

Research Article





Phytochemical screening & evaluation of antidiarrheal mode of action of leaves extracts of Paederia foetida linn

Abstract

For medicinal purpose, decoction of the whole plant P. foetida or leaf, stem bark juice are prepared. Tender leaves are boiled and eaten with chili and salt as food. Tribal people of Chittagong hill tracts of Bangladesh and Aka tribe of Arunachal Pradesh, India consume the leaf juice to treat diarrhea, dysentery, and burns or scalding. It has also been reported that, the powder form of the whole plant is taken by certain tribal communities in India for weakness and rheumatic joint pains. Though the plant has traditional usage as an anti diarrheal, there were limited studies to evaluate its anti diarrheal mode of action. Current study evaluates the preliminary phytochemical screening and anti diarrheal mode of action of aqueous & methanolic extracts of leaves of *P. foetida*. Preliminary phytochemical study revealed the presence of steroid, alkaloid, saponin, flavonoid, phenolic acids and tannin in the leaves extracts. Swiss albino mice of either sex were divided into six groups (five/ group): Group-I served as control and received vehicle (0.9% NaCl-saline) at a dose of 2 ml/kg orally; Group-II served as standard and received loperamide at the dose of 5mg/ kg i.p; Group-III and IV received aqueous extract of Paederia foetida, whereas group-V and VI received ethyl-acetate extract of Paederia foetida at doses of 200 and 300mg/ kg i.p, respectively. Castor-oil induced diarrhea, castor-oil induced enteropooling and gastrointestinal motility test were performed to investigate anti diarrheal activity of these extracts at given groups and concentrations. Acute-toxicity test was also performed at four different doses. The diarrheal episode was inhibited by 39.68% and 49.21% for aqueous extract whereas 47.62% and 53.97% for ethyl acetate extract at the doses of 200 and 300mg/kg respectively. The aqueous extract significantly (p<0.05) decreased the intestinal volume (0.48±0.04ml for 300mg/kg). Ethyl acetate extract also shown the similar result (0.47±0.027ml for 300mg/kg) for the same dose compared to control (0.65±0.03ml) in castor-oil induced enter opooling. Furthermore, decreased intestinal transit (57.1–62.34%) were observed for aqueous extract whereas (59.43-63.50%) for ethyl acetate extract compared to standard (loperamide 5mg/kg). No delayed toxicity was also observed for both of these extracts. Though both the extracts showed potential antidiarrheal activity however the ethyl acetate extract showed more significant activity than the aqueous extract of Paederia foetida. The mode of anti diarrheal activity is due to both the inhibition of gastrointestinal motility and enteropooling activities.

Keywords: anti diarrheal, *Paederia foetida*, castor oil, intestinal transit, acute-toxicity

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Introduction

In developing countries, diarrhea is a major health problem. Acute diarrhea is the leading cause of morbidity and mortality amongst children in developing countries,1 Diarrhea occurs due to a series of imbalanced physiological steps such as increasing motility of the gastrointestinal tract, increased secretion of ions and a decrease in the absorption of fluid and as a consequence loss of electrolytes, particularly Na⁺ and water.² Diarrhea has long been recognized as one of the most important health problem in the developing countries. Worldwide, diarrhea accounts for more than 5-8million deaths in infants and small children less than 5 years each year. According to World Health Organization (WHO) estimation for the year 1998, there were about 7.1 million deaths due to diarrhea.³ In the developing countries, diarrheal disease caused 5.2% of overall deaths and became the sixth leading cause of death in the year of 2007.4 Diarrhea causes one third of the child death in Bangladesh.5 In this scenario, WHO proposed the use of traditional plant in its Diarrhea Control Program.⁶

