Protein S100B: a potential biomarker for brain damage in paediatric patients with congenital heart disease?

Abstract

Objective: S100B protein has been implicated as a biomarker of brain injury in several clinical conditions. The aim of this study is to determine and to correlate concentrations of the protein within preoperative conditions in pediatric population with congenital heart disease: with perinatal suffering, neurological pathological antecedents, arterial O2 saturation and oxidative stress parameters values.

Material and Methods: We measured serum levels of S100β, lactate, nitrates, Nitrites, Malondialdehyde and total antioxidant capacity in the pre-operative period of 72 pediatric patients with congenital heart disease: 47 of them with perinatal neurological history (fetal suffering) and 25 of them without such antecedents.

Results: Patients with perinatal suffering and neurological background showed higher maximum lactate levels after cardiac surgery [3.2 (2.3-4.8) mmol/L; p<0.005]. As well as higher serie concentration of S100β protein [0.0188 (0.0155-0.0478) ug/l; p=0.0.0.019] (micrograms), when compared to the group without such clinical history. An inversely proportional correlation was observed when S100β concentration was compared with total antioxidant capacity. At serum levels of 0.188ug/l (nanograms) of S100β the sensitivity/specificity curve (ROC) was respectively 51% and 72%.

Conclusion: Although we do not know the true clinical significance and the specific anatomical substrate that induces the release of the S100β it can be considered as a biomarker of brain damage, only if complemented with other parameters such as lactate and total antioxidant capacity.

Keywords: biomarker, neurological damage, congenital heart disease, S100B

Abbreviations: NO, nitrogenmonoxide; NO2, nitrite anion; MDA, malondialdehyde; CPB, cardiopulmonary bypass; TAC, total antioxidant capacity; PBP, pulmonary bypass; CVE, cerebral vascular events; ALZ, alzheimer

Introduction

Congenital heart disease is the commonest malformation in paediatrics, with a prevalence of 6 to 8 cases for each 1000 newborns alive. The advances in the diagnosis and the surgical techniques and the peri-operative handling, have permitted that many patients reach adult life; but the possibility of damage to the central nervous system continues to be one of the most feared associated morbidities in cardiovascular surgery.1,2 The presence of a vast variety of neurodevelopmental alterations has been identified; up to a 50% of children, with corrected congenital heart disease, but unfortunately, we cannot predict it’s possible appearance: the diagnosis depends on neurological examinations, neuropsychiological studies, imagenological scanning such as: computed tomography, PET/CT, AMR with or without spectroscopy, SPECT, among others.

These studies cannot be accomplished immediately, because of hemodynamic instability, critical condition of patients, not availability of the infrastructure or the cost itself of the study. Besides, not always these studies can supply always the desired information regarding the sensibility or the specificity in clinical terms to evaluate and quantify the exact measure of the neurological insult and neither can predict it’s outcome.3,4

The need for a biomarker is therefore needed not only to determine neuronal damage but to predict it’s evolution and outcome. The protein S100β, abundant in the brain, produced by astrocytes in physiological conditions has been used in different clinical situations: CET, cerebral ischemia, cerebral tumors, neurodegenerative disorders or during chronical inflammatory cerebral disease.5-11 In cardiac arrest as well elevation of S100β as well as cardiopulmonary bypass associated to neurological suffering because of cardiovascular surgical complications.12-13 This increment of sensical values of the S100β has been mentioned as evidence of permeability of the BBB and CI.14,15

The objective of the present study was to determine and to correlate the serum concentration of S100β protein in pediatric population with congenital heart disease with perinatal neurological antecedents (particularly fetal suffering) and neurological antecedents during developmental period prior to surgery, with oxygen saturation, oxidative stress in a period before it’s surgical correction of the cardiopathy.

Material and methods

This study is analytical, prospective and transversal study of patients admitted to the cardiopедiatry department with an age <18...
years of age, that were submitted to surgical elective correction of their congenital heart disease with or without cardiopulmonary bypass in a period of 3 months.

The exclusion criteria were antecedent of previous cardiothoracic surgery or emergency cardiac catheterism. Clinical history was completed in all patients included, considering gender, anthropometric measures, O₂ saturation presurgical, pathological and syndromatical antecedents, perinatal-history PBP requirement, time under bypass, aortic clamping, special emphasis in fetal suffering and Appgar score (-5) after birth, cardiac arrest, neurodevelopment, metabolic issues and convulsions.

