

Binding of autoantibodies and apoptotic response in erythroid cells in the mouse model of autoimmune hemolytic anemia

Abstract

C57BL/6 mice were given repeated weekly injections of rat erythrocytes to induce autoimmune hemolytic anemia (AIHA), characterized by decreased erythrocyte count and hemoglobin content, presence of membrane bound autoantibodies on circulating erythrocytes and a decline in the bone marrow erythropoietic activity. Erythroid cells in the bone marrow and spleen of AIHA mice were examined for the presence of membrane bound autoantibodies using a fluorochrome conjugated anti-mouse IgG antibody. Apoptotic activity was also assessed flow cytometrically using Annexin V and 7AAD. Results showed that there is significant binding of autoantibodies to both spleen and bone marrow, but the apoptosis is evident only in bone marrow, that too in later stages of differentiation.

Keywords: autoantibody, erythropoiesis, apoptosis, hemoglobin, intraperitoneal injection

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Abbreviations: AIHA, autoimmune hemolytic anemia; DIB, double *in vivo* biotinylation; BM, bone marrow; PS, phosphatidylserine; ROS, reactive oxygen species; MFI, mean fluorescence intensity; SEM, standard error of the mean; SOP, standard operating procedures; SAv-APC, streptavidin-allophycocyanin; NHS, N-hydroxysuccinimide; 7AAD, 7-aminoactinomycin D; FITC, fluorescein isothiocyanate; DMF, dimethylformamide; FBS, fetal bovine serum; EDTA, ethylene diamine tetra acetic acid; DIB, double *In vivo* biotinylation; BNX, biotin-X-NHS

Introduction

Autoimmune hemolytic anemia (AIHA), characterized by the presence of self-reactive autoantibodies against erythrocytes,¹⁻³ can be induced in C57BL/6 mice by repeated intraperitoneal injection of rat erythrocytes⁴⁻⁶ following the Playfair and Clarke Model.⁷ Pathogenesis of AIHA involves two underlying mechanisms, viz, erythrophagocytosis of autoantibody (IgG) coated erythrocytes by macrophages in the reticuloendothelial system in liver and spleen,⁸⁻⁹ and complement mediated lysis of erythrocytes with membrane-bound IgM autoantibodies.¹⁰ AIHA has been extensively studied for the pathogenesis, clinical features and prognosis of the disease.^{3,11} However the modulation of erythropoietic homeostasis and the turnover pattern of circulating erythrocytes are poorly understood. In a recent work from our group, Chatterjee et al.⁶ have studied all the different stages of erythroid system, from early proerythroblasts to reticulocytes, to the different age groups of circulating erythrocytes in AIHA mice⁶ and have unraveled a link between the autoantibody binding and the decline in erythroid cells. The overall decline in erythropoietic activity and erythrocyte count as evidenced in this study was characterized by a higher susceptibility of the more differentiated erythroblasts C and the youngest erythrocytes in circulation.⁶ However the apoptotic activity, which plays a crucial role in the destruction of cells, was not studied in these erythroid cells. In the present study we set onto study a correlation, if any, between the membrane-bound autoantibody and the apoptotic activity in the erythroid cells.

Although erythrocytes lack nucleus, a process similar to apoptosis, termed 'eryptosis' has been identified in erythrocytes.^{12,13} Circulating erythrocytes that are damaged to an extent that would compromise their integrity, even before reaching senescence, undergo a mechanism of a regulated and coordinated suicidal death, termed 'eryptosis'.^{13,14} Eryptosis is very similar to apoptosis, the programmed cell death in nucleated cells,¹⁵ which involves cell shrinkage, membrane blebbing, and plasma membrane scrambling leading to phosphatidylserine (PS) externalization.¹⁴ However since erythrocytes lack a nucleus and organelles, eryptosis also lacks several hallmarks of apoptosis, like mitochondrial depolarization and nuclear condensation. Just like apoptotic cells, PS-exposing eryptotic cells are recognized and rapidly cleared from the circulation through erythrophagocytosis by macrophages.^{16,17} Disturbance of erythrocyte membrane lipids asymmetry and externalization of PS in mature erythrocytes is often triggered oxidative stress.¹⁸ Oxidative stress and high levels of reactive oxygen species (ROS) has been linked to autoimmunity in a variety of diseases¹⁹⁻²¹ and has also been shown as a critical factor in the rat erythrocyte administered mouse model of AIHA.⁶ PS externalization was therefore studied as a marker for both apoptotic and eryptotic activities in erythroid cells in AIHA mice.