Paederia foetida (Bengali name- Gondhabadali, Gondhal; English

name- King's Tonic) is a climbing twining shrub belonging to the family Rubiaceae and found all over the Bangladesh.^{7,8} Leaf juice of the plant acts as astringent and is given to children in diarrhea. The leaves extract shows diuretic activity whereas, the roots and barks are widely used to induce nausea and vomiting and in the treatment of hemorrhoids, inflammation of spleen and chest pain and liver pain. Fruits of *P. foetida* is specifically used against toothache. The methanolic extract of P. foetida (leaves) showed significant results on the investigation of anti-microbial, cytotoxic and anti-helmentic activity. 10,11 Whether, ethanolic extract of leaves showed anti-bacterial and hepatoprotective activity. 12,13 The root parts of P. foetida help to treat gastric-ulcer.14 On the other hand, the whole plant exerts anti-nociceptive activity. Phytochemical screening showed that arial parts contain a methyl mercaptane, crystalline keto alcohol, paederolone, aketo compound, paederone and beta-sitosterol, protein, carbohydrates, glucosides essential oils, quinines, alkaloids, iridoid glycosides, sitosterol, stigmasterol, amino acid and volatile oil etc. 15-17 Though the plant has been used traditionally as a remedy for diarrhea and dysentery in many parts of Bangladesh for long time,





a few studies have reported. Afroz et al., ¹⁸ reported that ethanolic extract of *P. foetida* have potential Antidiarrheal activity on limited mice models for ant diarrheal and gastrointestinal motility. Haider et al., ¹⁹ performed a randomized double blind clinical trial of the dried plant extract against a small group of patients with acute shigellosis and found moderate effect in comparison to ampicillin. In addition, recently Swarnamoni et al., ²⁰ reported that ethanolic leaves extracts exhibited significant amelioration of acetic acid induced colitis in mice model.

Therefore, all of the previous study justified the present work for more and in depth investigations of the antidiarrheal activities of the leaves of the plants in different animal models of various extracts from different solvents such as aqueous and ethyl-acetate to explore the antidiarrheal mode of action along with acute toxicity to get in site the mechanism of antidiarrheal action as well as the safety of the plant extracts.

Materials and methods

Plant material

The selected plant (*Paederia foetida*) was collected from a local area (Colonel Hat) of Chittagong district, Bangladesh and authenticated by Dr. Shaikh Bokhtear Uddin, Chittagong University Herbarium, Department of Botany, University of Chittagong, Chittagong, Bangladesh (accession no. SBU 2371. Dt. 11.05.2014. CTGUH).

Preparation of extract

The leaves were shade dried and ground. The ground leaves (300g for each solvent) were soaked in sufficient amount of distilled water and ethyl acetate for two weeks at room temperature with occasional shaking and stirring. In case of aqueous extraction a small amount of sodium azide is added as a preservative. After that both the extracts were filtered through a cotton plug followed by Whitman filter paper No. 1. The mixer was vacuum dried at room temperature to yield semisolid. The extract was then preserved in a refrigerator till further use.

Animals

Swiss Albino mice of either sex, weighting 25 to 30gm was obtained from animal house of Department of Pharmacy, Jahangirnagar University, Dhaka, Bangladesh and housed in polypropylene cages under controlled conditions. The animals were exposed to alternative 12h light/12h dark cycle at an ambient temperature of $27\pm1^{\circ}$ C with a relative humidity of 55%-65%. Animals were allowed free access to drinking water *ad libitum* and pellet diet (obtained from International Centre for Diarrheal Disease and Research, Bangladesh). All the animals were acclimatized for 10days in the laboratory environment prior to the study. For conducting all experiments with animals, we followed the guidelines of Institutional Animals Ethics Committee (IAEC) and before the experiments, study protocols were approved by the University of Science and Technology Medical Ethics, Biosafety and Biosecurity Committee (USTMEBBC) at the Life Science Faculty, University of Science and Technology Chittagong, Bangladesh.

