214.
8. Sanderson KE. Revised linkage map of *Salmonella typhimurium*. *Bacteriol Rev*. 1967;31(4):354–372.
9. Zinder ND, Lederberg J. Genetic exchange in *Salmonella*. *J Bacteriol*. 1952;64(5):679–699.
10. Sutton A, Buencamino R, Eisenstark A. rpoS mutants in archival cultures of *Salmonella entericaserovar typhimurium*. *J Bacteriol*. 2000;182(16):4375–4379.
11. Eisenstark A. Genetic diversity among offspring from archived *Salmonella enterica ssp. entericaserovar typhimurium* (Demerec Collection): in search of survival strategies. *Annu Rev Microbiol*. 2010;64:277–292.
12. Edwards K, Linetsky I, Hueser C, et al. Genetic variability among archival cultures of *Salmonella typhimurium*. *FEMS Microbiol Lett*. 2001;199:215–219.
13. Finkel SE, Kolter R. Evolution of microbial diversity during prolonged starvation. *Proc Natl Acad Sci USA*. 1999;96(7):4023–4027.
14. Lenski RE, Mongold JA, Sniegowski PD, et al. Evolution of competitive fitness in experimental populations of *E. coli*: what makes one genotype a better competitor than another? *Antonie Leeuwenhoek*. 1998;73(1):35–47.
15. Papadopoulos D, Schneider D, Meier–Eiss J, et al. Genomic evolution during a 10,000–generation experiment with bacteria. *Proc Natl Acad Sci USA*. 1999;96(7):3807–3812.
16. Erickson M, Newman D, Helm RA, et al. Competition among isolates of *Salmonella enterica ssp. entericaserovar Typhimurium*: role of prophage/ phage in archived cultures. *FEMS Microbiol Lett*. 2009;294(1):37–44.
17. Gómez JM. Aging in bacteria, immortality or not—a critical review. *Curr Aging Sci*. 2010;3(3):198–218.