Materials and methods

Animals

Inbred C57BL/6 male mice (8-12weeks old, 20-25g body weight) and female Wistar rats (2months old, 250-300g body weight) were used throughout this study. Animals were bred and maintained in microbe free environment in the animal house facility at Jawaharlal Nehru University (JNU), New Delhi or obtained from the National Institute of Nutrition (NIN), Hyderabad. The animals were housed in positive-pressure air conditioned units (25°C, 50% relative humidity) and kept on a 12h light/dark cycle. Water and mouse chow were provided *ad libitum*. All the experimental protocols were conducted strictly in compliance with the Standard Operating Procedures (SOP)

for Institutional Animal Ethics Committee (IAEC) of the Committee for the Purpose of Control and Supervision on Experiments on Animals (CPCSEA), Ministry of Environment, Forest and Climate Change, Govt. of India. The study was duly approved by Institutional Animal Ethics Committee of Jawaharlal Nehru University (IAEC Approved Project Code: 35/2012). All mice were randomly assigned to experimental groups. Experiments were designed so as to use the minimum number of mice.

Reagents and other supplies

Biotin-X-NHS (N-hydroxysuccinimide ester of biotin) was obtained from Sigma Aldrich (St. Louis, MO, USA). Streptavidin-Allophycocyanin (SAv-APC), rat anti-mouse Ter-119-APC, rat anti-mouse CD71-Fluorescein isothiocyanate (FITC) monoclonal antibodies, anti-mouse CD16/CD32 purified and Annexin V-PE recombinant proteins were purchased from BD Biosciences (San Diego, CA, USA) or from Affymetrix eBioscience (San Diego, CA, USA). Goat F(ab')₂ anti-mouse IgG-PE polyclonal antibody, rat IgG1κ-FITC and rat IgG2bκ-APC isotype controls, and 7-Aminoactinomycin D (7AAD) were procured from Affymetrix eBioscience (San Diego, CA, USA). Fetal bovine serum (FBS) was obtained from Hyclone (South Logan, UT, USA). RPMI was obtained from Sigma-Aldrich (St. Louis, MO, USA). HEPES, Dimethylformamide (DMF), Ethylene diamine tetra acetic acid (EDTA) and other analytical reagents were from Sigma-Aldrich (India). All other chemicals were purchased locally and were of analytical grade.

Induction of autoimmune hemolytic anemia in mice

Experimental autoimmune hemolytic anemia (AIHA) was induced in mice following the Playfair and Clarke model of repeated injections with rat erythrocytes.⁴⁻⁷ Rat RBCs derived from tail vein were washed 3 times in phosphate-buffered saline (PBS, pH 7.4) and adjusted to a concentration of 1x10⁹ cells/ml. Mice were given weekly injections of 2x10⁸ rat RBCs intraperitoneally for 5 weeks. Membrane-bound autoantibodies on erythrocytes were detected flow cytometrically by staining with F(ab')₂-anti-mouse IgG-PE.²²⁻²³

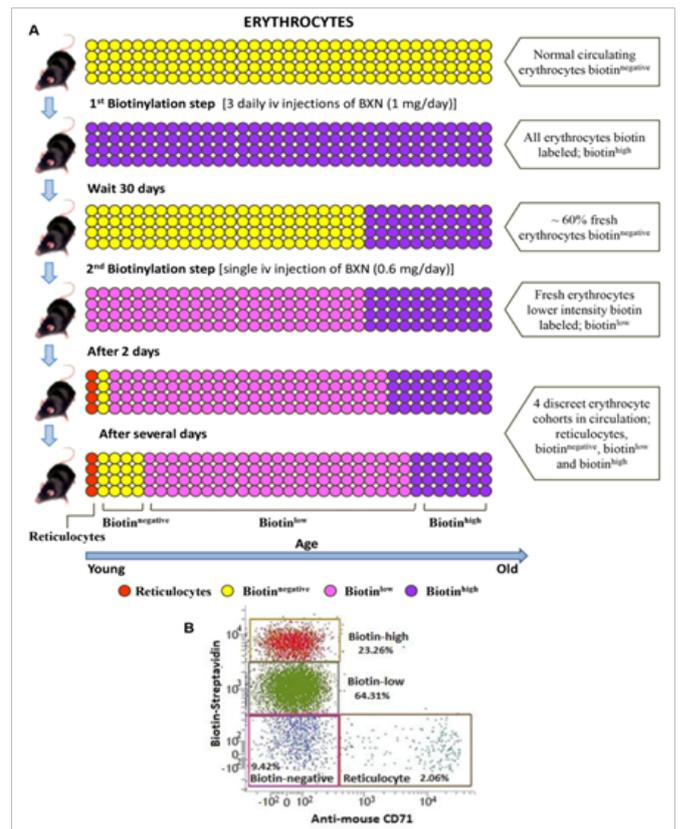
Double *in vivo* biotinylation (DIB) of erythrocytes