Chemicals

Following chemicals were used for evaluation of ant diarrheal activity of *P. foetida*

1. Loperamide hydrochloride 99.71% (Square Pharmaceuticals Ltd., Bangladesh)

- 2. Castor oil (WELL's Heath Care, Spain)
- 0.9% NaCl/Normal saline (Popular Pharmaceuticals Ltd., Bangladesh) and
- Charcoal preparation (10% activated charcoal in 5% gum acacia).

Phytochemical screening

Both the extracts of *P. foetida* were evaluated by phytochemical qualitative reactions for usual plant secondary metabolites. The phytochemical screening was conducted for the identification of steroids, alkaloids, coumarins, flavonoids, saponins, tannins and phenolic acids.^{21–24}

Following reagents and chemicals were used for phytochemical screenings of the extracts: Dragendroff's reagents (for alkaloids),mg and HCl (for flavonoids); Ferric chloride and Potassium dichromate solutions (for tannins) and saponins with ability to produce stable foam and Libermann-Burchard reagent (for steroids). Phenolic acids derivatives were identified in the studied of water extract of *P. foetida* leaves by the Carrez reagent.

Acute toxicity test

We utilized twenty Swiss Albino mice (4-5weeks old, weighing 20-25g) for investigating acute toxicity and with this we estimated $\rm LD_{50}$ value. Both the aqueous and ethyl-acetate extract was dissolved in saline water and administered orally to four groups of mice, each group containing five mice, at different doses (200, 400, 1000 and 2000mg/kg, b/w). After 24hours, $\rm LD_{50}$ was evaluated by recording mortality.²⁵

Ant diarrheal Activity

Castor oil induced diarrhea

We followed the method of Awouters et al to evaluate the antidiarrheal activity of Paederia foetida.26 The experimental mice were fasted 18h with water ad libitum before experiment. We divided the mice in six groups with each group containing five mice. The groups were marked as- Group I (Control group: treated with saline 2ml/kg body weight orally), Group II (Standard group: treated with loperamide 5mg/kg b. wt. i.p), Group III-IV (treated with 200 and 300mg/kg b. wt. dose of P. foetida aqueous extract i.p.) and Group V-VI (treated with 200 and 300mg/kg b. wt. dose of P. foetida ethyl acetate extract i.p.). After 1hour interval, each animal was given castor oil orally in the purpose of inducing diarrhea. Then the mice were caged with white blotting paper placed at the bottom of the cage and the papers were changed every 1hour for the purpose of ease in stool count. The total number of both dry and wet feces was counted every hour and the process continued up to 4hours. Then the total number of excreted feces was compared with control group whereas total number of diarrheal feces of the control group was considered

Castor oil induced enteropooling

For experimenting castor oil induced enter pooling, we followed the method of Robert et al.²⁷ The animals were fasted for 18 hours prior to experiment with water ad libitum and the fasted mice were grouped as Group I (Control group: treated with saline 2ml/kg body weight orally), Group II (Standard group: treated with loperamide 5mg/kg b. wt. i.p), Group III-IV (treated with 200 and 300mg/kg b. wt. dose of *P. foetida* aqueous extract i.p.) and Group V-VI (treated with 200 and 300mg/kg b. wt. dose of *P. foetida* ethyl acetate extract i.p. with each

group containing five animals. After 1 hour, the animals were given castor oil orally for inducing diarrhea. The with 1hour interval each animal was sacrificed with over dose of chloroform anesthesia and the sacrificed mice were dissected to separate the small intestine from pyloric sphincter to ileocecal junction. The intestine was weighed and the contents were milked out with graduated tube and volume of the contents was measured. Then the intestines were reweighed and weight gap between full and empty stomach were calculated.²⁸