Patients were divided in one group with neurological perinatal antecedents and a second group without such antecedents. The evolution was monitored until it’s withdraw from hospital with particular interest in neurological morbidity.

Sample processing

Peripheral blood samples were obtained from the patients included in the study, previous to cardiac corrective surgery. Such samples were extracted through venous puncture and centrifugated at 3000 rpm during 15´ at room temperature. Aliquots were obtained from the serum and proceeded to freeze the samples down to -80°C until their analysis. Levels of S100β were measured as well as concentration of Malondialdehyde, nitrates, nitrates and TAC.

Concentration of S100β

The concentration of S100β was measured by ELISA technique with two incubation periods with a total time of 120 minutes. During the first incubation period a monoclonal specific antibody was added (biotinilated anti S100β antibody) for a period of 60 minutes. Afterwards, it was added conjugated HRP-estreptavidine; which after the first incubation period a monoclonal specific antibody was added (biotinilated anti S100β antibody) for a period of 60 minutes. The reaction was stopped by the addition of an acid solution and the absorbance of the resulting product was measured: which results proportional to the concentration of S100β.

A standard curve was constructed representing the values of absorbance against concentrations that were expressed in micrograms/l.

Malondialdehyde (MDA)

This molecule, was determined in serum by capillary zone electrophoresis, the sample was desproteinized, with cold methanol, in a proportion 1:1.Proceeded to centrifuge at 16000xg during 15´ and purified with nitrocellulose membrane filters of 0.22um (Millipore, Billerica, MA, USA).Then it was diluted 1:10 with cold sodium hydroxide 0.1 M for 2 minutes, distilled water for another 2 minutes and buffer for 4 minutes. The concentration of MDA was expressed in UM and through a standard curve.

Nitrites and nitrates

The quantification of nitrites and nitrates was performed at 37°C. We added 100uL of the sample+100ul of saturated vanadium chloride in chlorhidric acid 1M.Homogenized vigorously and then added 100uL of a mixture 1:1 of sulfanilamide 2% recently prepared in chlorhydric acid at 5% conc. and N-(1naftil)-etilendiamina in 0.1% distilled water. Gently homogenized and immediately read at 540nm. In both determinations the 0 absorbance was adjusted with a mixture of reactive. The concentrations of nitrites and nitrates were determined through respective curves sodium nitrite and nitrate grade HPLC humidity free in a range of 0 to 500 pmoles/mL.

Total antioxidative capacity (TAC)

The CUPRAC methodology was applied in an assay tube 1ml of CuCl₂ 1x10-2M+1ml of neocuproine 7.5x10-3M and a solution of ammonium acetate at a pH of 7,these as a chromogenic agent. Afterwards the extracts prepared in water (1.1xml) to complete a final volume of 4.1 ml. The assay tubes were sealed and after 30 minutes of incubation an absorbance of 450 nm was registered against a reactive blank. In order to determine equivalence in Trolox units (umoles per gram of extract) we used a value of ETrolox=1.67x104Lmol-cm-1.

Statistical analysis

We used conventional statistical analysis of central tendency to resume the main characteristics of the sample. In the exploratory analysis, the numerical data had a different distribution than the normal standard. The comparison of the numerical variables between individuals with neurological perinatal antecedents with the ones that did not had them, was done with the test of U-Mann Whitney and the data are presented as median and percentiles 25 and 75. The categorical variables were analyzed with the test of Fisher when required and are presented as absolute frequencies and proportions. The statistical significance was established in p<=0.05.To study the correlation between S100β and ordinal variables and discrete quantitative it was used the Spearman correlation.

S100β as a dependent variable was submitted to logarithmic transformation, after confirming the corresponding suppositions; we applied a linear regression analysis in order to identify the variables that would explain better the risk of developing neurological issues in the studied groups.

The model was constructed one variable at a time. The final model included variables with biological relevance, with statistical relevance or both. The co variables included in the model were the ones that presented a coefficient =0.3 with a p<0.15 complying with this and adjusting to one variable the TAC.