Double *in vivo* biotinylation (DIB) of erythrocytes was done as described previously.^{6,24-32} The DIB technique involves two steps of intravenous (*i.v.*) administration of biotin-X-NHS ester (BXN), through the tail vein of mice. In the first step of high intensity *in vivo* biotinylation, 3 daily *i.v.* injections of biotin (1mg BXN dissolved in 20µl of DMF and 250µl of PBS) were given, followed after 30 days, by a low intensity biotinylation with a single lower dose (0.6mg BXN in 12µl DMF and 250µl PBS). This low intensity biotinylation labels the fresh erythrocytes that were released in circulation in the 30-day period between the two biotinylation steps. At any time point after the second biotinylation, biotin intensity on circulating erythrocytes could be analyzed by flow cytometry using streptavidin coupled to any appropriate fluorochrome.^{24,29} Biotin^{negative} erythrocytes in circulation would represent the youngest erythrocytes released in blood after the second biotinylation step. Biotin^{low} erythrocytes would represent the cohort of erythrocytes released in circulation between the first and the second steps of biotinylation, and the biotin^{high} erythrocytes would represent the population of old residual erythrocytes that were present in blood during the first biotinylation.²⁹ The DIB technique is summarized in Supplementary Figure 1.

Erythroid differentiation in bone marrow and spleen

For deriving bone marrow (BM) and spleen cells, mice were

euthanized by CO₂ asphyxiation before the organs were dissected out. BM cells were flushed out of femur and tibia using a 25-gauge needle and re-suspended in RPMI medium with 10% FBS. Single cell suspensions of spleen cells were made by gently teasing the spleen in a small volume of PBS. Splenic and BM cells were strained through a fine sieve, pelleted by centrifugation, and re-suspended at desired concentration in RPMI with 10% FBS. For delineating erythroid precursors at different stages of differentiation, freshly prepared single cell suspensions from BM or spleen were first incubated with anti-CD16/32 monoclonal antibody (Fc block, 1µg/10⁶ cells in 50µl of PBS+2% FBS) for 10mins followed by staining with anti-mouse CD71-FITC and anti-mouse Ter-119-APC for 20mins at 4°C.^{6,32-34}



Supplementary Figure 1 Double *in vivo* biotinylation (DIB) for tracking age related changes on circulating erythrocytes.

C57BL/6 mice were administered three daily *i.v.* doses of 1mg BXN (1st biotinylation step). After a rest for 30days, a single additional dose of 0.6mg BXN was administered (2nd biotinylation step). Blood was collected at different time points and distribution of biotin label on erythrocytes was examined by staining the cells with streptavidin-APC followed by flow cytometry. Time schedule for the experiment is given in panel A. Principle of DIB protocol B is given in panel B, and Biotin label on circulating erythrocytes at different time points is given in panel C. Erythrocyte populations in boxes X, Y and Z represent biotin^{high}, biotin^{low} and biotin^{negative} populations respectively. Values in parentheses represent percentage of cells in different boxes.

Determination of phosphatidylserine (PS) exposing erythroid cells

Circulating erythrocytes and erythroid cells from BM and spleen that express Phosphatidylserine (PS) on their surface were estimated by incubating the cells with fluorochrome conjugated Annexin V recombinant protein along with 7AAD.³² For the *in vitro* assay, mouse erythrocytes (2% suspension) were incubated with serum isolated

from AIHA mice in the 96 well U bottom culture plates for 2hrs at 37°C and 5% CO₂. For BM and splenic assay single cell suspension of erythroid cells were prepared. These cells were washed and re-suspended in Annexin V binding buffer (10mM Hepes, pH 7.4, 140mM NaCl, 2.5mM CaCl₂), followed by staining with 7AAD and Annexin V-PE for 20mins at 4°C. After incubation, erythrocytes were washed and re-suspended in HBS with 2.5mM CaCl₂ and analyzed on flow cytometer immediately. Apoptotic/eryptotic cells were identified as 7AAD⁻ Annexin V⁺ cells.