Gastrointestinal motility test

We carried out the experiment of gastrointestinal motility test according to the method described by Mascolo et al.²⁹ The animals were fasted for 18hours prior to experiment with water *ad libitum* and the fasted mice were grouped as- Group I (Control group: treated with saline 2ml/kg body weight orally), Group II (Standard group: treated with loperamide 5mg/kg b. wt. i.p), Group III-IV (treated with 200 and 300mg/kg b. wt. dose of *P. foetida*aqueous extract i.p.) and Group V-VI (treated with 200 and 300mg/kg b. wt. dose of *P. foetida* ethyl acetate extract i.p. with five animals in each group. Before 1hour of treating the groups with respective materials, the animals were given castor oil orally to induce diarrhea. After 1hour of treating the animals

with respective material, the animals were given charcoal preparation (10% charcoal suspension in 5% gum acacia) orally for tracking the movement of the contents afterwards. With one hour interval, the animals were sacrificed by overdose of chloroform anesthesia and the distance traveled by the charcoal preparation from pylorus to caecum was measured and expressed as a percentage of the total distance of the intestine.

Statistical analysis

All results are expressed as (mean±standard error) and to represent the significance, one-way ANOVA (Analysis of Variance) followed by Bonferroni test were used. P value less than 0.05 or 0.001 was considered statistically significant. All data were calculated with SPSS software (version: 16; IBM corporation, New York, USA).

Results

Phytochemical screening

Preliminary phytochemical screening revealed that different components such as steroid, tannins, alkaloids and phenolic acids are present in the extract which is demonstrated in Table 1.

Table I Result of chemical group test of different extracts of P. foetida

| Extracts | Steroids | Alkaloids | Coumarins | Flavonoids | Saponins | Tannins | Phenolic acids |
|----------|----------|-----------|-----------|------------|----------|---------|----------------|
| PFA | - | ++ | - | ++ | ++ | +++ | ++ |
| PFEA | + | + | - | ++ | - | ++ | - |

^{*} PFA, P.foetida aqueous extract, PFEA, P.foetida ethyl acetate extract

Acute toxicity test

For the LD_{50} dose determination, aqueous and ethyl-acetate extract of *P. foetida* leaves were administered up to dose 2000mg/kg body weight. In the 14days monitoring period, we did not observe any visible signs of acute toxicity and mortality.

Castor oil induced diarrhea

In the castor oil-induced diarrhea experiment, the leaves extract of

P. foetida produced a marked ant diarrheal effect in the mice (Table 2). At doses of 200 and 300mg/kg, the aqueous and ethylacetate extract displayed significant (p<0.001) defecation compared to control group. The decreased number of excreted feces after administration of castor oil, for aqueous extract the values are $7.20\pm0.37(200\text{mg/kg})$ and 6.40 ± 0.51 (300mg/kg), whereas, for ethyle acetate extract the values are 7.20 ± 0.37 (200mg/kg) and 6.40 ± 0.51 (300mg/kg). On the other hand, for loperamide, the number of feces decreased to 4.80 ± 0.37 at the dose of 5mg/kg compared to control group (Figure 1).

Table 2 Effect of aqueous and ethyl acetate-extract of P. foetida on castor-oil induced diarrhea in mice

| Groups | | Treatment | Total number of feces | % Inhibition of defecation | Total number of diarrheal feces | % Inhibition of diarrhea |
|-------------------|----|---|-----------------------|----------------------------|---------------------------------|--------------------------|
| Control | ı | Castor oil + Saline (2mL/kg p.o) | 15.4±0.51 | | 12.60±0.51 | |
| Positive | п | Castor oil +Loperamide (5mg/kg i.p) | 7.60±0.51** | 50.67 | 4.80±0.37** | 61.90 |
| P. foetida | Ш | Castor oil + Extract (200mg/kg i.p) | II.8±0.37** | 19.48 | 7.20±0.37** | 39.68 |
| (Aqueous extract) | IV | Castor oil + Extract (300mg/kg i.p) | 10.0±0.71** | 35.06 | 6.40±0.51** | 49.21 |
| P. foetida (Ethyl | ٧ | Castor oil + Extract (200mg/kg i.p) | 12.2±0.74* | 20.78 | 6.60±0.51** | 47.62 |
| Acetate extract) | ۷I | Castor oil + Extract (300mg/kg i.p) | 11.6±0.51* | 24.68 | 5.80±0.40** | 53.97 |