The reference values of S100β were calculated in pediatric population with perinatal antecedents by means of a ROC curve (sensibility and sensitivity curve) for differences between groups with and without perinatal antecedents (basically hypoxic). The data were analyzed with the program SPSS version 18.0 (SPSS, Chicago Il) and STATA version 11.0.
Results

72 Patients were included for the final analysis, which were divided in 2 groups: 47 of them with neurological perinatal antecedents (52% of them masculine), and 25 of them without these antecedents. The demographic and clinical characteristics are shown in Table 1.

We can observe that the group with neurological perinatal antecedents had significant differences in presence of congenital cyanotic heart disease [61.7%(n=29); p=0.0001], pathological antecedents [68% [68%(n=32); p=0.009], brain damage suspected [45%(n=21); p=0.001], chronic hypoxia [79%(n=79% p<0.000], and cardiac arrest event [23%(n=11) p=0.032], compared to the group without perinatal antecedents.

The group with perinatal antecedents revealed higher statistically significant maximum post surgical lactate [3.2(2.3-4.8) mmol/L; p=0.005] and serum levels of protein S100β [0.0188(0.0155-0.0478) ug/L; p=0.19] compared with the group without neurological perinatal antecedents. (Table 2)

Table 1 Characteristics demographics of study groups

<table>
<thead>
<tr>
<th>Variables</th>
<th>With perinatal antecedents (n=47)</th>
<th>Without perinatal antecedents (n=25)</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td>Gender</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Male n (%)</td>
<td>24 (51%)</td>
<td>13 (52%)</td>
<td>0.94</td>
</tr>
<tr>
<td>Female n (%)</td>
<td>23 (49%)</td>
<td>12 (48%)</td>
<td></td>
</tr>
<tr>
<td>Age (months/years)</td>
<td>3 (1-10)</td>
<td>3 (1-10)</td>
<td>0.648</td>
</tr>
<tr>
<td>Weight (Kg)</td>
<td>12 (5.2-21)</td>
<td>12.9 (7.3-43.0)</td>
<td>0.271</td>
</tr>
<tr>
<td>Congenital cyanotic heart disease</td>
<td>29 (61.70)</td>
<td>8 (32)</td>
<td>0.016</td>
</tr>
<tr>
<td>O₂ Saturation</td>
<td>78 (70-93)</td>
<td>91 (85-94)</td>
<td>0.004</td>
</tr>
<tr>
<td>Cardiopulmonar bypass</td>
<td>41 (87.23)</td>
<td>23 (92)</td>
<td>0.54</td>
</tr>
<tr>
<td>Cardiopulmonar bypass (min)</td>
<td>93 (40-129)</td>
<td>90 (60-126)</td>
<td>0.543</td>
</tr>
<tr>
<td>Transoperatory complications</td>
<td>20 (42.55)</td>
<td>6 (24)</td>
<td>0.119</td>
</tr>
<tr>
<td>Postoperatory complications</td>
<td>35 (74.47)</td>
<td>15 (60)</td>
<td>0.205</td>
</tr>
<tr>
<td>Sindromatic antecedents</td>
<td>9 (19)</td>
<td>5 (20)</td>
<td>0.931</td>
</tr>
<tr>
<td>Pathological antecedents</td>
<td>32(68)</td>
<td>9 (36)</td>
<td>0.009</td>
</tr>
<tr>
<td>Fetal Suffering</td>
<td>11 (23.40)</td>
<td>3 (12)</td>
<td>0.244</td>
</tr>
<tr>
<td>Pre-eclampsia</td>
<td>4(9)</td>
<td>3 (12)</td>
<td>0.463</td>
</tr>
<tr>
<td>Eclampsia</td>
<td>2(4)</td>
<td>0</td>
<td>0.423</td>
</tr>
<tr>
<td>Apgar&lt;7</td>
<td>25(53)</td>
<td>15(60)</td>
<td>0.58</td>
</tr>
<tr>
<td>Seizures</td>
<td>2(4)</td>
<td>2(8)</td>
<td>0.433</td>
</tr>
<tr>
<td>Metabolic disorders, n (%)</td>
<td>5 (11)</td>
<td>2(8)</td>
<td>0.537</td>
</tr>
<tr>
<td>Brain damage n (%)</td>
<td>21(45)</td>
<td>2(8)</td>
<td>0.001</td>
</tr>
</tbody>
</table>