Flow cytometric analysis

Mouse blood was collected in PBS containing 5mM EDTA and washed 3 times with ice cold saline containing HEPES buffer (10mM, pH-7.4) and 2% FBS. Biotin-labeled erythrocytes (1×10⁶) were stained *ex vivo* with streptavidin-APC and anti-mouse CD71-FITC in dark for 30mins to identify the different age cohorts of erythrocytes, as described before.^{24,29} For enumerating erythroid cells at different stages of differentiation in BM and spleen the technique of double staining with anti-mouse CD71 and anti-mouse Ter119 was used, as described before.^{6,32-34} Briefly freshly prepared single cell suspensions (1×10⁶) from BM or spleen were incubated with anti-mouse CD16/CD32 antibody for 10min followed by staining with anti-mouse CD71-FITC and anti-mouse Ter-119-APC for 20min in dark at 4°C.

To determine apoptotic marker PS externalization, erythroid cells (1×10⁶) were co-stained with Annexin V-PE and 7AAD. To detect autoantibodies bound to erythroid cells, rat serum was used for blocking instead of mouse serum, and cells were stained with F(ab')₂ anti-mouse IgG-PE. The cells were appropriately gated on the basis of streptavidin/CD71 staining or CD71/Ter119 staining, and the corresponding expression was assessed. All the stained cells were washed and analyzed immediately on a flow cytometer. For all the flow cytometric analysis 7AAD was used as viability dye and immunophenotyping was carried out on gated live 7AAD⁻ cells. A minimum of 50,000 events were recorded. All the flow cytometric analyses were performed on a BD FACSVerser flow cytometer (Becton Dickinson, San Jose, CA, USA) using FACSuite software for acquisition and analysis.

Statistical analysis

Each experiment was repeated at least three times. Statistical analysis by Student's t-test and ANOVA was carried out using SigmaPlot software. Data are presented as means±SEM. A level of p<0.05 was accepted as statistically significant.

Results and discussion

Binding of autoantibodies and phosphatidylserine externalization in circulating erythrocytes in AIHA mice

C57BL/6 mice were immunized weekly with rat erythrocytes intraperitoneally for 5-6weeks to induce autoimmune hemolytic anemia (AIHA). Generation of anti-mouse erythrocyte autoantibody was estimated by flow cytometric analysis of erythrocytes stained with anti-mouse IgG/IgM-FITC polyclonal antibodies. The anti-mouse erythrocyte autoantibody level showed a sharp increase of 60% in the MFI of the FITC-conjugated secondary antibody, from 7.10±0.25 in control to 11.46±0.74 in mice after 5 immunizations. The proportion of erythrocytes with membrane-bound anti-mouse autoantibody also increased, almost 3-fold from 3.5±1.11 % to 10.3±1.86 % (Figure 1).

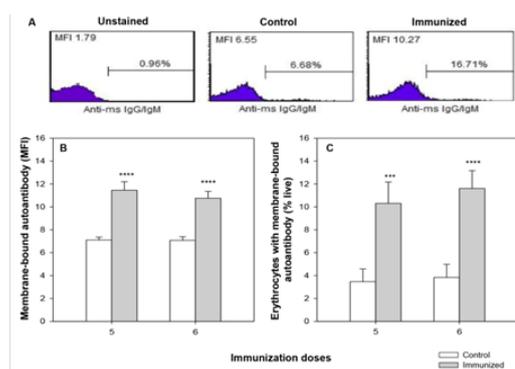


Figure 1 Generation of anti-mouse erythrocyte autoantibody in mice with induced AIHA.