^{*[}Values are expressed as mean±SEM (n=5).*p<0.05, **p<0.001 when compared with control group]

^{* [+++,} Strong intensity reaction, ++, Medium intensity reaction, +, Weak intensity reaction, -, not detected]

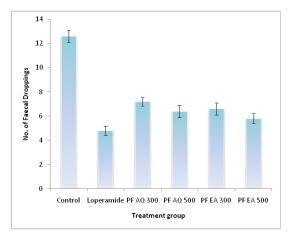


Figure I Number of diarrheal droppings.

*PFAQ, P. foetida aqueous extract; P. foetida ethyl acetate extract

Castor oil induced enteropooling

In case of Castor oil induced enteropooling test, the P. foetida

extracts (200 and 300mg/kg) produced a significant reduction in intestinal weight and volume in a dose dependent manner (Table 3). The intestinal volume was decreased by 17.35% (200mg/kg) and 26.02% (300mg/kg) for aqueous extract whereas for ethyl-acetate extract it was 21.43% (200mg/kg) and 27.55% (300mg/kg). The standard drug, loperamide (5mg/kg), also significantly inhibited (p<0.05) intestinal fluid accumulation which is 37.75% compared to control group.

Gastrointestinal motility test

Both the extracts of *P. foetida* significantly (p<0.001) reduced the gastrointestinal transit. In case of the aqueous extract these were 22.33 ± 1.20 cm (51.1%) for 200mg/kg and 19.33 ± 1.20 cm (62.34%) for 300mg/kg respectively. Whereas, in case of ethyl acetate extract these were 21.42 ± 0.87 cm (59.43%) for 200mg/kg and 19.26 ± 1.20 cm (63.50%) for 300mg/kg respectively. These values clearly revealed that both extracts decreased travel of the charcoal preparation while compared to the control (40.67 ± 3.48 cm) group by inhibition of GI motility (Table 4). However, lope amide (5mg/kg) also produced a significant decrease 14.66 ± 1.45 cm (72.18%) in the propulsion of charcoal preparation through gastrointestinal tract.

 Table 3 Effect of aqueous and ethyl-acetate extract of P. foetida on castor-oil induced enteropooling in mice

| Groups | | Treatment | Weight of intestinal content(g) | Volume of intestinal content(mL) | (%) Inhibition |
|-------------------------|----|--|---------------------------------|----------------------------------|----------------|
| Control | ı | Castor oil + Saline (2mL/kg p.o) | 1.75±0.08 | 0.65±0.03 | |
| Positive | п | Castor oil + Loperamide (5mg/kg i.p) | 1.50±0.03 | 0.41±0.01* | 37.75 |
| P. foetida (Aqueous | Ш | Castor oil + Extract (200mg/kg i.p) | 1.69±0.05 | 0.54±0.25 | 17.35 |
| extract) | IV | Castor oil + Extract (300mg/kg i.p) | 1.55±0.05 | 0.48±0.04* | 26.02 |
| P. foetida | ٧ | Castor oil + Extract (200mg/kg i.p) | 1.72±0.03 | 0.51±0.02 | 21.43 |
| (Ethyl acetate extract) | ۷I | Castor oil + Extract (300mg/kg i.p) | 1.57±0.03 | 0.47±0.02* | 27.55 |

^{*[}Values are expressed as mean \pm SEM (n=5). *p < 0.05, **p<0.001 when compared with control group]