Table 2 Biochemical features of study groups

<table>
<thead>
<tr>
<th>Variable</th>
<th>With antecedents perinatal (n=47)</th>
<th>Without antecedents perinatal (n=25)</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td>Minimum lactate (mmol/L)</td>
<td>1.4 (1.1-1.7)</td>
<td>1.2 (1.1-1.4)</td>
<td>0.076</td>
</tr>
<tr>
<td>Maximum lactate (mmol/L)</td>
<td>3.2 (2.3-4.8)</td>
<td>2.1 (1.5-2.6)</td>
<td>0.005</td>
</tr>
<tr>
<td>Protein S100β (ug/L)</td>
<td>0.0188 (0.0155-0.0478)</td>
<td>0.0155 (0.0144-0.0211)</td>
<td>0.019</td>
</tr>
<tr>
<td>Malondialdehyde, MDA (pmoles/mL)</td>
<td>0.427 (0.181-1.021)</td>
<td>0.448 (0.139-0.933)</td>
<td>0.438</td>
</tr>
<tr>
<td>TAC (moles/L)</td>
<td>482.7 (297.0-615.9)</td>
<td>459.4 (381.2-600.3)</td>
<td>0.827</td>
</tr>
<tr>
<td>[NO] (pmoles/mL)</td>
<td>10.2 (7.2-15.0)</td>
<td>12.3 (6.3-17.0)</td>
<td>0.594</td>
</tr>
<tr>
<td>[NO₂] (pmoles/mL)</td>
<td>42.9 (21.4-92.0)</td>
<td>51.3 (34.9-71.9)</td>
<td>0.35</td>
</tr>
<tr>
<td>[NO₃] (pmoles/mL)</td>
<td>318.9 (208.9-491.7)</td>
<td>287.6 (70.0-366.1)</td>
<td>0.172</td>
</tr>
</tbody>
</table>
The correlation of S100β with biochemical parameters shows significant statistical differences between the group with individuals with neurological perinatal antecedents when correlated with TAC (Spearman’s Rho= -0.516, p=0.0002), meaning that patients with perinatal antecedents mainly hypoxic with their cardiopathy with a descent in their levels of TAC presented elevated levels of S100β. (Table 3) Which is represented in Figure 1. We did not observe any other biochemical correlation of S100β in this work.

To study the relation and potential effect of confusion factors, we applied a linear regression model in the group with neurological perinatal antecedents. The only variable which maintained a significant statistical relation with levels of S100β was TAC (β=-0.864, 95% confidence interval =-1.001 to -0.720, R²=0.6697, p<0.000).

We posted the reference values of S100β in our total Pediatric population, by age <1 year and >1 year, by classification with and without perinatal antecedents. (Table 4) The Cohort dots of S100β were determined by ROC curves to determine the value of the protein in the presence or not of such perinatal neurological antecedents. (Table 5) The mayor area below the curve was AUC= 0.6677 (95% confidence interval 0.546-773) corresponding to a value of S100β of 0.188 ug/L. The sensibility and specificity of this value to identify patients with perinatal antecedents was 51% and 72% respectively. (Figure 2)
Protein S100B: a potential biomarker for brain damage in paediatric patients with congenital heart disease?

The literature has mentioned protein S100β as a possible biomarker, it was discovered by Moore in 1965 when he isolated a subcellular fraction of Bovine brain. This fraction was called S100 because it’s constituents are soluble in ammonium sulfate 100% saturated at neutral pH. This protein forms part of a family of about 25 members (E-F Hand proteins Calcium binding) with various configurations: Units alpha or beta. The subunit β-β (S100β) is highly specific in the brain, glial cells and Schwann cells; α-β is in the glial system, and alpha-alpha we can find in striate muscle, heart and kidney.21,24

The synthesis of S100 is regulated by glial cells mainly astrocytes and has the capacity to regulate synaptic plasticity,25,26 most of the protein acts intracytoplasmatic, regulating Ca++, and transcription, axonal growth; when in the extracellular space it participates interacting with R:A:G:E receptors, elevates IL6 and 8,Glutamate; it has the characteristics of a growth factor.6,25,26

In our study the elevated serum levels are considered to be altered metabolism of astrocytes, acute events are most definitely associated to BBB disruption associated to acute insults being associated17,22,27–34 in which even migration from subependimarian areas is stopped as found in autopsies in our department by the author. Further imagenological studies and correlations are needed.