C57BL/6 mice were given intraperitoneal injections of 2×10⁸ rat erythrocytes weekly for 5-6weeks to induce autoimmune hemolytic anemia (AIHA). At intended time points mice were bled and erythrocytes (1×10⁶) were stained with anti-mouse IgG/IgM-FITC polyclonal antibody to assess the presence of membrane-bound autoantibody in erythrocytes in control and immunized mice. Representative flow histograms showing anti-mouse IgG/IgM-FITC staining is given in panel A. Presence of membrane-bound autoantibody on circulating erythrocytes in terms of binding (MFI) and proportion of erythrocytes with membrane-bound autoantibody is given in panels B and C respectively. Each bar in the graph represents mean±SEM of observations from 10mice. ***p<0.005 and ****p<0.001 for comparison of the groups (Student t-test).

Erythrocytes from AIHA mice were examined for phosphatidylserine (PS) externalization. However, Annexin V staining in erythrocytes from control and immunized mice revealed no significant change, probably because PS⁺ erythrocytes are readily removed from circulation. Therefore to assess the extent of PS externalization on erythrocyte membrane after binding of autoantibodies, an indirect method was employed, where erythrocytes from a separate healthy mouse were incubated for 2hrs with serum from AIHA mice, followed by staining with Annexin V-PE and 7AAD. PS⁺ erythrocytes were identified as Annexin V⁺ 7AAD⁻ cells. Simultaneous staining with anti-mouse IgG-FITC would indicate the binding of autoantibodies to these erythrocytes. To examine the PS externalization in different age groups of erythrocytes, the donor mouse was administered double *in vivo* biotinylation (DIB, concept explained in Supplementary Figure 1) before the process. Briefly, in this DIB protocol,^{6,24,29,32} mice were given the two *in vivo* biotin injections at an interval of 30days, and the blood was collected after 10 days of second biotinylation, to get two distinct subgroups of very young biotin^{negative} (<10 days old) and very old biotin^{high} (>40days old) erythrocytes. After incubation with serum from AIHA mice, the erythrocytes were stained with streptavidin APC, CD71-FITC, Annexin V-PE and 7AAD. Different age groups of erythrocytes were gated and proportion of PS⁺ cells were determined in each of these cohorts. Results in Table 1 revealed a marginal, yet significant increase in the proportion of PS⁺ cells in the younger cohorts (biotin^{negative} and reticulocytes) of erythrocytes in mice with induced AIHA. The extent of PS externalization in the aged erythrocytes remained more or less unchanged. This pattern was similar to the extent of autoantibody binding in these cells.

Binding of autoantibodies and phosphatidylserine externalization in erythroid cells in bone marrow and spleen of AIHA mice

Bone marrow (BM) and spleen, the two prime erythropoietic sites

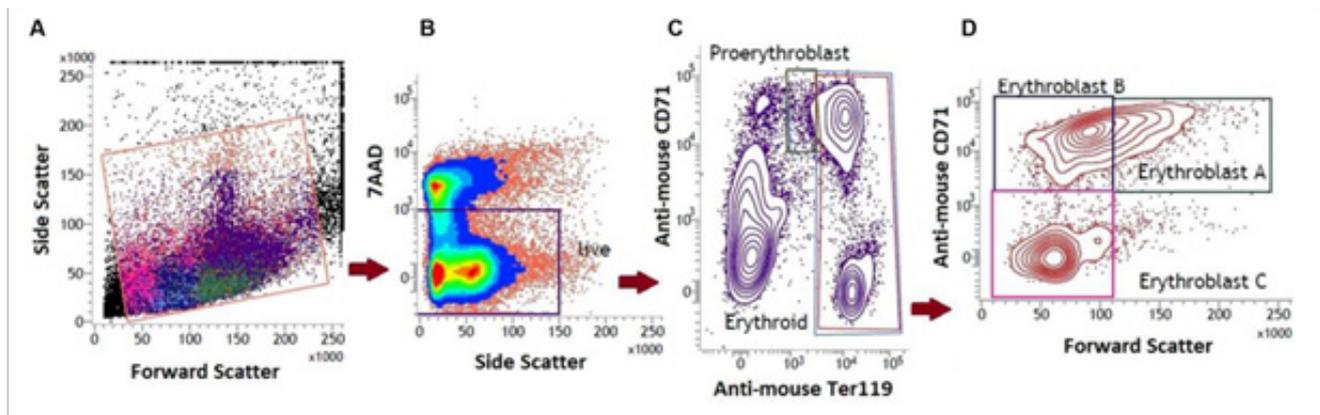
in adult mice, were examined for the presence of membrane-bound autoantibody and apoptotic activity. Erythroid line of differentiation in bone marrow and spleen starts with the early progenitor pro-erythroblasts that are derived from the pluripotent stem cells. Pro-erythroblasts further differentiate in successive stages viz erythroblast A, B and C.^{35,36} Differential expression of Ter119 and CD71 molecules on erythroid cell surface in BM and spleen were used to delineate four distinct stages of erythroid differentiation: early pro-erythroblasts (Ter119^{med} CD71^{high}), early basophilic erythroblasts (Ter119^{high}