Table 4 Effect of aqueous and ethyl-acetate extract of P. foetida on small-intestinal transit in mice

| Groups | | Treatment | Total length of intestine (cm) | Distance travelled by marker (cm) | Inhibition (%) |
|---------------------------|-----|---------------------------------------|--------------------------------|-----------------------------------|----------------|
| Control | ı | Castor oil+Saline (2mL/kg p.o) | 54.00±0.58 | 40.67±3.48 | |
| Positive | II | Castor oil+Loperamide (5mg/kg i.p) | 52.70±0.88 | 14.66±1.45** | 72.18 |
| P. foetida (Aqueous | III | Castor oil+Extract (200mg/kg i.p) | 52.0±1.15 | 22.33±1.20** | 57.1 |
| extract) | IV | Castor oil+Extract (300mg/kg i.p) | 51.33±0.88 | 19.33±1.20** | 62.34 |
| P. foetida (Ethyl acetate | ٧ | Castor oil+Extract (200mg/kg i.p) | 52.80±0.44 | 21.42±0.87** | 59.43 |
| extract) | VI | Castor oil+Extract (300mg/kg i.p) | 52.77±0.77 | 19.26±1.20** | 63.50 |

^{*[}Values are expressed as mean \pm SEM (n=5).* p<0.05, ** p<0.001 when compared with control group]

Discussion

As part of the traditional therapy, many people from different regions of the world use medicinal plants. Not only whole plant but also some specific parts (such as leaves, bark, etc.) of the plant used as the herbal therapy. As a part of traditional use, leaves of *Paederia foetida* also used for the treatment of diarrhea in some hilly regions of Bangladesh. We found through our preliminary phytochemical screening thatboth aqueous and ethyl acetate extract of *P. foetida* (leaves) possess some compounds like alkaloid, tannin, flavonoids, steroids, saponin etc. which may be responsible for amelioration of diarrhea.

The aqueous and ethylacetate extracts of P. foetida (leaves) displayed the significant antidirrheal activity (p<0.001) at 200mg/ kg and 300mg/kg doses which is comparable to the standard drug loperamide (5mg/kg) and also possess 19.48% and 35.06% inhibition of defecation for the aqueous extract while 20.78% and 24.68% inhibition for the ethyl acetate extract respectively. On the other hand, in case of Castrol oil induced enteropooling, both the extracts displayed significant (p<0.05) effect at 300mg/kg dose as well as reduced the intra-luminal contents volume. These effects, which have direct consequences to reduce water and electrolytes secretion into the small intestine,³⁰ suggested that the extracts may enhance electrolyte absorption from the intestinal lumen consistent with inhibition of hyper-secretion. Hyper-motility characterizes diarrhea where the secretory component is not the causal factor.³¹ Pre-treatment with both the extracts reduced the peristaltic movement or transit of charcoal preparation through the gastrointestinal tract which significantly (p<0.001) indicates that it may be feasible to reduce the frequency of stooling in diarrheal conditions with the leaves extract of P. foetida such as 57.1% and 62.34% inhibition by aqueous extracts whereas 59.43% and 63.50% inhibited by ethylacetate extract at a respective dose of 200mg/kg and 300mg/kg.

Castor oil induces diarrhea by following mechanisms-inhibition of intestinal Na+ K+ ATPase activity, thus reducing normal fluid absorption,³² activation of adenylate cyclase or mucosal cAMPmediated active secretion,33 stimulation of prostaglandin formation and platelet activating factor.³⁴ In the recent time, it is also proposed that nitric oxide also plays an important role in diarrheal effect of castor oil. However, it is an established concept that a compound named ricinoleic acid found in castor oil is responsible for inducing diarrhea for its hyper secretory activity.35,36 Above observations suggested that both the extracts reduced diarrhea by inhibiting peristalsis, gastrointestinal motility and castor oil induced enter pooling in a dose dependent manner. Earlier studies proved that tannins, alkaloids, flavonoids, sterol and/or triterpenes are responsible for anti-dysenteric and antidiarrheal properties of medicinal plants.^{37–39} Hence, phytochemical screening confirmed the presence of tannins, alkaloids and sterol in the leaves of the plant so these bio active compounds might be responsible for the mechanism of action of P. foetida ant diarrheal activity.