This protein has been considered a biomarker in various scenarios, Cerebral vascular event, Alzheimer, head trauma, damage to BBB, PBP.6,13–42

On the other hand it is known that oxidative stress occurs at neurological level when generation of free radicals exceeds the defense antioxidant mechanisms.

The serum concentrations of different oxidants (MDA, NO, NO) can be and were measured aside, but TAC may resume it all, evidencing the antioxidative status of plasma.6–43

In our study there was an inverse proportional relation between pre-operative TAC and levels of S100β. Besides, the group with neurological perinatal antecedents had a major significative presence in the cyanotic cardiopathies, patients with brain damage probability, chronic hypoxia and cardiac arrest, compared to the group without perinatal antecedents. The inverse relation between TAC and S100β levels can be explained by the tolerance developed by chronic hypoxic patients (cyanotic) through preconditioning in which sublethal hypoxic insults, over express transcription protecting hypoxia factors observed in various tissues, renal, cardiac, cerebral, hepatic.46 This mechanisms involve TIF1 Alpha and HIF (transcription inductible and Hypoxia inductible factors) that regulate genes in response to cellular hypoxic insults, over express transcription protecting hypoxia factors observed in various tissues, renal, cardiac, cerebral, hepatic.46 This mechanisms involve TIF1 Alpha and HIF (transcription inductible and Hypoxia inductible factors) that regulate genes in response to cellular hypoxic insults, over express transcription protecting hypoxia factors observed in various tissues, renal, cardiac, cerebral, hepatic.46 This mechanisms involve TIF1 Alpha and HIF (transcription inductible and Hypoxia inductible factors) that regulate genes in response to cellular hypoxic insults, over express transcription protecting hypoxia factors observed in various tissues, renal, cardiac, cerebral, hepatic.46 This mechanisms involve TIF1 Alpha and HIF (transcription inductible and Hypoxia inductible factors) that regulate genes in response to cellular hypoxic insults, over express transcription protecting hypoxia factors observed in various tissues, renal, cardiac, cerebral, hepatic.46 This mechanisms involve TIF1 Alpha and HIF (transcription inductible and Hypoxia inductible factors) that regulate genes in response to cellular hypoxic insults, over express transcription protecting hypoxia factors observed in various tissues, renal, cardiac, cerebral, hepatic.46 This mechanisms involve TIF1 Alpha and HIF (transcription inductible and Hypoxia inductible factors) that regulate genes in response to cellular hypoxic insults, over express transcription protecting hypoxia factors observed in various tissues, renal, cardiac, cerebral, hepatic.46 This mechanisms involve TIF1 Alpha and HIF (transcription inductible and Hypoxia inductible factors) that regulate genes in response to cellular hypoxic insults, over express transcription protecting hypoxia factors observed in various tissues, renal, cardiac, cerebral, hepatic.46 This mechanisms involve TIF1 Alpha and HIF (transcription inductible and Hypoxia inductible factors) that regulate genes in response to cellular hypoxic insults, over express transcription protecting hypoxia factors observed in various tissues, renal, cardiac, cerebral, hepatic.46

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This mechanisms involve TIF1 Alpha and HIF (transcription inductible and Hypoxia inductible factors) that regulate genes in response to cellular hypoxia and make them more resistant to it (Chronio) by means of anaerobic glycolisis and preserving mitochondrial function.13,47 With low saturations observed in this study it is clear that this mechanisms are called for as well as the explanation of S100β high levels in the preoperative stage. Further specific correlations are needed.

We believe that S100β is a biomarker if we fill the metabolic gaps and basic and clinical gaps surrounding it’s anatomical and clinical interpretation and correlations surrounding hypoxic phenomena both chronic and acute and it’s understanding might be twice important as to develop further research to blockade its effect on R.A.G.E.
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receptors through its TRKT12 link for a true brain protection during congenital heart disease surgery specially in non cyanotic congenital heart disease that do not seem to have this protective adaptive mechanisms of the cytoxic which is our current investigation.

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None.

**Conflict of interest**

The author declares no conflict of interest.

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