CD71^{high} FSC^{high}, erythroblast A), late basophilic, polychromatic and orthochromatic erythroblasts (Ter119^{high} CD71^{med} FSC^{low}, erythroblast B) and orthochromatic erythroblasts with mature erythrocytes (Ter119^{high} CD71^{low} FSC^{low}, erythroblast C), as described elsewhere^{6,32-34} (Supplementary Figure 2). Total erythroid population in BM and spleen was delineated and enumerated by setting an inverted L-shaped gate in the flow histogram that includes all the above mentioned four stages of erythroid differentiation (Supplementary Figure 2).^{33,34}

Table I Phosphatidylserine (PS) externalization in erythrocytes

Erythrocyte age cohorts	PS+Erythrocytes (%)		Autoantibody (MFI)	
	Control sera	Immunized sera	Control sera	Immunized sera
Total erythrocytes	1.06±0.31	1.18±0.08	24.7±1.20	32.7±2.12*
Biotin ^{high} (SAV ^{high} CD71 ^{low})	2.86±0.95	2.73±0.19	29.3±1.45	33.7±1.41
Biotin ^{low} (SAV ^{med} CD71 ^{low})	0.88±0.45	0.96±0.11	22.3±0.88	30.2±2.08*
Biotin ^{negative} (SAV ^{low} CD71 ^{low})	0.27±0.02	0.45±0.04*	21.7±1.45	31.7±2.49*
Reticulocyte (SAV ^{low} CD71 ^{high})	1.32±0.19	2.35±0.17**	67.3±2.40	98.3±4.58***

Mice were given weekly *i.p.* injections of rat erythrocytes for 5weeks to induce AIHA, blood samples were collected thereafter, and serum isolated. A separate group of mice were given biotin injections as per schedule in Supplementary figure 1, 10days after the 2nd biotinylation, erythrocytes were collected and incubated *in vitro* with the sera from control and AIHA mice. Erythrocytes were then washed and stained with streptavidin-APC, anti-mouse CD71-FITC and Annexin V-PE. Erythrocytes of different age groups were gated and the PS externalization and presence of membrane-bound autoantibody was assessed in each of these groups. Each value represents mean±SEM of observations. n=3 control and 7 immunized sera. *p<0.05, **p<0.01 and ***p<0.005 for comparison of the groups (Student t-test).



Supplementary Figure 2 Erythropoietic activity in the bone marrow (BM) and spleen in mice with induced AIHA.

Mice were given intraperitoneal injections of 2x10⁸ rat erythrocytes weekly for 5-6weeks to induce AIHA. Mice were sacrificed 3days after 5th and 6th immunizations and their BM and spleen cells were harvested. Cells isolated were stained with anti-mouse CD71-PE, anti-mouse Ter119-APC and 7AAD, and the proportions of erythroid cells were determined. Four distinct stages of erythroid differentiation could be identified within an inverted 'L' shaped gate in the flow diagram. These include early proerythroblasts (Ter119^{med}CD71^{high}), early basophilic erythroblasts or erythroblasts A (Ter119^{high}CD71^{high}FSC^{high}), late basophilic polychromatic and orthochromatic erythroblasts or erythroblasts B (Ter119^{high} CD71^{med}FSC^{low}), and orthochromatic erythroblasts with mature erythrocytes or erythroblasts C (Ter119^{high}CD71^{low}FSC^{low}).