Conclusion

The results of this investigation revealed that the aqueous and ethyl acetate extract of *P. foetida* leaves possess significant anti diarrheal properties. The mechanism of antidiarrheal activity of the plant is due to the significant inhibition of gastrointestinal motility and enteropooling activities. Advanced research is needed to isolate specific compounds from both aqueous and ethyl acetate extracts and find out the specific molecule(s) responsible for the observed anti diarrheal activity.

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Conflict of interest

There is no conflict of interest associated with the authors of this paper.

References

- Coker MF, Berky S, Pandou C. New development in acute diarrhea current problem. *Paediatrics*. 1998;24:15–17.
- Rang HP, Dale MM, Ritter JM, et al. *Pharmacology*. London: Churchill Livingstone; 2003.
- 3. Park K. *Park's Text book of Preventive and Social Medicine*. Jabalpur: Banarsidas Bharat publishers; 2015.
- 4. World Health Organization: Top ten causes of death. Fact sheet $N^{\circ}310$. 2007.
- Victora CG, Huttly SR, Fuchs SC, et al. International differences in clinical patterns of diarrheal deaths: a comparison of children from Brazil, Senegal, Bangladesh, and India. J Diarrh Dis Res. 1993;11(1):25–29.
- Syder JD, Merson MH. The magnitude of the global problem of acute diarrheal disease. A review of surveillance data. *Bull World Health Organ*. 1982;60(4):605–613.
- Kantutan Paederia foetida linn medicinal herbs Philippine Alternative Medicine. Stuart Xchange.
- 8. Basu KR, Basu BD. *Indian medicinal plants*. In: Bishen Singh, Mahendra P Singh, editors. 1987;4(2):1796–1799.
- Ghani A. Medicinal plants of Bangladesh with chemical constituents and uses. 2nd ed. Asiatic Society of Bangladesh; 2003:331–332.
- Morshed H, Islam MS, Parvin S, et al. Antimicrobial and cytotoxic activity of the methanol extract of *Paederia foetida* linn. (rubiaceae). J Appl Pharm Science. 2012;2(1):77–80.
- Dey YN, Pal MK. Evaluation of anti-helmintic activity of leaves of Paederia foetida. Int J of Pharma Bio Sci. 2011;2(1):227–231.
- Uddin B, Nahar T, Khalil MI, et al. In vitro antibacterial activity of the ethanol extract of *Paederia foetida* L. (Rubiaceae) leaves. *Bangladesh J Life Sci.* 2007;19(2):141–143.
- Uddin B, Nahar T, Basunia MA, et al. Paederia foetida protects liver against hepatotoxin-induced oxidative damage. Adv Biol Res. 2011;5(5):267–272.
- Reddy KS, Kumar AS, Ganapaty S. Evaluation of anti–ulcer activity of *P. foetida* root extracts in experimentally induced gastric ulcer in rats. *Int J Res Ayur Pharm.* 2011;2(5):1556–1559.
- De S, Ravishankar B, Bhavsar GC. Investigation of the anti–inflammatory effects of Paederia foetida. *J Ethnopharmacol*. 1994;43(1):31–38.
- Osman H, Rahim AA, Isa NM, et al. Antioxidant activity and phenolic content of Paederia foetida and Syzygium aqueum. *Molecules*. 2009;14(3):970–978.
- 17. Blatter E, Caius JF, et al. Indian medicinal plants. 1981;2:1297-1299.
- Afroz S, Alamgir M, Khan MT, et al. Antidiarrheal activity of the ethanol extract of *Paederia foetida* Linn. (Rubiaceae). *J Ethnopharmacol*. 2006;105(1–2):125–130.