In our previous study,⁶ examination of erythroid cells for membrane-bound autoantibody revealed a significant increase in the autoantibody binding in both BM and spleen in the total erythroid population (Figure 2) as well as the different stages of differentiation (more significant in erythroblasts B and C). As such presence of membrane-bound autoantibodies on erythroid cells, particularly in BM, has been regarded as a critical factor for the decline in overall erythropoietic activity in the AIHA mice. This observation was in line with several earlier studies on SLE (systemic lupus erythematosus), where presence of inhibitory autoantibody on BM progenitor cells had been linked to the development of hypoplasia, and even pure red cell

aplasia.^{37,38} In the present study erythroid cells in BM and spleen were examined for their apoptotic response. Annexin-V staining showed a marked increase in the proportion of apoptotic cells (Annexin-V⁺ 7AAD⁻) in BM erythroid cells (from 4.20±0.27% in control to 5.16±0.40% in AIHA mice; Figure 3). This increase was also evident in the erythroblasts B and C subpopulations in BM, which showed a 30% increase in the proportion of apoptotic cells (Figure 4). The apoptotic activity in BM therefore followed the same pattern as observed in case of membrane-bound autoantibody. The splenic erythroid cells however showed no change in apoptotic response (Figure 3) & (Figure 4).

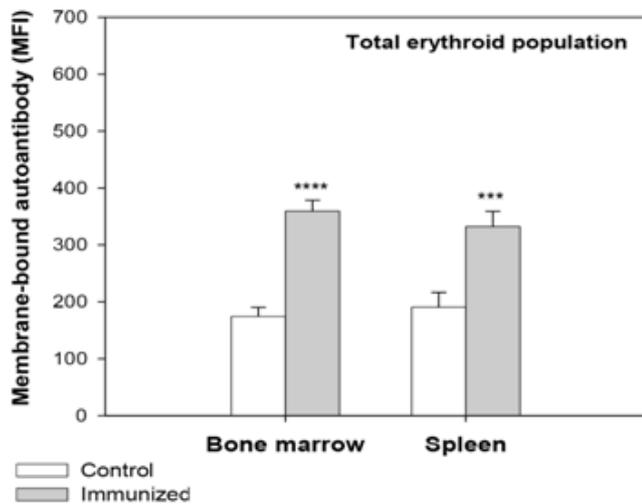


Figure 2 Presence of membrane-bound anti-mouse autoantibody on erythroid cells of bone marrow (BM) and spleen.

Mice were given intraperitoneal injections of 2×10^8 rat erythrocytes weekly for 5-6 weeks to induce AIHA. Mice were sacrificed and their BM and spleen cells harvested. Cells isolated were stained with anti-mouse CD71-PE, anti-mouse Ter119-APC and 7AAD, to determine the proportion of live erythroid cells. The erythroid cells were co-stained with F(ab')₂ anti-mouse IgG-PE to detect the presence of autoantibodies. Presence of membrane-bound autoantibody in the total erythroid populations of BM and spleen after 5 immunizations is shown above (ANOVA test for both BM and spleen, $p < 0.001$). Each bar on the graph represents mean \pm SEM of observations. $n = 4$ control and 6 AIHA mice. *** $p < 0.005$ and **** $p < 0.001$ for comparison of groups (Student t-test).

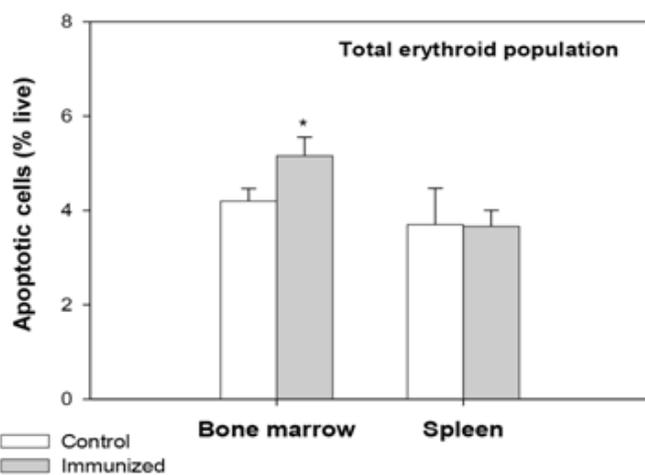


Figure 3 Apoptotic response in erythroid cells of BM and spleen in mice with induced AIHA.

Mice were given intraperitoneal injections of 2×10^8 rat erythrocytes weekly for 5-6 weeks to induce AIHA. Mice were sacrificed and their BM and spleen cells were harvested. Cells isolated were stained with anti-mouse CD71-FITC, anti-mouse Ter119-APC and 7AAD, to determine the proportion of live erythroid cells. Erythroid cells were incubated with Annexin V-PE and the apoptotic cells were identified as Annexin V⁺7AAD⁻ cells. Proportion of apoptotic cells in the whole erythroid population in BM and spleen is given above. Each bar on the graph represents mean \pm SEM of observations. $n = 4$ control and 6 AIHA mice. * $p < 0.05$ for comparison of groups (Student t-test).