- Haider R, Khan AK, Aziz KM, et al. Evaluation of indigenous plants in the treatment of acute shigellosis. *Trop Geogr Med*. 1991;43(3):266–270.
- Das S, Kanodia L, Mukherjee A, et al. Effect of ethanolic extract of leaves of *Paederia foetida* Linn. on acetic acid induced colitis in albino rats. *Indian J Pharmacol*. 2013;45(5):453–457.
- Farnsworth NR. Biological and phytochemical screening of plants. J Pharm Sci. 1966;55(3):225–276.
- Evans WC. Trease and evans' pharmacognosy. London: Sauders Elsevier; 2009:616.
- Harborne JB. A guide to modern techniques of plant analysis. London, New York: Chapman and Hall; 1998.
- Pearson DE. The chemical analysis of foods. New York Churchill Livingstone; 1976.
- 25. Litchfield JT, Wilcoxon F. A simplified method of evaluating dose–effect experiments. *J Pharmacol Exp Ther*. 1949;96:99–113.
- Awouters F, Niemegeers CJE, Lenaerts FM, et al. Delay of castor oil diarrhea in rats; a new way to evaluate inhibitors of prostaglandin biosynthesis. J Pharmacol Pharma. 1998;30(1):41–45.
- Robert A, Nezamis JE, Lancaster C, et al. Enteropooling assay;
 A test for diarrhea produced by prostaglandins. *Prostaglandins*. 1976;11(5):809–828.
- Qnais EY, Elokda AS, Ghalyun YYA, et al. Antidiarrheal activity of the aqueous extract of Punica granatum (Pomegranate) peels. *Pharm Biol*. 2007;45(9):715–720.
- Mascolo N, Izzo AA, Autore G, et al. Nitric oxide and castor oil induced diarrhea. J Pharmacol Exp Ther. 1994;268(1):291–295.
- Shah S. Evaluation of diarrhea: the challenge continues! Part–1. *Indian J Med Sci.* 2004;58(2):75–78.

- 31. Chitme HR, Chandra R, Kaushik S. Studies on antidiarrheal activity of Calotropis gigantean in experimental animals. *J Pharmacol Pharm Sci.* 2004;7(1):70–75.
- Capasso F, Mascolo N, Izzo AA, et al. Dissociation of castor oil–induced diarrhea and intestinal mucosal injury in rat: effect of NG–nitro–L– arginine methyl ester. *British J Pharmacol*. 1994;113(4):1127–1130.
- Pinto A, Autore G, Mascolo N, et al. Time course of PAF formation by gastrointestinal tissue in rats after castor oil challenge. *J Pharm Pharmacol*. 1992;44(3):224–226.
- Mascolo N, Izzo AA, Gaginella TS, et al. Relationship between nitric oxide and plateletactivating factor in castor oil-induced mucosal injury in the rat duodenum. *Naunyn-Schmiedeberg's Arch Pharmacol*. 1996;353(6):680-684.
- Racusen LC, Binder HJ. Ricinoleic acid stimulation of active anion secretion in colonic mucosa of the rat. J Clin Invest. 1979;63(4):743–749.
- Vieira C, Evangelista S, Cirillo R, et al. Effect of ricinoleic acid in acute and subchronic experimental models of inflammation. *Mediators Inflamm*. 2000;9(5):223–228.
- 37. Galvez J, Zarzuelo A, Crespo ME. Antidiarrhoeic activity of Scleroarya birrea bark extract and its active tannin constituent in rats. *Phyto Res.* 1991;5(6):276–278.
- Galvez J, Zarzuelo A, Crespo ME, et al. Antidiarrhoeic activity of Euphorbia hirta extract and isolation of an active flavonoid constituent. *Planta Medica*. 1993;59(4):333–336.
- Longanga OA, Vercruysse A, Foriers A. Contribution to the ethnobotanical, phytochemical and pharmacological studies of traditionally used medicinal plant in the treatment of dysentery and diarrhea in Lomela area, Democratic Republic of Congo (DRC). J Ethnopharmacol. 2000;71(3):411–423.