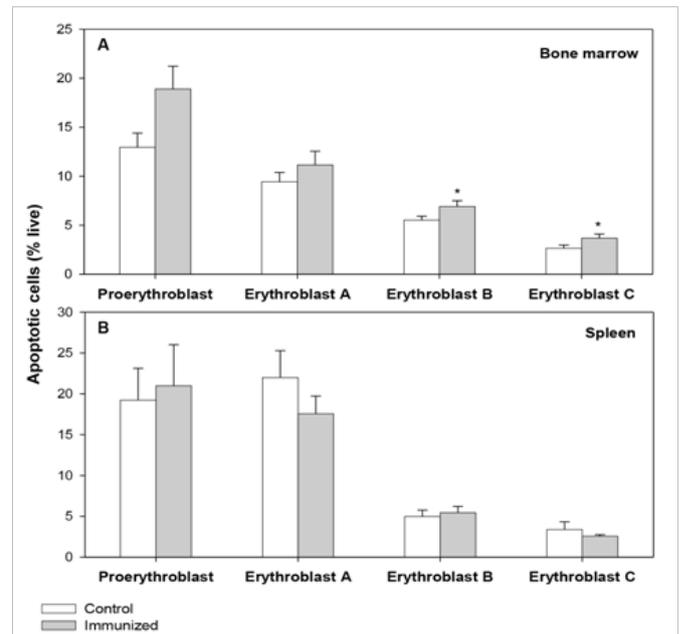


Figure 4 Apoptotic response in erythroid cells of BM and spleen at different stages of maturation.

Mice were given intraperitoneal injections of 2×10^8 rat erythrocytes weekly for 5-6 weeks to induce AIHA. Mice were sacrificed and their BM and spleen cells were harvested. Cells isolated were stained with anti-mouse CD71-FITC, anti-mouse Ter119-APC and 7AAD, to determine the proportion of live erythroid cells. Erythroid cells were incubated with Annexin V-PE and the apoptotic cells were identified as Annexin V⁺7AAD⁻ cells. Apoptotic cells in erythroid populations at different stages of maturation in BM and spleen are shown in panels A and B respectively. Each bar on the graph represents mean \pm SEM of observations. $n = 4$ control and 6 AIHA mice. * $p < 0.05$ for comparison of groups (Student t-test).

Conclusion

Autoimmune anemia is characterized by the presence of anti-erythrocyte autoantibodies,⁵ which is evident from the presence of membrane-bound autoantibodies on circulating erythrocytes. These autoantibodies seem to be directed not only against the mature erythrocytes, but also the erythroid precursors at every stage of differentiation.⁶ Since binding of autoantibody on the erythroid membrane could cause a membrane perturbation, it might also trigger an apoptotic response in these erythroid cells. Additionally autoantibody binding has been shown to be linked with enhanced oxidative stress,^{6,19-21} and oxidative stress on the other hand, is known to be a critical factor triggering apoptosis.¹⁸

In this study apoptotic activity in erythroid progenitors and eryptotic activity in mature non-nucleated erythrocytes were examined as a consequence of autoimmune response in AIHA mice. Results showed increased apoptotic/eryptotic activities in erythroid cells with the highest titer of membrane-bound autoantibodies. Although membrane-bound autoantibodies could be evidenced in each and every stage of erythroid system, this selective activation of apoptosis/eryptosis in erythroid cells could indicate a critical density of autoantibodies that is necessary to elicit the apoptotic response. As such, only the cells with highest deposition of autoantibodies on their membrane show significant apoptotic activity, which might be a causative factor for their selective destruction. Splenic erythroid

cells in AIHA mice however, did not show any apoptotic activity in spite of very high titer of autoantibodies on their membrane. Similar trend was also observed for the intracellular ROS generation in these cells.⁶ The splenic population might harbor protective mechanisms to counteract the perils of autoantibody binding that might also be linked to the activation of stress response in spleen erythroid cells in conditions of BM hypoplasia/aplasia and acute anemia. This requires further studies.

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Conflict of interest

The author declares no conflict of interest.

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